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# **APPENDIX A**

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APPLICATION FOR LETTERS PATENT

for

***STREPTOCOCCUS SUI*S VACCINES AND DIAGNOSTIC TESTS**

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## STREPTOCOCCUS SUIIS VACCINES AND DIAGNOSTIC TESTS

### CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This application claims priority to, and is a continuation of, International Application No. PCT/NL99/00460, filed on July 19, 1999, designating the United States of America, the contents of which are incorporated herein by this reference, the PCT International Patent Application itself claiming priority from European Patent Office Application Serial No. 98202465.5 filed July 22, 1998 and European Patent Office Application Serial No. 98202467.1 filed July 22, 1998.

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### TECHNICAL FIELD

[0002] The invention relates to *Streptococcus* infections in pigs, vaccines directed against those infections, tests for diagnosing *Streptococcus* infections and bacterial vaccines. More particularly, the invention relates to vaccines directed against *Streptococcus* infections. TECH CENTER 1600/2900

### BACKGROUND OF THE INVENTION

[0003] *Streptococcus* species, of which a large variety cause infections in domestic animals and man, are often grouped according to Lancefield's groups. Typing according to Lancefield occurs on the basis of serological determinants or antigens that are, among others, present in the capsule of the bacterium, and allows for only an approximate determination. Often, bacteria from different groups show cross-reactivity with each other, while other Streptococci cannot be assigned a group-determinant at all. Within groups, further differentiation is often possible on the basis of serotyping. These serotypes further contribute to the large antigenic variability of Streptococci, a fact that creates an array of difficulties within diagnosis of and vaccination against Streptococcal infections.

[0004] Lancefield group A *Streptococcus species* (Group A *Streptococci* "GAS," *Streptococcus pyogenes*) are common in children, causing nasopharyngeal infections and complications thereof. Among animals, cattle are especially susceptible to GAS infection which can cause mastitis.

[0005] Group A streptococci are the etiologic agents of streptococcal pharyngitis and impetigo, two of the most common bacterial infections in children, as well as a variety of less common, but potentially life-threatening, infections including soft tissue infections, bacteremia, and pneumonia. In addition, GAS are uniquely associated with the post-infectious autoimmune syndromes of acute rheumatic fever and post streptococcal glomerulonephritis.

[0006] Several recent reports suggest that the incidence of both serious infections due to GAS and acute rheumatic fever has increased during the past decade, focusing renewed interest on defining the attributes or virulence factors of the organism that may play a role in the pathogenesis of these diseases.

[0007] GAS produce several surface components and extracellular products that may be important in virulence. The major surface protein, M protein, has been studied in the most detail and has been convincingly shown to play a role in both virulence and immunity. Isolates rich in M protein are able to grow in human blood, a property thought to reflect the capacity of N protein to interfere with phagocytosis, and these isolates tend to be virulent in experimental animals.

[0008] Lancefield group B *Streptococcus* (“GBS”) are most often seen in cattle, causing mastitis; however, human infants are susceptible as well, often with fatal consequences. Group B streptococci (GBS) constitute a major cause of bacterial sepsis and meningitis among human neonates born in the United States and Western Europe and are emerging as significant neonatal pathogens in developing countries as well.

[0009] It is estimated that GBS strains are responsible for 10,000 to 15,000 cases of invasive infection in neonates in the United States alone. Despite advances in early diagnosis and treatment, neonatal sepsis due to GBS continues to carry a mortality rate of 15 to 20%. In addition, survivors of GBS meningitis have 30 to 50% incidence of long-term neurologic sequelae. Over the past two decades, increasing recognition of GBS as an important pathogen for human infants has generated renewed interest in defining the bacterial and host factors important in virulence of GBS and in the immune response to GBS infection.

[0010] Particular attention has focused on the capsular polysaccharide as the predominant surface antigen of the organisms. In a modification of the system originally developed by Rebecca Lancefield, GBS strains are serotyped on the basis of antigenic differences in their capsular

polysaccharides and the presence or absence of serologically defined C proteins. While GBS isolated from non-human sources often lack a serologically detectable capsular, a large majority of strains associated with neonatal infection belong to one of four major capsular serotypes, Ia, Ib, II or III. The capsular polysaccharide forms the outermost layer around the exterior of the bacterial cell, superficial to the cell wall. The capsule is distinct from the cell wall-associated group B carbohydrate. It has been suggested that the presence of sialic acid, in the capsule of bacteria that causes meningitis, is important for allowing these bacteria to breach the blood-brain barrier. Indeed, in *S. agalactiae*, sialic acid has been shown to be critical for the virulence function of the type III capsule. The capsule of *S. suis* serotype is composed of glucose, galactose, N-acetylglucosamine, rhamnose and sialic acid.

[0011] The group B polysaccharide, in contrast to the type-specific capsule, is present on all GBS strains and is the basis for serogrouping the organisms into Lancefield's group B. Early studies by Lancefield and co-workers showed that antibodies raised in rabbits against whole GBS organisms protected mice against challenge with strains of homologous capsular type, demonstrating the central role of the capsular polysaccharide as a protective antigen. Studies in the 1970s by Baker and Kasper demonstrated that cord blood of human infants with type III GBS sepsis uniformly had low or undetectable levels of antibodies directed against the type III capsule, suggesting that a deficiency of anticapsular antibody was a key factor in susceptibility of human neonates to GBS disease.

[0012] Lancefield group C infections, such as those with *S. equi*, *S. zooepidemicus*, *S. dysgalactiae*, and others, are mainly seen in horses, cattle and pigs, but can also cross the species barrier to humans. Lancefield group D (*S. bovis*) infections are found in all mammals and some birds, sometimes resulting in endocarditis or septicemia.

[0013] Lancefield groups E, G, L, P, U and V (*S. porcinus*, *s. canis*, *s. dysgalactiae*) are found in various hosts, causing neonatal infections, nasopharyngeal infections or mastitis.

[0014] Within Lancefield groups R, S and T (and with ungrouped types), *Streptococcus suis* is an important cause of meningitis, septicemia, arthritis and sudden death in young pigs (4, 46). Incidentally, it can also cause meningitis in man (1). *S. suis* strains are usually identified and classified by their morphological, biochemical and serological characteristics (58, 59, 46).

Serological classification is based on the presence of specific antigenic polysaccharides. So far, 35 different serotypes have been described (9, 56, 14). In several European countries, *S. suis* serotype 2 is the most prevalent type isolated from diseased pigs, followed by serotypes 9 and 1. Serological typing of *S. suis* is performed using different types of agglutination tests. In these tests, isolated and biochemically characterized *S. suis* cells are agglutinated with a panel of 35 specific sera. These methods are very laborious and time-consuming.

[0015] Little is known about the pathogenesis of the disease caused by *S. suis*, let alone about its various serotypes such as type 2. Various bacterial components, such as extracellular and cell-membrane associated proteins, fimbriae, hemagglutinins, and hemolysis, have been suggested as virulence factors (9, 10, 11, 15, 16, 47, 49). However, the precise role of these protein components in the pathogenesis of the disease remains unclear (37). It is well known that the polysaccharide capsule of various Streptococci and other gram-positive bacteria plays an important role in pathogenesis (3, 6, 35, 51, 52). The capsule enables these microorganisms to resist phagocytosis and is therefore regarded as an important virulence factor. Recently, a role of the capsule of *S. suis* in the pathogenesis was suggested as well (5). However, the structure, organization and function of the genes responsible for capsule polysaccharide synthesis (“*cps*”) in *S. suis* is unknown. Within *S. suis*, serotype 1 and 2 strains can differ in virulence for pigs (41, 45, 49). Some type 1 and 2 strains are virulent, other strains are not. Because both virulent and non-virulent strains of serotype 1 and 2 strains are fully encapsulated, it may even be that the capsule is not a relevant factor required for virulence.

[0016] Attempts to control *S. suis* infections or disease are still hampered by the lack of knowledge about the epidemiology of the disease and the lack of effective vaccines and sensitive diagnostics. It is well known and generally accepted that the polysaccharide capsule of various Streptococci and other gram-positive bacteria plays an important role in pathogenesis. The capsule enables these microorganisms to resist phagocytosis and is therefore regarded as an important virulence factor.

[0017] Compared to encapsulated *S. suis* strains, non-encapsulated *S. suis* strains are phagocytosed by murine polymorphonuclear leucocytes to a greater degree. Moreover, an increase in thickness of capsule was noted for *in vivo* grown virulent strains while no increase was observed for

avirulent strains. Therefore, these data again demonstrate the role of the capsule in the pathogenesis for *S. suis* as well.

[0018] Ungrouped *Streptococcus species*, such as *S. mutans*, causing caries in humans, *S. uberis*, causing mastitis in cattle, and *S. pneumoniae*, causing major infections in humans, and *Enterococcus faecilalis* and *E. faecium*, further contribute to the large group of Streptococci.

[0019] *Streptococcus pneumoniae* (the pneumococcus) is a human pathogen causing invasive diseases, such as pneumonia, bacteremia, and meningitis. Despite the availability of antibiotics, pneumococcal infections remain common and can still be fatal, especially in high-risk groups, such as young children and elderly people. Particularly in developing countries, many children under the age of five years die each year from pneumococcal pneumonia. *S. pneumoniae* is also the leading cause of otitis media and sinusitis. These infections are less serious, but nevertheless incur substantial medical costs, especially when leading to complications, such as permanent deafness. The normal ecological niche of the pneumococcus is the nasopharynx of man. The entire human population is colonized by the pneumococcus at one time or another, and at a given time, up to 60% of individuals may be carriers. Nasopharyngeal carriage of pneumococci by man is often accompanied by the development of protection against infection by the same serotype. Most infections do not occur after prolonged carriage but follow exposure to recently acquired strains. Many bacteria contain surface polysaccharides that act as a protective layer against the environment. Surface polysaccharides of pathogenic bacteria usually make the bacteria resistant to the defense mechanisms of the host, for example, the lytic action of serum or phagocytosis. In this respect, the serotype-specific capsular polysaccharide (“CP”) of *Streptococcus pneumoniae*, is an important virulence factor. Unencapsulated strains are avirulent, and antibodies directed against the CP are protective. Protection is serotype specific; each serotype has its own, specific CP structure. Ninety different capsular serotypes have been identified. Currently, CPs of 23 serotypes are included in a vaccine.

[0020] Vaccines directed against *Streptococcus* infections typically aim to utilize an immune response directed against the polysaccharide capsule of the various *Streptococcus species*, especially since the capsule is considered a primary virulence factor for these bacteria. During

infection, the capsule provides resistance against phagocytosis and thus protects the bacteria from the immune system of the host, and from elimination by macrophages and neutrophils.

[0021] The capsule particularly confers the bacterium resistance to complement-mediated opsonophagocytosis. In addition, some bacteria express capsular polysaccharides (CPs) that mimic host molecules, thereby avoiding the immune system of the host. Also, even when the bacteria have been phagocytosed, intracellular killing is hampered by the presence of a capsule.

[0022] It is generally thought that the bacterium will get recognized by the immune system through the anticapsular-antibodies or serum-factors bound to its capsule and will, through opsonization, get phagocytosed and killed only when the host has antibodies or other serum factors directed against capsule antigens.

[0023] However, these antibodies are serotype-specific, and will often only confer protection against only one of the many serotypes known within a group of *Streptococci*.

[0024] For example, current commercially available *S. suis* vaccines, which are generally based on whole-cell-bacterial preparations, or on capsule-enriched fractions of *S. suis*, confer only limited protection against heterologous strains. Also, the current pneumococcal vaccine that was licensed in the United States in 1983, consists of purified CPs of 23 pneumococcal serotypes whereas at least 90 CP types exist.

[0025] The composition of this pneumococcal vaccine was based, on the frequency of the occurrence of disease isolates in the US and cross-reactivity between various serotypes. Although this vaccine protects healthy adults against infections caused by serotypes included in the vaccine, it fails to raise a protective immune response in infants younger than 18 months and it is less effective in elderly people. In addition, the vaccine confers only limited protection in patients with immunodeficiencies and hematology malignancies.

[0026] Thus, improved vaccines are needed against *Streptococcus* infections. Much attention is directed toward producing CP vaccines by producing the relevant polysaccharides via chemical or recombinant means. However, chemical synthesis of polysaccharides is costly, and capsular polysaccharide synthesis by recombinant means necessitates knowledge about the relevant genes, which is not always available, and needs to be determined for every relevant serotype.



## DISCLOSURE OF THE INVENTION

[0027] The invention provides an isolated or recombinant nucleic acid encoding a capsular (*cps*) gene cluster of *Streptococcus suis*. Biosynthesis of capsule polysaccharides has generally been studied in a number of Gram-positive and Gram-negative bacteria (32). In Gram-negative bacteria, but also in a number of Gram-positive bacteria, genes which are involved in the biosynthesis of polysaccharides are clustered at a single locus.

[0028] *Streptococcus suis* capsular genes, as provided by the invention, show a common genetic organization involving three distinct regions. The central region is serotype specific and encodes enzymes responsible for the synthesis and polymerization of the polysaccharides. The central region is flanked by two regions conserved in *Streptococcus suis* which encode proteins for common functions, such as transport of the polysaccharide across the cellular membrane. However, between species, only low homologies exist, hampering easy comparison and detection of seemingly similar genes. Knowing the nucleic acid encoding the flanking regions allows type-specific determination of nucleic acid of the central region of *Streptococcus suis* serotypes as, for example, described herein.

[0029] The invention provides an isolated or recombinant nucleic acid encoding a capsular gene cluster of *Streptococcus suis* or a gene or gene fragment derived thereof. Such a nucleic acid is, for example, provided by hybridizing chromosomal DNA derived from any one of the *Streptococcus suis* serotypes to a nucleic acid encoding a gene derived from a *Streptococcus suis* serotype 1, 2 or 9 capsular gene cluster, as provided by the invention (*see*, for example, Tables 4 and 5) and cloning of (type-specific) genes as, for example, described herein. At least 14 open reading frames are identified. Most of the genes belong to a single transcriptional unit, identifying a coordinate control of these genes. The genes, and the enzymes and proteins they encode, act in concert to provide the capsule with the relevant polysaccharides.

[0030] The invention provides *cps* genes and proteins encoded thereof involved in regulation (CpsA), chain length determination (CpsB, C), export (CpsC) and biosynthesis (CpsE, F, G, H, J, K). Although, at first glance, the overall organization seemed to be similar to that of the *cps* and *eps* gene clusters of a number of Gram-positive bacteria (19, 32, 42), overall homologies are low

(see, table 3). The region involved in biosynthesis is located at the center of the gene cluster and is flanked by two regions containing genes with more common functions.

**[0031]** The invention provides an isolated or recombinant nucleic acid encoding a capsular gene cluster of *Streptococcus suis* serotype 2, or a gene or gene fragment derived thereof, preferably as identified in FIG. 3. Genes in this gene cluster are involved in polysaccharide biosynthesis of capsular components and antigens. For a further description of such genes see, for example, Table 2. For example, a *cpsA* gene is provided functionally encoding regulation of capsular polysaccharide synthesis, whereas *cpsB* and *cpsC* are functionally involved in chain-in-chain length determination. Other genes, such as *cpsD*, *E*, *F*, *G*, *H*, *I*, *J*, *K* and related genes, are involved in polysaccharide synthesis, functioning, for example, as glucosyl- or glycosyltransferase. The *cpsF*, *G*, *H*, *I*, *J* genes encode more type-specific proteins than the flanking genes which are found more-or-less conserved throughout the species and can serve as a base for selection of primers or probes in PCR-amplification or cross-hybridization experiments for subsequent cloning.

**[0032]** The invention further provides an isolated or recombinant nucleic acid encoding a capsular gene cluster of *Streptococcus suis* serotype 1 or a gene or gene fragment derived thereof, preferably as identified in FIG. 4.

**[0033]** In addition, the invention provides an isolated or recombinant nucleic acid encoding a capsular gene cluster of *Streptococcus suis* serotype 9 or a gene or gene fragment derived thereof, preferably as identified in FIG. 5.

**[0034]** Furthermore, the invention provides, for example, a fragment of the *cps* locus, or parts thereof, involved in the capsular polysaccharide biosynthesis of *S. suis* exemplified herein for serotypes 1, 2 or 9, and allows easy identification or detection of related fragments derived of other serotypes of *S. suis*.

**[0035]** The invention provides a nucleic acid probe or primer derived from a nucleic acid according to the invention allowing species or serotype-specific detection of *Streptococcus suis*. Such a probe or primer (used interchangeably herein) is, for example, a DNA, RNA or PNA (peptide nucleic acid) probe hybridizing with capsular nucleic acid as provided by the invention. Species-specific detection is provided preferably by selecting a probe or primer sequence from a species-specific region (e.g. flanking region) whereas serotype-specific detection is provided

preferably by selecting a probe or primer sequence from a type-specific region (*e.g.* central region) of a capsular gene cluster as provided by the invention. Such a probe or primer can be used in a further unmodified form, for example, in cross-hybridization or polymerase-chain reaction (PCR) experiments as, for example described in the experimental part herein. The invention provides the isolation and molecular characterization of additional type-specific *cps* genes of *S. suis* types 1 and 9.

In addition, we describe the genetic diversity of the *cps* loci of serotypes 1, 2 and 9 among the 35 *S. suis* serotypes known. Type-specific probes are identified. Also, a type-specific PCR, for example, for serotype 9, is provided, being a rapid, reliable and sensitive assay used directly on nasal or tonsillar swabs or other samples of infected or carrier animals.

[0036] The invention also provides a probe or primer according to the invention with at least one reporter molecule. Examples of reporter molecules are manifold and known in the art; for example, a reporter molecule can include additional nucleic acid provided with a specific sequence (*e.g.* oligo-dT) hybridizing to a corresponding sequence in which hybridization can easily be detected, for example, because it has been immobilized to a solid support.

[0037] Yet other reporter molecules include chromophores, *e.g.* fluorochromes for visual detection, for example, by light microscopy or fluorescent *in situ* hybridization (“FISH”) techniques, or include an enzyme such as horseradish peroxidase for enzymatic detection, *e.g.* in enzyme-linked assays (“EIA”). Yet other reporter molecules include radioactive compounds for detection in radiation-based assays.

[0038] In a preferred embodiment of the invention, at least one probe or primer according to the invention is provided (labeled) with a reporter molecule and a quencher molecule, together with an unlabeled probe or primer in a PCR-based test allowing rapid detection of specific hybridization.

[0039] The invention further provides a diagnostic test or test kit including a probe or primer as provided by the invention. Such a test or test kit is, for example, a cross-hybridization test or PCR-based test advantageously used in rapid detection and/or serotyping of *Streptococcus suis*.

[0040] The invention further provides a protein or fragment thereof encoded by a nucleic acid according to the invention. Examples of such a protein or fragment are proteins described in Table 2. For example, a *cpsA* protein is provided that functionally encodes regulation of capsular

polysaccharide synthesis, whereas *cpsB* and *cpsC* are functionally involved in chain-in-chain length determination. Other proteins or functional fragments thereof, as provided by the invention, such as *cpsD*, *E*, *F*, *G*, *H*, *I*, *J*, *K* and related proteins, are involved in polysaccharide biosynthesis, functioning, for example, as glucosyl- or glycosyltransferase in polysaccharide biosynthesis of *Streptococcus suis* capsular antigen.

[0041] The invention also provides a method of producing a *Streptococcus suis* capsular antigen including using a protein or functional fragment thereof as provided by the invention, and provides therewith a *Streptococcus suis* capsular antigen obtainable by such a method.

[0042] A comparison of the predicted amino acid sequences of the *cps2* genes with sequences found in the databases allowed the assignment of functions to the open reading frames. The central region contains the type-specific glycosyltransferases and the putative polysaccharide polymerase. This region is flanked by two regions encoding for proteins with common functions, such as regulation and transport of polysaccharide across the membrane. Biosynthesis of *Streptococcus* capsular polysaccharide antigen using a protein or functional fragment thereof is advantageously used in chemo-enzymatic synthesis and the development of vaccines which offer protection against serotype-specific Streptococcal disease, and is also advantageously used in the synthesis and development of multivalent vaccines against Streptococcal infections. Such vaccines elicit anticapsular antibodies which confer protection.

[0043] Furthermore, the invention provides an acapsular *Streptococcus* mutant for use in a vaccine, a vaccine strain derived thereof and a vaccine derived thereof. Surprisingly, and against the grain of common doctrine, the invention provides use of a *Streptococcus* mutant deficient in capsular expression in a vaccine.

[0044] Acapsular *Streptococcus* mutants have long been known in the art and can be found in nature. Griffith (*J. Hyg.* 27:113-159, 1928) demonstrated that pneumococci could be transformed from one type to another. If he injected live rough (acapsular or unencapsulated) type 2 pneumococci into mice, the mice would survive. If, however, he injected the same dose of live rough type 2 mixed with heat-killed smooth (encapsulated) type 1 into a mouse, the mouse would die, and, from the blood, he could isolate live smooth type 1 pneumococci. At that time, the significance of this transforming principle was not understood. However, understanding came when

it was shown that DNA constituted the genetic material responsible for phenotypic changes during transformation.

[0045] *Streptococcus* mutants deficient in capsular expression are found in several forms. Some are fully deficient and have no capsule at all, others form a deficient capsule, characterized by a mutation in a capsular gene cluster. Deficiency can, for instance, include capsular formation wherein the organization of the capsular material has been rearranged as, for example, demonstrable by electron microscopy. Yet others have a nearly fully developed capsule which is only deficient in a particular sugar component.

[0046] Now, after much advance of biotechnology and despite the fact that little is still known about the exact localization and sequence of genes involved in capsular synthesis in *Streptococci*, it is possible to create mutants of *Streptococci*, for example, by homologous recombination or transposon mutagenesis, which has, for example, been done for GAS (Wessels et al., *PNAS* 88:8317-8321, 1991), for GBS (Wessels et al., *PNAS* 86: 8983-8987, 1989), for *S. suis* (Smith, ID-DLO Annual report 1996, page 18-19; Charland et al., *Microbiol.* 144:325-332, 1998) and *S. pneumoniae* (Kolkman et al., *J. Bact.* 178:3736-3741, 1996). Such recombinant derived mutants, or isogenic mutants, can easily be compared with the wild-type strains from which they have been derived.

[0047] In a preferred embodiment, the invention provides use of a recombinant-derived *Streptococcus* mutant deficient in capsular expression in a vaccine. Recombinant techniques useful in producing such mutants are, for example, homologous recombination, transposon mutagenesis, and others, wherein deletions, insertions or (point) mutations are introduced in the genome. Advantages of using recombinant techniques include the stability of the obtained mutants (especially with homologous recombination and double cross-over techniques), and the knowledge about the exact site of the deletion, mutation or insertion.

[0048] In another embodiment, the invention provides a stable mutant deficient in capsular expression obtained, for example, through homologous recombination or cross-over integration events. Examples of such a mutant can be found herein, such as mutants 10cpsB or 10cpsEF are stable mutants as provided by the invention.

[0049] The invention also provides a *Streptococcus* vaccine strain and vaccine that has been derived from a *Streptococcus* mutant deficient in capsular expression. In general, the strain or vaccine is applicable within the whole range of Streptococcal infections, including animals or man or with zoonotic infections. It is, of course, now possible to first select a common vaccine strain and derive a *Streptococcus* mutant deficient in capsular expression thereof for the selection of a vaccine strain and use in a vaccine according to the invention.

[0050] In a preferred embodiment, the invention provides use of a *Streptococcus* mutant deficient in capsular expression in a vaccine wherein the *Streptococcus* mutant is selected from the group composed of *Streptococcus* group A, *Streptococcus* group B, *Streptococcus suis* and *Streptococcus pneumoniae*. Herewith the invention provides vaccine strains and vaccines for use with these notoriously heterologous Streptococci, of which a multitude of serotypes exist. With a vaccine, as provided by the invention, that is derived from a specific *Streptococcus* mutant that is deficient in capsular expression, the difficulties relating to lack of heterologous protection can be circumvented since these mutants do not rely on capsular antigens, per se, to induce protection.

[0051] In a preferred embodiment, the vaccine strain is selected for its ability to survive, or even replicate, in an immune-competent host or host cells and thus can persist for a certain period, varying from 1-2 days to more than one or two weeks, in a host, despite its deficient character.

[0052] Although an immunodeficient host will support replication of a wide range of bacteria that are deficient in one or more virulence factors, in general, it is considered a characteristic of pathogenicity of Streptococci that they can survive for certain periods or replicate in a normal host or host cells such as macrophages. For example, Williams and Blakemore (*Neuropath. Appl. Neurobiol.* 16, 345-356, 1990; *Neuropath. Appl. Neurobiol.* 16, 377-392, 1990; *J. Infect. Dis.* 162, 474-481, 1990) show that both polymorphonuclear cells and macrophage cells are capable of phagocytosing pathogenic *S. suis* in pigs lacking anti-*S. suis* antibodies; only pathogenic bacteria could survive and multiply inside macrophages and the pig.

[0053] In a preferred embodiment, the invention, however, provides a deficient or avirulent mutant or vaccine strain which is capable of surviving at least 4-5 days, preferably at least 8-10 days in the host, thereby allowing the development of a solid immune response to subsequent *Streptococcus* infection,

[0054] Due to its persistent but avirulent character, a *Streptococcus* mutant or vaccine strain, as provided by the invention, is well suited to generate specific and/or long-lasting immune responses against Streptococcal antigens. Moreover, possible specific immune responses of the host directed against a capsule are relatively irrelevant because a vaccine strain, as provided by the invention, is typically not recognized by such antibodies.

[0055] In addition, the invention provides a *Streptococcus* vaccine strain, according to the invention, which strain includes a mutant capable of expressing a *Streptococcus* virulence factor or antigenic determinant.

[0056] In a preferred embodiment, the invention provides a *Streptococcus* vaccine strain, according to the invention, which includes a mutant capable of expressing a *Streptococcus* virulence factor wherein the virulence factor or antigenic determinant is selected from a group of cellular components, such as muramidase-released protein ("MRP"), extracellular factor ("EF"), and cell-membrane associated proteins, 60kDA heat shock protein, pneumococcal surface protein A (Psp A), pneumolysin, C protein, protein M, fimbriae, hemagglutinins and hemolysis or components functionally related thereto.

[0057] In a preferred embodiment, the invention provides a *Streptococcus* vaccine strain including a mutant capable of over-expressing the virulence factor. In this way, the invention provides a vaccine strain for incorporation in a vaccine which specifically causes a host immune response directed against antigenically important determinants of virulence (listed above), thereby providing specific protection against the determinants. Over-expression can, for example, be achieved by cloning the gene involved behind a strong promoter, which is, for example, constitutionally expressed in a multicopy system, either in a plasmid or via integration in a genome.

[0058] In yet another embodiment, the invention provides a *Streptococcus* vaccine strain, according to the invention, including a mutant capable of expressing a non-*Streptococcus* protein. Such a vector-*Streptococcus* vaccine strain allows, when used in a vaccine, protection against pathogens other than *Streptococcus*.

[0059] Due to its persistent but avirulent character, a *Streptococcus* vaccine strain or mutant as provided by the invention is well suited to generate specific and long-lasting immune responses, not only against Streptococcal antigens, but also against other antigens expressed by the

strain. Specifically, antigens derived from another pathogen are now expressed without the detrimental effects of the antigen or pathogen which would otherwise have harmed the host.

[0060] An example of such a vector is a *Streptococcus* vaccine strain or mutant wherein the antigen is derived from a pathogen, such as *Actinobacillus pleuropneumonia*, *Mycoplasma*, *Bordetella*, *pasteurella*, *E. coli*, *Salmonella*, *campylobacter*, *Serpulina* and others.

[0061] The invention also provides a vaccine including a *Streptococcus* vaccine strain or mutant according to the invention and a pharmaceutically acceptable carrier or adjuvant. Carriers or adjuvants are well known in the art; examples are phosphate buffered saline, physiological salt solutions, (double-) oil-in-water emulsions, aluminumhydroxide, Specol, block- or co-polymers, and others.

[0062] A vaccine according to the invention can include a vaccine strain either in a killed or live form. For example, a killed vaccine including a strain having (over)expressed a Streptococcal or heterologous antigen or virulence factor is very well suited for eliciting an immune response. In a preferred embodiment, the invention provides a vaccine wherein the strain is live, due to its persistent but avirulent character; a *Streptococcus* vaccine strain, as provided by the invention, is well suited to generate specific and long-lasting immune responses.

[0063] The invention also provides a method for controlling or eradicating a Streptococcal disease in a population comprising vaccinating subjects in the population with a vaccine according to the invention.

[0064] In a preferred embodiment, a method for controlling or eradicating a Streptococcal disease is provided including testing a sample, such as a blood sample, or nasal or throat swab, feces, urine, or other samples such as can be sampled at or after slaughter, collected from at least-one subject, such as an infant or a pig, in a population partly or wholly vaccinated with a vaccine according to the invention for the presence of encapsulated Streptococcal strains or mutants. Since a vaccine strain or mutant according to the invention is not pathogenic, and can be distinguished from wild-type strains by capsular expression, the detection of (fully) encapsulated Streptococcal strains indicates that wild-type infections are still present. Such wild-type infected subjects can then be isolated from the remainder of the population until the infection has passed. With domestic animals, such as pigs, it is even possible to remove the infected subject from the population as a whole by



culling. Detection of wild-type strains can be achieved via traditional culturing techniques, or by rapid detection techniques such as PCR detection.

**[0065]** In yet another embodiment, the invention provides a method for controlling or eradicating a Streptococcal disease including testing a sample collected from at least one subject in a population partly or wholly vaccinated with a vaccine according to the invention for the presence of capsule-specific antibodies directed against Streptococcal strains. Capsule-specific antibodies can be detected with classical techniques known in the art, such as used for Lancefield's group typing or serotyping.

**[0066]** A preferred embodiment for controlling or eradicating a Streptococcal disease in a population includes vaccinating subjects in the population with a vaccine according to the invention and testing a sample collected from at least one subject in the population for the presence of encapsulated Streptococcal strains and/or for the presence of capsule-specific antibodies directed against Streptococcal strains.

**[0067]** For example, a method is provided wherein the Streptococcal disease is caused by *Streptococcus suis*.

**[0068]** The invention also provides a diagnostic assay for testing a sample for use in a method according to the invention including at least one means for the detection of encapsulated Streptococcal strains and/or for the detection of capsule-specific antibodies directed against Streptococcal strains.

**[0069]** The invention further provides a vaccine including an antigen according to the invention and a suitable carrier or adjuvant. The immunogenicity of a capsular antigen provided by the invention is, for example, increased by linking to a carrier (such as a carrier protein), allowing the recruitment of T-cell help in developing an immune response.

**[0070]** The invention further provides a recombinant microorganism provided with at least a part of a capsular gene cluster derived from *Streptococcus suis*. The invention provides for example a lactic acid bacterium provided with at least a part of a capsular gene cluster derived from *Streptococcus suis*. Various food-grade lactic acid bacteria (*Lactococcus lactis*, *Lactobacillus casei*, *Lactobacillus plantarium* and *Streptococcus gordonii*) have been used as delivery systems for mucosal immunization. It has now been shown that oral (or mucosal) administration of recombinant

*L. lactis*, *Lactobacillus*, and *Streptococcus gordonii* can elicit local IgA and/or IgG antibody responses to an expressed antigen. The use of oral routes for immunization against infective diseases is desirable because oral vaccines are easier to administer and have higher compliance rates, and because mucosal surfaces are the portals of entry for many pathogenic microbial agents. It is within the skill of the artisan to provide such microorganisms with (additional) genes.

[0071] The invention further provides a recombinant *Streptococcus suis* mutant provided with a modified capsular gene cluster. It is within the skill of the artisan to swap genes within a species. In a preferred embodiment, an avirulent *Streptococcus suis* mutant is selected to be provided with at least a part of a modified capsular gene cluster according to the invention.

[0072] The invention further provides a vaccine including a microorganism or a mutant provided by the invention. An advantage of such a vaccine over currently used vaccines is that they include accurately defined microorganisms and well-characterized antigens, allowing accurate determination of immune responses against various antigens of choice.

[0073] The invention is further explained in the experimental part of this description without limiting the invention thereto.

## DESCRIPTION OF THE DRAWINGS

[0074] FIG. 1 illustrates the organization of the *cps2* gene cluster of *S. suis* type 2.

[0075] (A) Genetic map of the *cps2* gene cluster. The shadowed arrows represent potential ORFs. Interrupted ORFs indicate the presence of stop codons or frame-shift mutations. Gene designations are indicated below the ORFs. The closed arrows indicate the position of the potential promoter sequences. | indicates the position of the potential transcription regulator sequence. ||| indicates the position of the 100-bp repeated sequence.

[0076] (B) Physical map of the *cps2* locus. Restriction sites are as follows: A: *AluI*; C: *ClaI*; E: *EcoRI*; H: *HindIII*; K: *KpnI*; M: *MluI*; N: *NsiI*; P: *PstI*; S: *SnaBI*; Sa: *SacI*; X: *XbaI*.

[0077] (C) The DNA fragments cloned in the various plasmids.

[0078] FIG. 2 illustrates ethidium bromide stained agarose gel showing PCR products obtained with chromosomal DNA of *S. suis* strains belonging to the serotypes 1,2, 1/2, 9 and 14 and *cps2J*, *cps1I*, and *cps9H* primer sets as described herein.

[0079] (A) *cpsII* primers; (B) *cps2J* primers and (C) *cps9H* primers.

[0080] Lanes 1-3: serotype 1 strains; lanes 4-6: serotype 2 strains; lanes 7-9: serotype 1/2 strains; lanes 10-12: serotype 9 strains and lanes 13-15: serotype 14 strains.

[0081] (B) Ethidium bromide stained agarose gel showing PCR products obtained with tonsillar swabs collected from pigs carrying *S. suis* type 2, type 1 or type 9 strains and *cps2J*, *cpsII* and *cpsH* primer sets as described in Materials and Methods. Bacterial DNA suitable for PCR was prepared by using the multiscreen methods as described previously (20).

[0082] (C) *cpsII* primers. (B) *cps2J* primers and (C) *cps9H* primers.

[0083] Lanes 1-3: PCR products obtained with tonsillar swabs collected from pigs carrying *S. suis* type 1 strains; lanes 4-6: PCR products obtained with tonsillar swabs collected from pigs carrying *S. suis* type 2 strains; lanes 7-9: PCR products obtained with tonsillar swabs collected from pigs carrying *S. suis* type 9 strains; lanes 10-12: PCR products obtained with chromosomal DNA from serotype 9, 2 and 1 strains respectively; lane 13: negative control, no DNA present.

[0084] FIG. 3 illustrates the CPS2 nucleotide sequences and corresponding amino acid sequences from the open reading frames.

[0085] FIG. 4 illustrates the CPS1 nucleotide sequences and corresponding amino acid sequences from the open reading frames.

[0086] FIG. 5 illustrates the CPS9 nucleotide sequences and corresponding amino acid sequences from the open reading frames.

[0087] FIG. 6 illustrates the CPS7 nucleotide sequences and corresponding amino acid sequences from the open reading frames.

[0088] FIG. 7 illustrates alignment of the N-terminal parts of Cps2J and Cps2K.

[0089] Identical amino acids are marked by bars. The amino acids shown in bold are also conserved in Cps14I, Cps24J of *S. pneumoniae* and several other glycosyltransferases (19). The aspartate residues marked by asterisks are strongly conserved.

[0090] FIG. 8 illustrates transmission electron micrographs of thin sections of various *S. suis* strains.

[0091] (A) wild-type strain 10;

[0092] (B) mutant strain 10cpsB;

[0093] (C) mutant strain 10cpsEF.

[0094] Bar = 100 nm

[0095] **FIG. 9** illustrates the kinetics of phagocytosis of wild-type and mutant *S. suis* strains.

[0096] (A) Kinetics of phagocytosis of wild-type and mutant *S. suis* strains by porcine alveolar macrophages. Phagocytosis was determined as described herein. The Y-axis represents the number of CFU per milliliter in the supernatant fluids as determined by plate counting, the X-axis represents time in minutes.

[0097] □ wild-type strain 10;

[0098] ○ mutant strain 10cpsB;

[0099] △ mutant strain 10cpsEF.

[00100] (B) Kinetics of intracellular killing of wild-type and mutant *S. suis* strains by porcine AM. The intracellular killing was determined as described herein. The Y-axis represents the number of CFU per ml in the supernatant fluids after lysis of the macrophages as determined by plate counting, the X-axis represents time in minutes.

[00101] □ wild-type strain 10;

[00102] ○ mutant strain 10cpsB;

[00103] △ mutant strain 10cpsEF.

[00104] **FIG. 10** illustrates the nucleotide sequence alignment of the highly conserved 100-bp repeated element.

[00105] 1) 100-bp repeat between cps2G and cps2H

[00106] 2) 100-bp repeat within “cps2M”

[00107] 3) 100-bp repeat between cps20 and cps2P

[00108] **FIG. 11** illustrates the cps2, cps9 and cps7 gene clusters of *S. suis* serotypes 2, 9 and 7.

[00109] (A) Genetic organization of the cps2 gene cluster [84]. The large arrows represent potential ORFs. Gene designations are indicated below the ORFs. Identically filled arrows represent ORFs which showed homology. The small closed arrows indicate the position of the potential promoter sequences. | indicates the position of the potential transcription regulator sequence.

[00110] (B) Physical map and genetic organization of the *cps9* gene cluster [15]. Restriction sites are as follows: B: *Bam*HI; P: *Pst*I; H: *Hind*III; X: *Xba*I. The DNA fragments cloned in the various plasmids are indicated. The open arrows represent potential ORFs.

[00111] (C) Physical map and genetic organization of the *cps7* gene cluster. Restriction sites are as follows: C: *Clal*; P: *Pst*I; Sc: *Sca*I. The DNA fragments cloned in the various plasmids are indicated. The open arrows represent potential ORFs.

[00112] **FIG. 12** illustrates ethidium bromide stained agarose gel showing PCR products.

[00113] (A) Ethidium bromide stained agarose gel showing PCR products obtained with chromosomal DNA of *S. suis* strains belonging to the serotypes 1, 2, 9 and 7 and the *cps7H* primer set. Strain designations are indicated above the lanes. C: negative control, no DNA present. M: molecular size marker (lambda digested with *Eco*RI and *Hind*III).

[00114] (B) Ethidium bromide stained agarose gel showing PCR products obtained with serotype 7 strains collected in different countries and from different organs. Bacterial DNA suitable for PCR was prepared by using the multiscreen method as described herein [89]. Strain designations are indicated above the lanes. M: molecular size marker (lambda digested with *Eco*RI and *Hind*III).

## DETAILED DESCRIPTION OF THE INVENTION

### Experimental part

### MATERIAL AND METHODS

#### Bacterial strains and growth conditions.

[00115] The bacterial strains and plasmids used in this study are listed in Table 1. *S. suis* strains were grown in Todd-Hewitt broth (code CM189, Oxoid), and plated on Columbia agar blood base (code CM331, Oxoid) containing 6% (v/v) horse blood. *E. coli* strains were grown in Luria broth (28) and plated on Luria broth containing 1.5% (w/v) agar. If required, antibiotics were added to the plates at the following concentrations: spectinomycin: 100 µg/ml for *S. suis* and 50 µg/ml for *E. coli* and ampicillin, 50 µg/ml.

[00116] **Serotyping.** The *S. suis* strains were serotyped by the slide agglutination test with serotype-specific antibodies (44).

**[00117] DNA techniques.** Routine DNA manipulations were performed as described by Sambrook et al. (36).

**[00118] Alkaline phosphatase activity.** To screen for PhoA fusions in *E. coli*, plasmid libraries were constructed. Therefore, chromosomal DNA of *S. suis* type 2 was digested with *AluI*. The 300-500-bp fragments were ligated to *SmaI*-digested pPHOS2. Ligation mixtures were transformed to the PhoA<sup>-</sup> *E. coli* strain CC118. Transformants were plated on LB media supplemented with 5- Bromo-4-chloro-3-indolylfosfaat (BCIP, 50 µg/ml, Boehringer, Mannheim, Germany). Blue colonies were purified on fresh LB/BCIP plates to verify the blue phenotype.

**[00119] DNA sequence analysis.** DNA sequences were determined on a 373A DNA Sequencing System (Applied Biosystems, Warrington, GB). Samples were prepared by using an ABI/PRISM dye terminator cycle sequencing ready reaction kit (Applied Biosystems). Sequencing data were assembled and analyzed using the MacMollyTetra program. Custom-made sequencing primers were purchased from Life Technologies. Hydrophobic stretches within proteins were predicted by the method of Klein et al. (17). The BLAST program available on Netscape Navigator™ was used to search for protein sequences related to the deduced amino acid sequences.

**[00120] Construction of gene-specific knock-out mutants of *S. suis*.** To construct the mutant strains 10cpsB and 10cpsEF, we electrotransformed the pathogenic serotype 2 strain 10 (45, 49) of *S. suis* with pCPS11 and pCPS28 respectively. In these plasmids, the *cpsB* and *cpsEF* genes were disturbed by the insertion of a spectinomycin-resistance gene. To create pCPS11, the internal 400 bp *PstI*-*BamHI* fragment of the *cpsB* gene in pCPS7 was replaced by the Spc<sup>R</sup> gene. For this purpose, pCPS7 was digested with *PstI* and *BamHI* and ligated to the 1,200-bp *PstI*-*BamHI* fragment, containing the Spc<sup>R</sup> gene, from pIC-spc. To construct pCPS28, we have used pIC20R. In this plasmid we inserted the *KpnI*-*SalI* fragment from pCPS17 (resulting in pCPS25) and the *XbaI*-*ClaI* fragment from pCPS20 (resulting in pCPS27). pCPS27 was digested with *PstI* and *XhoI* and ligated to the 1,200-bp *PstI*-*XhoI* fragment, containing the Spc<sup>R</sup> gene of pIC-spc. The electrotransformation to *S. suis* was carried out as described before (38).

**[00121] Southern blotting and hybridization.** Chromosomal DNA was isolated as described by Sambrook et al. (36). DNA fragments were separated on 0.8% agarose gels and transferred to Zeta-Probe GT membranes (Bio-Rad) as described by Sambrook et al. (36). DNA

probes were labeled with [(<sup>32</sup>P)dCTP (3000 Ci mmol<sup>-1</sup> Amersham) by use of a random primed labeling kit (Boehringer). The DNA on the blots was hybridized at 65°C with appropriate DNA probes as recommended by the supplier of the Zeta-Probe membranes. After hybridization, the membranes were washed twice with a solution of 40 mM sodium phosphate, pH 7.2, 1 mM EDTA, 5% SDS for 30 min at 65°C and twice with a solution of 40 mM sodium phosphate, pH 7.2, 1 mM EDTA, 1% SDS for 30 min at 65°C.

**[00122] PCR.** The primers used in the *cps2J* PCR correspond to the positions 13791-13813 and 14465-14443 in the *S. suis cps2* locus. The sequences were: 5'-CAAACGCAAGGAATTACGGTATC-3' (SEQ ID NO:1) and 5'-GAGTATCTAAAGAATGCCTATTG-3' (SEQ ID NO:2). The primers used for the *cps1I* PCR correspond to the positions 4398-4417 and 4839-4821 in the *S. suis cps1* sequence. The sequences were: 5'-GGCGGTCTAGCAGATGCTCG-3' (SEQ ID NO:3) and 5'-GCGAACTGTTAGCAATGAC-3' (SEQ ID NO:4). The primers used in the *cps9H* PCR correspond to the positions 4406-4126 and 4494-4475 in the *S. suis cps9* sequence. The sequences were: 5'-GGCTACATATAATGGAAGCCC3' (SEQ ID NO:5) and 5'-CGGAAGTATCTGGGCTACTG-3' (SEQ ID NO:6).

**[00123] Construction of gene-specific knock-out mutants of *S. suis*.** To construct the mutant strains 10cpsB and 10cpsEF, we electrotransformed the pathogenic serotype 2 strain 10 of *S. suis* with pCPS11 and pCPS28 respectively. In these plasmids, the *cpsB* and *cpsEF* genes were disturbed by the insertion of a spectinomycin-resistance gene. To create pCPS11, the internal 400 bp *PstI*-*BamHI* fragment of the *cpsB* gene in pCPS7 was replaced by the Spc<sup>R</sup> gene. For this purpose, pCPS7 was digested with *PstI* and *BamHI* and ligated to the 1,200-bp *PstI*-*BamHI* fragment, containing the Spc<sup>R</sup> gene, from pIC-spc. To construct pCPS28, we have used pIC20R. In this plasmid, we inserted the *KpnI*-*SalI* fragment from pCPS17 (resulting in pCPS25) and the *XbaI*-*ClaI* fragment from pCPS20 (resulting in pCPS27). pCPS27 was digested with *PstI* and *XhoI* and ligated to the 1,200-bp *PstI*-*XhoI* fragment, containing the spc<sup>R</sup> gene of pIC-Spc. The electrotransformation to *S. suis* was carried out as described before (38).

**[00124] Phagocytosis assay.** Phagocytosis assays were performed as described by Leij et al. (23). Briefly, to opsonize the cells, 10<sup>7</sup> *S. suis* cells were incubated with 6% SPF-pig serum for 30

min at 37°C in a head-over-head rotor at 6 rpm.  $10^7$  AM and  $10^7$  opsonized *S. suis* cells were combined and incubated at 37°C under continuous rotation at 6 rpm. At 0, 30, 60 and 90 min, 1-ml samples were collected and mixed with 4 ml of ice-cold EMEM to stop phagocytosis. Phagocytes were removed by centrifugation for 4 min at 110 x g and 4°C. The number of colony-forming units, (“CFU”) in the supernatants was determined. Control experiments were carried out simultaneously by combining  $10^7$  opsonized *S. suis* cells with EMEM (without AM).

**[00125] Killing assays.** AM ( $10^7$ /ml) and opsonized *S. suis* cells ( $10^7$ /ml) were mixed 1 : 1 and incubated for 10 min at 37°C under continuous rotation at 6 rpm. Ice-cold EMEM was added to stop further phagocytosis and killing. To remove extracellular *S. suis* cells, phagocytes were washed twice (4 min, 110 x g, 4°C) and resuspended in 5 ml EMEM containing 6% SPF serum. The tubes were incubated at 37°C under rotation at 6 rpm. After 0, 15, 30, 60 and 90 min, samples were collected and mixed with ice-cold EMEM to stop further killing. The samples were centrifuged for 4 min at 110 x g at 4°C and the phagocytic cells were lysed in EMEM containing 1% saponine for 20 min at room temperature. The number of CFU in the suspensions was determined.

**[00126] Pigs.** Germfree pigs, crossbreeds of Great Yorkshire and Dutch Landrace, were obtained from sows by caesarian sections. The surgery was performed in sterile flexible film isolators. Pigs were allotted to groups, each consisting of 4 pigs, and were housed in sterile stainless steel incubators.

**[00127] Experimental infections.** Pigs were inoculated intranasally with *S. suis* type 2 as described before. To predispose the pigs for infection with *S. suis*, five-day old pigs were inoculated intranasally with about  $10^7$  CFU of *Bordetella bronchiseptica* strain 92932. Two days later, the pigs were inoculated intranasally with *S. suis* type 2 ( $10^6$  CFU). Pigs were monitored twice daily for clinical signs of disease, such as fever, nervous signs and lameness. Blood samples were collected three times a week from each pig. White blood cells were counted with a cell counter. To monitor infection with *S. suis* and *B. bronchiseptica* and to check for absence of contaminants, we collected swabs of nasopharynx and feces daily. The swabs were plated directly onto Columbia agar containing 6% horse blood. After three weeks, the pigs were killed and examined for pathological changes. Tissue specimens from the central nervous system, serosae, and joints were examined bacteriologically and histologically as described herein (45, 49). Colonization of the serosae was



scored positively when *S. suis* was isolated from the pericardium, thoracic pleura or the peritoneum. Colonization of the joints was scored positively when *S. suis* was isolated from one or more joints (12 joints per animal were scored).

**[00128] Vaccination and challenge.** One week old pigs were vaccinated intravenously with a dosage of 10<sup>6</sup> cfu of the *S. suis* strains 10cpsEF or 10cpsB. Three weeks later, the pigs were challenged intravenously with the pathogenic serotype 2 strain 10 (10<sup>7</sup> cfu). Disease monitoring, hematological, serological and bacteriological examinations as well as post-mortum examinations were as described before under experimental infections.

**[00129] Electron Microscopy.** Bacteria were prepared for electron microscopy as described by Wagenaar et al. (50). Shortly, bacteria were mixed with agarose MP (Boehringer) of 37°C to a concentration of 0.7%. The mixture was immediately cooled on ice. Upon gelifying, samples were cut into 1 to 1.5 mm slices and incubated in a fixative containing 0.8% glutaraldehyde and 0.8% osmiumtetroxide. Subsequently, the samples were fixed and stained with uranyl acetate by microwave stimulation, dehydrated and imbedded in eponaraldite resin. Ultra-thin sections were counterstained with lead citrate and examined with a Philips CM 10 electron microscope at 80 kV. (FIG. 8.)

**[00130] Isolation of porcine alveolar macrophages (AM).** Porcine AM were obtained from the lungs of specific pathogen-free (“SPF”) pigs. Lung lavage samples were collected as described by van Leengoed et al. (43). Cells were suspended in EMEM containing 6% (v/v) SPF-pig serum and adjusted to 10 cells per ml.

## RESULTS

### Identification of the *cps* locus.

**[00131]** The *cps* locus of *S. suis* type 2 was identified through a strategy developed for the genetic identification of exported proteins (13, 31). In this system, a plasmid (pPHOS2) containing a truncated alkaline phosphatase gene (13) was used. The gene lacked the promoter sequence, the translational start site and the signal sequence. The truncated gene is preceded by a unique *Sma*I restriction site. Chromosomal DNA of *S. suis* type 2, digested with *Alu*I, was randomly cloned in this restriction site. Because translocation of PhoA across the cytoplasmic membrane of *E. coli* is

required for enzymatic activity, the system can be used to select for *S. suis* fragments containing a promoter sequence, a translational start site and a functional signal sequence. Among 560 individual *E. coli* clones tested, 16 displayed a dark blue phenotype when plated on media containing BCIP. DNA sequence analysis of the inserts from several of these plasmids was performed (results not shown) and the deduced amino acid sequences were analyzed. The hydrophobicity profile of one of the clones (pPHOS7, results not shown) showed that the N-terminal part of the sequence resembled the characteristics of a typical signal peptide: a short hydrophilic N-terminal region is followed by a hydrophobic region of 38 amino acids. These data indicate that the *phoA* system was successfully used for the selection of *S. suis* genes encoding exported proteins. Moreover, the sequences were analyzed for similarities present in the databases. The sequence of pPHOS7 showed a high similarity (37% identity) with the protein encoded by the *cps14C* gene of *Streptococcus pneumoniae* (19). This strongly suggests that pPHOS7 contains a part of the *cps* operon of *S. suis* type 2.

**[00132] Cloning of the flanking *cps* genes.** In order to clone the flanking *cps* genes of *S. suis* type 2, the insert of pPHOS7 was used as a probe to identify chromosomal DNA fragments which contain flanking *cps* genes. A 6-kb *Hind*III fragment was identified and cloned in pKUN19. This yielded clone pCPS6 (FIG. 1, part C). Sequence analysis of the insert of pCPS6 revealed that pCPS6 most probably contained the 5'-end of the *cps* locus, but still lacked the 3'-end. Therefore, sequences of the 3'-end of pCPS6 were in turn used as a probe to identify chromosomal fragments containing *cps* sequences located further downstream. These fragments were also cloned in pKUN19, resulting in pCPS17. Using the same system of chromosomal walking, plasmids pCPS18, pCPS20, pCPS23 and pCPS26, containing downstream *cps* sequences were subsequently generated.

**[00133] Analysis of the *cps* operon.** The complete nucleotide sequence of the cloned fragments was determined (FIG. 4). Examination of the compiled sequence revealed the presence of at least 13 potential open reading frames (Orfs), which were designated as Orf 2Y, Orf2X and Cps2A-Cps2K (FIG. 1, part A; FIG. 11, part A). Moreover, a 14th, incomplete Orf (Orf 2Z) was located at the 5'-end of the sequence. Two potential promoter sequences were identified. One was located 313 bp (locations 1885-1865 and 1884-1889) upstream of Orf2X. The other potential promoter sequence was located 68 bp upstream of Orf2Y (locations 2241-2236 and 2216-2211). Orf2Y is expressed in opposite orientation. Between Orfs 2Y and 2Z, the sequence contained a

potential stem-loop structure, which could act as a transcription terminator. Each Orf is preceded by a ribosome-binding site and the majority of the Orfs are very closely linked. The only significant intergenic gap was found between Cps2G and Cps2H (389 nucleotides). However, no obvious promoter sequences or potential stem-loop structures were found in this region. These data suggest that Orf2X and Cps2A-Cps2K are arranged as an operon.

[00134] An overview of all Orfs with their properties is shown in Table 2. The majority of the predicted gene products is related to proteins involved in polysaccharide biosynthesis. Orf2Z showed some similarity with the YitS protein of *Bacillus subtilis*. YitS was identified during the sequence analysis of the complete genome of *B. subtilis*. The function of the protein is unknown.

[00135] Orf2Y showed similarity with the YcxD protein of *B. subtilis* (53). Based on the similarity between YcxD and MocR of *Rhizobium meliloti* (33), YcxD was suggested to be a regulatory protein.

[00136] Orf2X showed similarity with the hypothetical YAAA proteins of *Haemophilus influenzae* and *E. coli*. The function of these proteins is unknown.

[00137] The gene products encoded by the *cps2A*, *cps2B*, *cps2C* and *cps2D* genes showed approximate similarity to the CpsA, CpsC, CpsD and CpsB proteins of several serotypes of *Streptococcus pneumoniae* (19), respectively. This suggests similar functions for these proteins. Hence, Cps2A may have a role in the regulation of the capsular polysaccharide synthesis. Cps2B and Cps2C could be involved in the chain length determination of the type 2 capsule and Cps2C can play an additional role in the export of the polysaccharide. The Cps2D protein of *S. suis* is related to the CpsB protein of *S. pneumoniae* and to proteins encoded by genes of several other Gram-positive bacteria involved in polysaccharide or exopolysaccharide synthesis, but their function is unknown (19).

[00138] The protein encoded by the *cps2E* gene showed similarity to several bacterial proteins with glycosyltransferase activities: Cps14E and Cps19fE of *S. pneumoniae* serotypes 14 and 19F (18, 19, 29), CpsE of *Streptococcus salvarius* (X94980) and CpsD of *Streptococcus agalactiae* (34). Recently, Kolkman et al. (18) showed that Cps14E is a glucosyl-1-phosphate transferase that links glucose to a lipid carrier, the first step in the biosynthesis of the *S. pneumoniae* type 14 repeating unit. Based on these data, a similar function may be fulfilled by Cps2E of *S. suis*.

[00139] The protein encoded by the *cps2F* gene showed similarity to the protein encoded by the *rfbU* gene of *Salmonella enteritica* (25). This similarity is most pronounced in the C-terminal regions of these proteins. The *rfbU* gene was shown to encode mannosyltransferase activity (25).

[00140] The *cps2G* gene encoded a protein that showed moderate similarity with the *rfbF* gene product of *Campylobacter hyoilei* (22), the *epsF* gene product of *S. thermophilus* (40) and the *capM* gene product of *S. aureus* (24). On the basis of similarity, the *rfbF*, *epsF* and *capM* genes are suggested to encode galactosyltransferase activities. Hence, a similar glycosyltransferase activity could be fulfilled by the *cps2G* gene product.

[00141] The *cps2H* gene encodes a protein that is similar to the N-terminal region of the *lgtD* gene product of *Haemophilus influenzae* (U32768). Moreover, the hydrophobicity plots of Cps2H and LgtD looked very similar in these regions (data not shown). Based on sequence similarity, the *lgtD* gene product was suggested to have glycosyltransferase activity (U32768).

[00142] The gene product encoded by the *cps2I* gene showed some similarity with a protein of *Actinobacillus actinomycetemcomitans* (AB002668). This protein is part of the gene cluster responsible for the serotype-b-specific antigen of *A. actinomycetemcomitans*. The function of the protein is unknown.

[00143] The gene products encoded by the *cps2J* and *cps2K* genes showed significant similarities to the Cps14J protein of *S. pneumoniae*. The *cps14J* gene of *S. pneumoniae* was shown to encode a  $\beta$ -1,4-galactosyltransferase activity. In *S. pneumoniae*, CpsJ is responsible for the addition of the fourth (*i.e.* last) sugar in the synthesis of the *S. pneumoniae* serotype 14 polysaccharide (20). Even some similarity was found between Cps2J and Cps2K (FIG. 2, 25.5% similarity). This similarity was most pronounced in the N-terminal regions of the proteins (FIG. 7). Recently, two small conserved regions were identified in the N-terminus of Cps14J and Cps14I and their homologues (20). These regions were predicted to be important for catalytic activity. Both regions, DXS and DXDD (FIG. 2), were also found in Cps2J and Cps2K.

[00144] **Distribution of the *cps2* genes in other *S. suis* serotypes.** To examine the relationship between the *cps2* genes and *cps* genes in the other *S. suis* serotypes, we performed cross-hybridization experiments. DNA fragments of the individual *cps2* genes were amplified by PCR, labeled with  $^{32}\text{P}$ , and used to probe Southern blots of chromosomal DNA of the reference

strains of the 35 different *S. suis* serotypes. Large variations in the hybridization patterns were observed (Table 4). As a positive control, we used a probe specific for 16S rRNA. The 16S rRNA probe hybridized with all serotypes tested. However, none of the other genes tested were common in all serotypes. Based on the genetic organization of the genes, it was previously suggested that *orfX* and *cpsA-cpsK* genes are part of one operon and that the proteins encoded by these genes are all involved in polysaccharide biosynthesis. OrfY and OrfZ are not a part of this operon, and their role in the polysaccharide biosynthesis is unclear. Based on sequence similarity data, OrfY may be involved in regulation of the *cps2* genes. OrfZ is proposed to be unrelated to polysaccharide biosynthesis. Probes specific for the *orfZ*, *orfY*, *orfX*, *cpsA*, *cpsB*, *cpsC* and *cpsD* genes hybridized with most other serotypes. This suggests that the proteins encoded by these genes are not type-specific, but may perform more common functions in biosynthesis of the capsular polysaccharide. This confirms previous data which showed that the *cps2A-cps2D* genes showed strong similarity to *cps* genes of several serotypes of *Streptococcus pneumoniae*. Based on this similarity, Cps2A is possibly a regulatory protein, whereas Cps2B and Cps2C may play a role in length determination and export of polysaccharide. The *cps2E* gene hybridized with DNA of serotypes 1, 2, 14 and 1/2. The *cps2E* gene showed a strong similarity to the *cps14E* gene of *S. pneumoniae* (18). This enzyme was shown to have a glucosyl-1-phosphate activity and catalyzed the transfer of glucose to a lipid carrier (18). These data indicate that a glycosyltransferase closely related to Cps14E may be responsible for the first step in the biosynthesis of polysaccharide in the *S. suis* serotypes 1, 2, 14 and 1/2. The *cps2F*, *cps2G*, *cps2H*, *cps2I* and *cps2J* genes hybridized with chromosomal DNA of serotypes 2 and 1/2 only. The *cps2G* gene showed an additional weak hybridization signal with DNA of serotype 34. In agglutination tests, serotype 1/2 showed agglutination with sera specific for serotype 2 as well as with sera specific for serotype 1. This suggests that serotype 1/2 shares antigenic determinants with both types 1 and 2. The hybridization data confirmed these data. All putative glycosyltransferases present in serotype 2 are also present in serotype 1/2. The *cps2K* gene showed a hybridization pattern similar to the *cps2E* gene. Hybridization was observed with DNA of serotypes 1, 2, 14 and 1/2. Taken together, these hybridization data show that the *cps2* gene cluster can be divided into three regions: a central region

containing the type-specific genes is flanked by two regions containing common genes for various serotypes.

**[00145] Cloning of the type-specific *cps* genes of serotypes 1 and 9.** To clone the type-specific *cps* genes of *S. suis* serotype 1, the *cps2E* gene was used as a probe to identify chromosomal DNA fragments of type 1 which contain flanking *cps* genes. A 5 kb *EcoRV* fragment was identified and cloned in pKUN19. This yielded pCPS1-1 (FIG. 1, part B). This fragment was in turn used as a probe to identify an overlapping 2.2 kb *HindIII* fragment. pKUN19 containing this *HindIII* fragment was designated pCPS1-2. The same strategy was followed to identify and clone the type-specific *cps* genes of serotype 9. In this case, we used the *cps2D* gene as a probe. A 0.8 kb *HindIII-XbaI* fragment was identified and cloned, yielding pCPS9-1 (FIG. 1, part C). This fragment was in turn used as a probe to identify a 4 kb *XbaI* fragment. pKUN19 containing this 4 kb *XbaI* fragment was designated pCPS9-2.

**[00146] Analysis of the cloned *cpsI* genes.** The complete nucleotide sequence of the inserts of pCPS1-1 and pCPS1-2 was determined (FIG. 5). Examination of the sequence revealed the presence of five complete and two incomplete Orfs (FIG. 1, part B). Each Orf is preceded by a ribosome-binding site. In accord with data obtained for the *cps2* genes of serotype 2, the majority of the Orfs is very closely linked. The only significant gap (718 bp) was found between Cps1G and Cps1H. No obvious promoter sequences or potential stem-loop structures could be found in this region. This suggests that, as in serotype 2, the *cps* genes in serotype 1 are arranged in an operon.

**[00147]** An overview of the Orfs and their properties is shown in Table 2. As expected on the basis of the hybridization data (Table 4), the protein encoded by the *cps1E* gene was related to Cps2E of *S. suis* type 2 (identity of 86%). The fragment cloned in pCPS1-1 lacked the coding region for the first 7 amino acids of the *cps1E* gene.

**[00148]** The protein encoded by the *cps1F* and *cps1G* genes showed strong similarity to the Cps14F and Cps14G proteins of *Streptococcus pneumoniae* serotype 14, respectively (20). The function of the Cps14F is not completely clear, but it has been suggested that Cps14F has a role in glycosyltransferase activity. The *cps14G* gene of *S. pneumoniae* was shown to encode  $\beta$ -1,4-galactosyltransferase activity. In *S. pneumoniae* type 14, this activity is required for the second step in the biosynthesis of the oligosaccharide subunit (20). Based on the similarity of the data,

similar glycosyltransferase and enhancing activities are suggested for the *cps1G* and *cps1F* genes of *S. suis* type 1.

**[00149]** The protein encoded by the *cps1H* gene showed similarity to the Cps14H protein of *S. pneumoniae* (20). Based on sequence similarity, Cps14H was proposed to be the polysaccharide polymerase (20).

**[00150]** The protein encoded by the *cps1I* gene showed some similarity with the Cps14J protein of *S. pneumoniae* (19). The *cps14J* gene was shown to encode a  $\beta$ -1,4-galactosyltransferase activity, responsible for the addition of the fourth (*i.e.* last) sugar in the synthesis of the *S. pneumoniae* serotype 14 polysaccharide.

**[00151]** Between Cps1G and Cps1H, a gap of 718 bp was found. This region revealed three small Orfs. The three Orfs were expressed in three different reading frames and were not preceded by potential ribosome binding sites, nor contained potential start sites. However, the three potential gene products encoded by this region showed some similarity with three successive regions of the C-terminal part of the EpsK protein of *Streptococcus thermophilus* (27% identity, 40). The region related to the first 82 amino acids is lacking.

**[00152] Analysis of the cloned *cps9* genes.** We also determined the complete nucleotide sequence of the inserts of pCPS9-1 and pCPS9-2 (FIG. 6). Examination of the sequence revealed the presence of three complete and two incomplete Orfs (FIG. 1, part C). As in serotypes 1 and 2, all Orfs are preceded by a ribosome-binding site and are very closely coupled. As suggested by the hybridization data (Table 4), the Cps2D and Cps9D proteins were highly related (Table 2). Based on sequence comparisons, pCPS9-1 lacked the first 27 amino acids of the Cps9D protein.

**[00153]** The protein encoded by the *cps9E* gene showed some similarity with the CapD protein of *Staphylococcus aureus* serotype 1 (24). Based on sequence similarity data, the Cap1D protein was suggested to be an epimerase or a dehydratase involved in the synthesis of N-acetylfructosamine or N- acetylgalactosamine (63).

**[00154]** Cps9F showed some similarity to the CapM proteins of *S. aureus* serotypes 5 and 8 (61, 64, 65). Based on sequence similarity data, Cap5M and Cap8M are proposed to be glycosyltransferases (63).

[00155] The protein encoded by the *cps9G* gene showed some similarity to a protein of *Actinobacillus actinomycetemcomitans* (AB002668\_4). This protein is part of a gene cluster responsible for the serotype b-specific antigens of *Actinobacillus actinomycetemcomitans*. The function of the protein is unknown.

[00156] The protein encoded by the *cps9H* gene showed some similarity to the *rfbB* gene of *Yersinia enterocolitica* (68). The RfbB protein was shown to be essential for O-antigen synthesis, but the function of the protein in the synthesis of the O:3 lipopolysaccharide is unknown.

[00157] **Serotype 1 and serotype 9-specific *cps* genes.** To determine whether the cloned fragments in pCPS1-1, pCPS1-2, pCPS9-1 and pCPS9-2 contained the type-specific genes for serotype 1 and 9, respectively, cross-hybridization experiments were performed. DNA fragments of the individual *cps1* and *cps9* genes were amplified by PCR, labeled with <sup>32</sup>P, and used to probe Southern blots of chromosomal DNA of the reference strains of the 35 different *S. suis* serotypes. The results are shown in Table 5. Based on the data obtained with the *cps2E* probe (Table 4), the *cps1E* probe was expected to hybridize with chromosomal DNA of *S. suis* serotypes 1, 2, 14, 27 and 1/2. The *cps1H*, *cps9E* and *cps9F* probes hybridized with most other serotypes. However, the *cps1F* and *cps1G* and *cps1I* probes hybridized with chromosomal DNA of serotypes 1 and 14 only. The *cps9G* and *cps9H* probes hybridized with serotype 9 only. These data suggest that the *cps9G* and *cps9H* probes are specific for serotype 9 and, therefore, could be useful tools for the development of rapid and sensitive diagnostic tests for *S. suis* type 9 infections.

[00158] **Type-specific PCR.** So far, the probes were tested on the 35 different reference strains only. To test the diagnostic value of the type-specific *cps* probes further, several other *S. suis* serotype 1, 2, 1/2, 9 and 14 strains were used. Moreover, since a PCR-based method would be even more rapid and sensitive than a hybridization test, we tested whether we could use a PCR for the serotyping of the *S. suis* strains. The oligonucleotide primer sets were chosen within the *cps2J*, *cps1I* and *cps9H* genes. Amplified fragments of 675 bp, 380 bp and 390 bp were expected, respectively. The results show that 675 bp fragments were amplified on type 2 and 1/2 strains using *cps2J* primers; 380 bp fragments were amplified on type 1 and 14 strains using *cps1I* primers and 390 bp fragments were amplified on type 9 strains using *cps9H* primers.



**[00159] Construction of mutants impaired in capsule production.** To evaluate the role of the capsule of *S. suis* type 2 in pathogenesis, we constructed two isogenic mutants in which capsule production was disturbed. To construct mutant 10cpsB, pCPS11 was used. In this plasmid, a part of the *cps2B* gene was replaced by the spectinomycin-resistance gene. To construct mutant strain 10cpsEF, the plasmid pCPS28 was used. In pCPS28, the 3'-end of *cps2E* gene, as well as the 5'-end of *cps2F* gene, were replaced by the spectinomycin-resistance gene. pCPS11 and pCPS28 were used to electrotransform strain 10 of *S. suis* type 2 and spectinomycin-resistant colonies were selected. Southern blotting and hybridization experiments were used to select double cross-over integration events (results not shown). To test whether the capsular structure of the strains 10cpsB and 10cpsEF was disturbed, we used a slide agglutination test using a suspension of the mutant strains in hyperimmune anti-*S. suis* type 2 serum (44). The results showed that even in the absence of serotype-specific antisera, the bacteria agglutinated. This indicates that, in the mutant strains, the capsular structure was disturbed. To confirm this, thin sections of wild-type and mutant strains were compared by electron microscopy. The results showed that, compared to the wild-type (FIG. 3, part A), the amount of capsule produced by the mutant strains was greatly reduced (FIG. 3, parts B and C). Almost no capsular material could be detected on the surface of the mutant strains.

#### **Capsular mutants are sensitive to phagocytosis and killing by porcine alveolar macrophages (“PAM”).**

**[00160]** The capsular mutants were tested for their ability to resist phagocytosis by PAM in the presence of porcine SPF serum. The wild-type strain 10 seemed to be resistant to phagocytosis under these conditions (FIGS. 9A and 9B). In contrast, the mutant strains were efficiently ingested by macrophages (FIGS. 9A and 9B). After 90 minutes, more than 99.7% (strain 10cpsB) and 99.8% (strain 10cpsEF) of the mutant cells were ingested by the macrophages. Moreover, as shown in FIGS. 9A and 9B, the ingested strains were efficiently killed by the macrophages. 90-98% of all ingested cells were killed within 90 min. No differences could be observed between wild-type and mutant strains. These data indicate that the capsule of *S. suis* type 2 efficiently protects the organism from uptake by macrophages *in vitro*.

**[00161] Capsular mutants are less virulent for germfree piglets.** The virulence properties of the wild-type and mutant strains were tested after experimental infection of newborn germfree pigs (45, 49). Table 1 shows that specific and nonspecific signs of disease could be observed in all pigs inoculated with the wild-type strain. Moreover, all pigs inoculated with the wild-type strain died during the course of the experiment or were killed because of serious illness or nervous disorders (Table 3). In contrast, the pigs inoculated with strains 10cpsB and 10cpsEF showed no specific signs of disease and all pigs survived until the end of the experiment (Table 6). The temperature of the pigs inoculated with the wild-type strain increased 2 days after inoculation and remained high until day 5 (Table 3). The temperature of the pigs inoculated with the mutant strains sometimes exceeded 40°C, however, we could observe significant differences in the fever index (*i.e.*, percent of observations in an experimental group during which pigs showed fever (>40°C)) between pigs inoculated with wild-type and mutant strains. All pigs showed increased numbers of polymorphonuclear leucocytes (PMLs) ( $>10 \times 10^9$  PMLs per liter) (Table 3). However, in pigs inoculated with the mutant strains, the percentage of samples with increased numbers of PMLs was considerably lower. *S. suis* strains and *B. bronchiseptica* could be isolated from the nasopharynx and feces swab samples of all pigs from 1 day post-infection until the end of the experiment (Table 3). Postmortem, the wild-type strain could frequently be isolated from the central nervous system (“CNS”), kidney, heart, liver, spleen, serosae, joints and tonsils. Mutant strains could easily be recovered from the tonsils, but were never recovered from the kidney, liver or spleen. Interestingly, low numbers of the mutant strains were isolated from the CNS, the serosae, the joints, the lungs and the heart. Taken together, these data strongly indicated that mutant *S. suis* strains, impaired in capsule production, are not virulent for young germfree pigs.

**[00162]** We describe the identification and the molecular characterization of the *cps* locus, involved in the capsular polysaccharide biosynthesis, of *S. suis*. Most of the genes seemed to belong to a single transcriptional unit, suggesting a coordinate control of these genes. Functions to most of the gene products were assigned. Regions involved in regulation (Cps2A), chain length determination (Cps2B, C), export (Cps2C) and biosynthesis (Cps2E, F, G, H, J, K) were identified. The region involved in biosynthesis is located at the center of the gene cluster and is flanked by two regions containing genes with more common functions. The incomplete *orf22* gene was located at

the 5'-end of the cloned fragment. Orf2Z showed some similarity with the YitS protein of *B. subtilis*. However, because the function of the YitS protein is unknown, this did not give us any information about the possible function of Orf2Z. Because the *orf2Z* gene is not a part of the *cps* operon, a role of this gene in polysaccharide biosynthesis is not expected. The Orf2Y protein showed some similarity with the YcxD protein of *B. subtilis* (53). The YcxD protein was suggested to be a regulatory protein. Similarly, Orf2Y may be involved in the regulation of polysaccharide biosynthesis. The Orf2X protein showed similarity with the YAAA proteins of *H. influenzae* and *E. coli*. The function of these proteins is unknown. In *S. suis* type 2, the *orf2X* gene seemed to be the first gene in the *cps2* operon. This suggests a role of Orf2X in the polysaccharide biosynthesis. In *H. influenzae* and *E. coli*, however, these proteins are not associated with capsular gene clusters. The analysis of isogenic mutants impaired in the expression of Orf2X should give more insight in the presumed role of Orf2X in the polysaccharide biosynthesis of *S. suis* type 2.

[00163] The gene products encoded by the *cps2E*, *cps2F*, *cps2G*, *cps2H*, *cps2J* and *cps2K* genes showed little similarity with glycosyltransferases of several Gram-positive or Gram-negative bacteria (18, 19, 20, 22, 25). The *cps2E* gene product shows some similarity with the Cps14E protein of *S. pneumoniae* (18, 19). Cps14E is a glucosyl-1-phosphate transferase that links glucose to a lipid carrier (18). In *S. pneumoniae* this is the first step in the biosynthesis of the oligosaccharide repeating unit. The structure of the *S. suis* serotype 2 capsule contains glucose, galactose, rhamnose, N-acetyl glucosamine and sialic acid in a ratio of 3:1:1:1:1 (7). Based on these data, we conclude that Cps2E of *S. suis* has glucosyltransferase activity and is involved in the linkage of the first sugar to the lipid carrier.

[00164] The C-terminal region of the *cps2F* gene product showed some similarity with the RfbU of *Salmonella enteritica*. RfbU was shown to have mannosyltransferase activity (24). Because mannosyl is not a component of the *S. suis* type 2 polysaccharide, a mannosyltransferase activity is not expected in this organism. Nevertheless, *cps2F* encodes a glycosyltransferase with another sugar specificity.

[00165] Cps2G showed moderate similarity to a family of gene products suggested to encode galactosyltransferase activities (22, 24, 40). Hence, a similar activity is shown for Cps2G.

[00166] Cps2H showed some similarity with LgtD of *H. influenzae* (U32768). Because LgtD was proposed to have glycosyltransferase activity, a similar activity is fulfilled by Cps2H.

[00167] Cps2J and Cps2K showed similarity to Cps14J of *S. pneumoniae* (20). Cps2J showed similarity with Cps14I of *S. pneumoniae* as well. Cps14I was shown to have N-acetyl glucosaminyltransferase activity, whereas Cps14J has a  $\beta$ -1,4-galactosyltransferase activity (20). In *S. pneumoniae*, Cps14I is responsible for the addition of the third sugar and Cps14J for the addition of the last sugar in the synthesis of the type 14 repeating unit (20). Because the capsule of *S. suis* type 2 contains galactose as well as N-acetyl glucosamine components, galactosyltransferase as well as N-acetyl glucoaminyltransferase activities could be envisaged for the *cps2J* and *cps2K* gene products, respectively. As was observed for Cps14I and Cps14J, the N-termini of Cps2J and Cps2K showed a significant degree of sequence similarity. Within the N-terminal domains of Cps14I and Cps14J, two small regions were identified, which were also conserved in several other glycosyltransferases (22). Within these two regions, two Asp residues were proposed to be important for catalytic activity. The two conserved regions, DXS and DXDD, were also found in Cps2J and Cps2K.

[00168] The function of Cps2I remains unclear. Cps2I showed some similarity with a protein of *A. actinomycetemcomitans*. Although this protein part is of the gene cluster responsible for the serotype-B-specific antigens, the function of the protein is unknown.

[00169] We further describe the identification and characterization of the *cps* genes specific for *S. suis* serotypes 1, 2 and 9. After the entire *cps2* locus of *S. suis* serotype 2 was cloned and characterized, functions for most of the *cps2* gene products could be assigned by sequence homologies. Based on these data, the glycosyltransferase activities, required for type specificity, could be located in the center of the operon. Cross-hybridization experiments, using the individual *cps2* genes as probes on chromosomal DNAs of the 35 different serotypes, confirmed this idea. The regions containing the type-specific genes of serotypes 1 and 9 could be cloned and characterized, showing that an identical genetic organization of the *cps* operons of other *S. suis* serotypes exists. The *cps1E*, *cps1F*, *cps1G*, *cps1H*, and *cps1I* genes revealed a striking similarity with *cps14E*, *cps14F*, *cps14G*, *cps14H* and *cps14J* genes of *S. pneumoniae*. Interestingly, *S. pneumoniae* serotype 14 is the serotype most commonly associated with pneumococcal infections in young children (54),

whereas *S. suis* serotype 1 strains are most commonly isolated from piglets younger than 8 weeks (46). In *S. pneumoniae*, the *cps14E*, *cps14G*, *cps14I* and *cps14J* encode the glycosyltransferases required for the synthesis of the type 14 tetrameric repeating unit, showing that the *cps1E*, *cps1G* and *cps1I* genes encoded glycosyltransferases. The precise functions of these genes as well as the substrate specificities of the enzymes can be established. In *S. pneumoniae*, the *cps14E* gene was shown to encode a glucosyl-1-phosphate transferase catalyzing the transfer of glucose to a lipid carrier. Moreover, *cpsE-like* genes were found in *S. pneumoniae* serotypes 9N, 13, 14, 15B, 15C, 18F, 18A and 19F (60). *CpsE* mutants were constructed in the serotypes 9N, 13, 14 and 15B. All mutant strains lacked glycosyltransferase activity (60). Moreover, in all these *S. pneumoniae* serotypes, the *cpsE* gene seemed to be responsible for the addition of glucose to the lipid carrier. Based on these data, we suggest that in *S. suis* type 1, the *cps1E* gene may fulfill a similar function. The structure of the *S. suis* type 1 capsule is unknown, but it is composed of glucose, galactose, N-acetyl glucosamine, N-acetyl galactosamine and sialic acid in a ratio of 1: 2.4: 1: 1:1.4 (5). Therefore, a role of a *cpsE-like* glycosyltransferase activity can easily be envisaged. *CpsE-like* sequences were also found in serotypes 2, 1/2 and 14.

[00170] For polysaccharide biosynthesis in *S. pneumoniae* type 14, transfer of the second sugar of the repeating unit to the first lipid-linked sugar is performed by the gene products of *cps14F* and *cps14G* (20). Similar to Cps14F and Cps14G, the *S. suis* type 1 proteins Cps1F and Cps1G may act as one glycosyltransferase performing the same reaction. Cps14F and Cps14G of *S. pneumoniae* showed similarity to the N-terminal half and C-terminal half of the SpsK protein of *Sphingomonas* (20, 67), respectively. This suggests a combined function for both proteins. Moreover, *cps14F*- and *cps14G*-like sequences were found in several serotypes of *S. pneumoniae* and these genes always seemed to exist together (60). The same was observed for *S. suis* type 1. The *cps1F* and *cps1G* probes hybridized with type 1 and type 14 strains.

[00171] According to the similarity found between the *cps1H* gene and the *cps14H* gene of *S. pneumoniae* (20), *cps1H* is expected to encode a polysaccharide polymerase.

[00172] The protein encoded by the *cps1I* gene showed some similarity with the Cps14J protein of *S. pneumoniae* (19). The *cps14J* gene was shown to encode a  $\beta$ -1,4-galactosyltransferase activity, responsible for the addition of the fourth (*i.e.* last) sugar in the synthesis of the *S.*

*pneumoniae* serotype 14 polysaccharide. In *S. suis* type 2, the proteins encoded by the *cps2J* and *cps2K* genes showed similarity to the Cps14J protein. However, no significant homologies were found between Cps2J, Cps2K and Cps1I. In the N-terminal regions of Cps14J and Cps14I, two small conserved regions, DXS and DXDD, were identified (19). These regions seemed to be important for catalytic activity (13). At the same positions in the sequence, Cps2I contained the regions DXS and DXED.

[00173] In the region between Cps1G and Cps1H, three small Orfs were identified. Since the Orfs were expressed in three different reading frames, and did not contain potential start sites, expression is not expected. However, the three potential gene products encoded by this region showed some similarity with three successive regions of the C-terminal part of the EpsK (protein of *Streptococcus thermophilus* (27% identity, 40). The region related to the first 82 amino acids is lacking. The EpsK protein was suggested to play a role in the export of the exopolysaccharide by rendering the polymerized exopolysaccharide more hydrophobic through a lipid modification. These data could suggest that the sequences in the region between Cps1G and Cps1H originated from an *epsK*-like sequence. Hybridization experiments showed that this *epsK*-like region is also present in other serotype 1 strains as well as in serotype 14 strains (results not shown).

[00174] The function of most of the cloned serotype 9 genes can be established. Based on sequence similarity data, the *cps9E* and *cps9F* genes could be glycosyltransferases (61, 24, 63, 64, 65). Moreover, the *cps9G* and *cps9H* genes showed similarity to genes located in regions involved in polysaccharide biosynthesis, but the function of these genes is unknown (68).

[00175] Cross-hybridization experiments using the individual *cps2*, *cps1* and *cps9* genes as probes showed that the *cps9G* and *cps9H* probes specifically hybridized with serotype 9 strains. Therefore, these are useful as tools for the identification of *S. suis* type 9 strains both for diagnostic purposes as well as in epidemiological and transmission studies. We previously developed a PCR method which can be used to detect *S. suis* strains in nasal and tonsil swabs of pigs (62). The method was used to identify pathogenic (EF-positive) strains of *S. suis* serotype 2. Besides *S. suis* type 2 strains, serotype 9 strains are frequently isolated from organs of diseased pigs. However, until now, a rapid and sensitive diagnostic test was not available for type 9 strains. Therefore, the type 9-specific probes or the type 9-specific PCR is of great diagnostic value. The *cps1F*, *cps1G* and

*cps1I* probes hybridized with serotype 1 as well as with serotype 14 strains. In coagglutination tests, type 1 strains react with the anti-type 1 as well as with the anti-type 14 antisera (56). This suggests the presence of common epitopes between these serotypes. On the other hand, type 1 strains agglutinated only with anti-type 1 serum (56, 57), indicating that it is possible to detect differences between those serotypes.

[00176] The *cps2F*, *cps2G*, *cps2H*, *cps2I* and *cps2J* probes hybridized with serotypes 2 and 1/2 only. Serotype 34 showed a weak hybridizing signal with the *cps2G* probe. As shown in agglutination tests, type 1/2 strains react with sera directed against type 1 as well as with sera directed against type 2 strains (46). Therefore, type 1/2 shared antigens with both types 1 and 2. Based on the hybridization patterns of serotype 1/2 strains with the *cps1*- and *cps2*-specific genes, serotype 1/2 seemed to be more closely related to type 2 strains than to type 1 strains. In our current studies, we identify type-specific genes, primers or probes which are used for the discrimination of serotypes 1, 14 and 2 and 1/2 and others of the 35 serotypes yet known. Furthermore, type-specific genes, primers or probes can now easily be developed for yet unknown serotypes, once they become isolated.

### **Cloning and characterization of a further part of the *cps2* locus.**

[00177] Based on the established sequence, 11 genes, designated *cps2L* to *cps2T*, *orf2U* and *orf2V*, were identified. A gene homologous to genes involved in the polymerization of the repeating oligosaccharide unit (*cps20*) as well as genes involved in the synthesis of sialic acid (*cps2P* to *cps2T*) were identified. Moreover, hybridization experiments showed that the genes involved in the sialic acid synthesis are present in *S. suis* serotypes 1, 2, 14, 27 and 1/2. The “*cps2M*” and “*cps2N*” regions showed similarity to proteins involved in the polysaccharide biosynthesis of other Gram-positive bacteria. However, these regions seemed to be truncated or were non-functional as the result of frame-shift or point mutations. At its 3'-end, the *cps2* locus contained two insertional elements (“*orf2U*” and “*orf2V*”), both of which seemed to be non-functional.

[00178] To clone the remaining part of the *cps2* locus, sequences of the 3'-end of pCPS26 (FIG. 1, part C) were used to identify a chromosomal fragment containing *cps2* sequences located further downstream. This fragment was cloned in pKUN19, resulting in pCPS29. Using a similar

approach, we subsequently isolated the plasmids pCPS30 and pCPS34 containing downstream *cps2* sequences (FIG. 1, part C).

### **Analysis of the *cps2* operon.**

[00179] The complete nucleotide sequence of the cloned fragments was determined. Examination of the compiled sequence revealed the presence of: a sequence encoding the C-terminal part of Cps2K, six apparently functional genes (designated *cps2O-cps2T*) and the remnants of 5 different ancestral genes (designated “*cps2L*,” “*cps2M*,” “*cps2N*,” “*orf2U*” and “*orf2V*”). The latter genes seemed to be truncated or incomplete as the result of the presence of stop codons or frame-shift mutations (FIG. 1, part A). Neither potential promoter sequences nor potential stem-loop structures could be identified within the sequenced region. A ribosome-binding site precedes each ORF and the majority of the ORFs are very closely linked. Three intergenic gaps were found: one between “*cps2M*” and “*cps2N*” (176 nucleotides), one between *cps2O* and *cps2P* (525 nucleotides), and one between *cps2T* and “*orf2U*” (200 nucleotides). These and our above data show that *Orf2X* and *Cps2A-Orf2T* are part of a single operon.

[00180] A list of all loci and their properties is shown in Table 4. The “*cps2L*” region contained three potential ORFs of 103, 79 and 152 amino acids, respectively, which were only separated from each other by stop codons. Only the first ORF is preceded by a potential ribosomal binding site and contained a methionine start codon. This suggests that “*cps2L*” originates from an ancestral *cps2L* gene, which coded for a protein of 339 amino acids. The function of this hypothetical *cps2L* protein remains unclear so far: no significant homologies were found between *Cps2L* and proteins present in the data libraries. It is not clear whether the first ORF of the “*cps2L*” region is expressed into a protein of 103 amino acids. The “*cps2M*” region showed homology to the N-terminal 134 amino acids of the NeuA proteins of *Streptococcus agalactiae* and *Escherichia coli* (AB017355, 32). However, although the “*cps2M*” region contained a potential ribosome binding site, a methionine start codon was absent. Compared with the *S. agalactiae* sequence, the ATG start codon was replaced by a lysin encoding AAG codon. Moreover, the region homologous to the first 58 amino acids of the *S. agalactiae* NeuA (identity 77%) was separated from the region homologous to amino acids 59-134 of NeuA by a repeated DNA sequence of 100-bp (*see herein*). In addition, the



region homologous to amino acids 59 to 95 of NeuA (identity 32%) and the region homologous to the amino acids 96 to 134 of NeuA (identity 50%) were present in different reading frames. Therefore, the partial and truncated NeuA homologue is probably nonfunctional in *S. suis*. The “cps2N” region showed homology to CpsJ of *S. agalactiae* (accession no. AB017355). However, sequences homologous to the first 88 amino acids of CpsJ were lacking in *S. suis*. Moreover, the homologous region was present in two different reading frames. The protein encoded by the cps2O gene showed homology to proteins of several streptococci involved in the transport of the oligosaccharide repeating unit (accession no. AB017355), suggesting a similar function for Cps2O. The proteins encoded by the cps2P, cps2S and cps2T genes showed homology to the NeuB, NeuD and NeuA proteins of *S. agalactiae* and *E. coli* (accession no AB017355). Because the “cps2M” region also showed homology to NeuA of *E. coli*, the *S. suis* cps2 locus contains a functional neuA gene (cps2T) as well as a nonfunctional (“cps2M”) gene. The mutual homology between these two regions showed an identity of 77% at the amino acid level over amino acids 1-58 and 49% over the amino acids 59-134. Cps2Q and Cps2R showed homology to the N-terminal and C-terminal parts of the NeuC protein of *S. agalactiae* and *E. coli*, respectively. This suggests that the function of the *S. agalactiae* NeuC protein in *S. suis* is likely fulfilled by two different proteins. In *E. coli*, the neu genes are known to be involved in the synthesis of sialic acid. NeuNAc is synthesized from N-acetylmannosamine and phosphoenolpyruvate by NeuNAc synthetase. Subsequently, NeuNAc is converted to CMP-NeuNAc by the enzyme CMP-NeuNAc synthetase. CMP-NeuNAc is the substrate for the synthesis of polysaccharide. In *E. coli*, K1 NeuB is the NeuNAc synthetase, and NeuA is the CMP-NeuNAc synthetase. NeuC has been implicated in the NeuNAc synthesis, but its precise role is not known. The precise role of NeuD is not known. A role of the Cps2P-Cps2T proteins in the synthesis of sialic acid can easily be envisaged, since the capsule of *S. suis* serotype 2 is rich in sialic acid. In *S. agalactiae*, sialic acid has been shown to be critical to the virulence function of the type III capsule. Moreover, it has been suggested that the presence of sialic acid in the capsule of bacteria which can cause meningitis may be important for these bacteria to breach the blood-brain barrier. So far, however, the requirement of the sialic acid for virulence of *S. suis* remains unclear.

[00181] “Orf2U” and “Orf2V” showed homology to proteins located on two different insertional elements. “Orf2U” is homologous to IS1194 of *Streptococcus thermophilus*, whereas “Orf2V” showed homology to a putative transposase of *Streptococcus pneumoniae*. This putative transposase was recently found to be associated with the type 2 capsular locus of *S. pneumoniae*. Compared with the original insertional elements in *S. thermophilus* and *S. pneumoniae*, both “Orf2U” and “Orf2V” are likely to be non-functional due to frame shift mutations within their coding regions.

[00182] A striking observation was the presence of a sequence of 100 bp (FIG. 10) which was repeated three times within the cps2 operon. The sequence is highly conserved (between 94% and 98%) and was found in the intergenic regions between cps2G and cps2H, within “cps2M” and between cps2O and cps2P. No significant homologies were found between this 100-bp direct repeat sequence and sequences present in the data libraries, suggesting that the sequence is unique for *S. suis*.

#### **Distribution of the cps2 sequences among the 35 *S. suis* serotypes.**

[00183] To examine the presence of sialic acid encoding genes in other *S. suis* serotypes, we performed cross-hybridization experiments. DNA fragments of the individual cps2 genes were amplified by PCR, radiolabeled with <sup>32</sup>P and hybridized to chromosomal DNA of the reference strains of the 35 different *S. suis* serotypes. As a positive control, we used a probe specific for *S. suis* 16S rRNA. The 16S rRNA probe hybridized with almost equal intensities to all serotypes tested (Table 4). The “cps2L” sequence hybridized with DNA of serotypes 1, 2, 14 and 1/2. The “cps2M”, cps2O, cps2P, cps2Q, cps2R, cps2S and cps2T genes hybridized with DNA of serotypes 1, 2, 14, 27 and 1/2. Because the cps2P-cps2T genes are most likely involved in the synthesis of sialic acid, these results suggest that sialic acid is also a part of the capsule in the *S. suis* serotypes 1, 2, 14, 27 and 1/2. This is in agreement with the finding that the serotypes 1, 2 and 1/2 possess a capsule that is rich in sialic acid. Although the chemical compositions of the capsules of serotypes 14 and 27 are unknown, recent agglutination studies using sialic acid-binding lectins suggested the presence of sialic acid in *S. suis* serotype 14, but not in serotype 27. In these studies, sialic acid was also detected in serotypes 15 and 16. Since the latter observation is not in agreement with our

hybridization studies, it might be that other genes, not homologous to the *cps2P-cps2T* genes, are responsible for the sialic acid synthesis in serotypes 15 and 16.

[00184] A probe based on “*cps2N*” sequences hybridized with DNA from serotypes 1, 2, 14 and 1/2. A probe specific for “*orf2U*” hybridized with serotypes 1, 2, 7, 14, 24, 27, 32, 34, and 1/2, whereas a probe specific for “*orf2V*” hybridized with many different serotypes. In addition, we prepared a probe specific for the 100-bp direct repeat sequence. This probe hybridized with the serotypes 1, 2, 13, 14, 22, 24, 27, 29, 32, 34 and 1/2 (Table 4). To analyze the number of copies of the direct repeat sequence within the *S. suis* serotype 2 chromosome, a Southern blot hybridization and analysis was performed. Therefore, chromosomal DNA of *S. suis* serotype 2 was digested with *NcoI* and hybridized with a <sup>32</sup>P-labeled direct repeat sequence. Only one hybridizing fragment, containing the three direct repeats present on the *cps2* locus, was found (results not shown). This indicates that the 100-bp direct repeat sequence is only associated with the *cps2* locus. In *S. pneumoniae*, a 115-bp long repeated sequence was found to be associated with the capsular genes of serotypes 1, 3, 14 and 19F. In *S. pneumoniae*, this 115-bp sequence was also found in the vicinity of other genes involved in pneumococcal virulence (hyaluronidase and neuraminidase genes). A regulatory role of the 115-bp sequence in coordinate control of these virulence-related genes was suggested.

[00185] To study the role of the capsule in resistance to phagocytosis and in virulence, we constructed two isogenic mutants in which capsule synthesis was disturbed. In 10*cpsB*, the *cps2B* gene was disturbed by the insertion of an antibiotic-resistance gene, whereas in 10*cpsEF*, parts of the *cps2E* and *cps2F* genes were replaced. Both mutant strains seemed to be completely unencapsulated. Because the *cps2* genes seemed to be part of an operon, polar effects cannot be excluded. Therefore, these data did not give any information about the role of Cps2B, Cps2E or Cps2F in the polysaccharide biosynthesis. However, the results clearly show that the capsular polysaccharide of *S. suis* type 2 is a surface component with antiphagocytic activity. *In vitro* wild-type encapsulated bacteria are ingested by phagocytes at a very low frequency, whereas the mutant unencapsulated bacteria are efficiently ingested by porcine macrophages. Within 2 hours, over 99.6% of mutant bacteria were ingested and over 92% of the ingested bacteria were killed. Intracellularly, wild-type as well as mutant strains seemed to be killed with the same efficiency. This suggests that the loss of

capsular material is associated with loss of capacity to resist uptake by macrophages. This loss of resistance to *in vitro* phagocytosis was associated with a substantial attenuation of the virulence in germfree pigs. All pigs inoculated with the mutant strains survived the experiment and did not show any specific clinical signs of disease. Only some aspecific clinical signs of disease could be observed. Moreover, mutant bacteria could be reisolated from the pigs. This supports the idea that, as in other pathogenic Streptococci, the capsule of *S. suis* acts as an important virulence factor. Transposon mutants prepared by Charland impaired in the capsule production showed a reduced virulence in pigs and mice. To construct these mutants, the type 2 reference strain S735 was used. We previously showed that this strain is only weakly virulent for young pigs. Moreover, the insertion site of the transposon is unsolved so far.

**As a further example herein a rapid PCT test for *Streptococcus suis* type 7 is described.**

[00186] Recent epidemiological studies on *Streptococcus suis* infections in pigs indicated that, besides serotypes 1, 2 and 9, serotype 7 is also frequently associated with diseased animals. For the latter serotype, however, no rapid and sensitive diagnostic methods are available. This hampers prevention and control programs. Here we describe the development of a type-specific PCR test for the rapid and sensitive detection of *S. suis* serotype 7. The test is based on DNA sequences of capsular (cps) genes specific for serotype 7. These sequences could be identified by cross-hybridization of several individual cps genes with the chromosomal DNAs of 35 different *S. suis* serotypes.

[00187] *Streptococcus suis* is an important cause of meningitis, septicemia, arthritis and sudden death in young pigs [69, 70]. It can, however, also cause meningitis in man (71). Attempts to control the disease are still hampered by the lack of sufficient knowledge about the epidemiology of the disease and the lack of effective vaccines and sensitive diagnostics.

[00188] *S. suis* strains can be identified and classified by their morphological, biochemical and serological characteristics (70, 73, 74). Serological classification is based on the presence of specific antigenic determinants. Isolated and biochemically characterized *S. suis* cells are agglutinated with a panel of specific sera. These typing methods are very laborious and

time-consuming and can only be performed on isolated colonies. Moreover, it has been reported that nonspecific cross-reactions may occur among different types of *S. suis* (75, 76).

[00189] So far, 35 different serotypes have been described (7, 78, 79). *S. suis* serotype 2 is the most prevalent type isolated from diseased pigs, followed by serotypes 9 and 1. However, recently, serotype 7 strains were also frequently isolated from diseased pigs (80, 81, 82). This suggests that infections with *S. suis* serotype 7 strains seemed to be an increasing problem. Moreover, the virulence of *S. suis* serotype 7 strains was confirmed by experimental infection of young pigs (83).

[00190] Recently, rapid and sensitive PCR assays specific for serotypes 2 (and 1/2), 1 (and 14) and 9 were developed (84). These assays were based on the *cps* loci of *S. suis* serotypes 2, 1 and 9 (84, 85). However, until now, no rapid and sensitive diagnostic test was available for *S. suis* serotype 7. Herein we describe the development of a PCR test for the rapid and sensitive detection of *S. suis* serotype 7 strains. The test is based on DNA sequences which form a part of the *cps* locus of *S. suis* serotype 7. Compared with the serological serotyping methods, the PCR assay was a rapid, reliable and sensitive assay. Therefore, this test, in combination with the PCR tests which we previously developed for serotypes 1, 2 and 9, will undoubtedly contribute to a more rapid and reliable diagnosis of *S. suis* and may facilitate control and eradication programs.

## **Materials and Methods**

### **Bacterial strains, growth conditions and serotyping.**

[00191] The bacterial strains and plasmids used in this study are listed in Table 7. The *S. suis* reference strains were obtained from M. Gottschalk, Canada. *S. suis* strains were grown in Todd-Hewitt broth (code CM189, Oxoid), and plated on Columbia agar blood base (code CM331, Oxoid) containing 6% (v/v) horse blood. *E. coli* strains were grown in Luria broth (86) and plated on Luria broth containing 1.5% (w/v) agar. If required, ampicillin was added to the plates. The *S. suis* strains were serotyped by the slide agglutination test with serotype-specific antibodies (70).

## **DNA techniques.**

[00192] Routine DNA manipulations and PCR reactions were performed as described by Sambrook et al. (88). Blotting and hybridization were performed as described previously (84, 86).

## **DNA sequence analysis.**

[00193] DNA sequences were determined on a 373A DNA Sequencing system (Applied Biosystems, Warrington, GB). Samples were prepared by use of an ABI/PRISM dye terminator cycle sequencing ready reaction kit (Applied Biosystems). Custom-made sequencing primers were purchased from Life Technologies. Sequencing data were assembled and analyzed using the McMollyTetra program. The BLAST program was used to search for protein sequences homologous to the deduced amino acid sequences.

## **PCR.**

[00194] The primers used for the *cps7H* PCR correspond to the positions 3334-3354 and 3585-3565 in the *S. suis* *cps7* locus. The sequences were: 5'-AGCTCTAACACGAAATAAGGC-3' (SEQ ID NO:7) and 5'-GTCAAACACCCTGGATAGCCG-3' (SEQ ID NO:8).

[00195] The reaction mixtures contained 10 mM Tris-HCl, pH 8.3; 1.5 mM MgCl<sub>2</sub>; 50 mM KCl; 0.2 mM of each of the four deoxynucleotide triphosphates; 1 microM of each of the primers and 1U of AmpliTaq Gold DNA polymerase (Perkin Elmer Applied Biosystems, New Jersey). DNA amplification was carried out in a Perkin Elmer 9600 thermal cycler and the program consisted of an incubation for 10 min at 95°C and 30 cycles of 1 min at 95°C, 2 min at 56°C and 2 min at 72°C.

## **Results and discussion**

### **Cloning of the serotype 7-specific *cps* genes.**

[00196] To isolate the type-specific *cps* genes of *S. suis* serotype 7, we used the *cps9E* gene of serotype 9 as a probe to identify chromosomal DNA fragments of type 7 containing homologous DNA sequences (84). A 1.6-kb PstI fragment was identified and cloned in pKUN19. This yielded pCPS7-1 (FIG. 11, part C). In turn, this fragment was used as a probe to identify an overlapping 2.7

kb ScaI-ClaI fragment. pGEM7 containing the latter fragment was designated pCPS7-2 (FIG. 11, part C).

#### **Analysis of the cloned cps 7 genes.**

[00197] The complete nucleotide sequences of the inserts of pCPS7-1, pCPS7-2 were determined. Examination of the cps7 sequence revealed the presence of two complete and two incomplete open reading frames (ORFs) (FIG. 11, part C). All ORFs are preceded by a ribosome-binding site. In accord with the data obtained for the cps1, cps2 and cps9 genes of serotypes 1, 2 and 9, respectively, the type 7 ORFs are very closely linked to each other. The only significant intergenic gap was that found between cps7E and cps7F (443 nucleotides). No obvious promoter sequences or potential stem-loop structures were found in this region. This suggests that, as in serotypes 1, 2 and 9, the cps genes in serotype 7 form part of an operon.

[00198] An overview of the ORFs and their properties is shown in Table 8. As expected on the basis of the hybridization data (84), the Cps9E and Cps7E proteins showed a high similarity (identity 99%, Table 8). Based on sequence comparisons between Cps9E and Cps7E, the PstI fragment of pCPS7-1 lacks the region encoding the first 371 codons of Cps7E. The C-terminal part of the protein encoded by the cps7F gene showed some similarity with the BplG protein of *Bordetella pertussis* (88), as well as with the C-terminal part of *S. suis* Cps2E (85). Both BplG and Cps2E were suggested to have glycosyltransferase activity and are probably involved in the linkage of the first sugar to the lipid carrier (85, 88). The protein encoded by the cps7G gene showed similarity with the BlpF protein of *Bordetella pertussis* (88). BlpF is likely to be involved in the biosynthesis of an amino sugar, suggesting a similar function for Cps7G. The protein encoded by the cps7H gene showed similarity with the WbdN protein of *E. coli* (89) as well as with the N-terminal part of the Cps2K protein of *S. suis* (81). Both WbdN and Cps2K were suggested to have glycosyltransferase activity (85, 89).

#### **Serotype 7-specific cps genes.**

[00199] To determine whether the cloned fragments in pCPS7-1 and pCPS7-2 contained serotype 7-specific DNA sequences, cross-hybridization experiments were performed. DNA

fragments of the individual *cps7* genes were amplified by PCR, labeled with <sup>32</sup>P, and used to probe spot blots of chromosomal DNA of the reference strains of 35 different *S. suis* serotypes. The results are summarized in Table 9. As expected, based on the data obtained with the *cps9E* probe (84), the *cps7E* probe hybridized with chromosomal DNA of many different *S. suis* serotypes. The *cps7F* and *cps7G* probes showed hybridization with chromosomal DNA of *S. suis* serotypes 4, 5, 7, 17, and 23. However, the *cps7H* probe hybridized with chromosomal DNA of serotype 7 only, indicating that this gene is specific for serotype 7.

### **Type-specific PCR.**

[00200] We tested whether we could use PCR instead of hybridization for the typing of the *S. suis* serotype 7 strains. For that purpose, we selected an oligonucleotide primer set within the *cps7H* gene with which an amplified fragment of 251-bp was expected. In addition, we included in our analysis several *S. suis* serotype 7 strains, other than the reference strain. These strains were obtained from different countries and were isolated from different organs (Table 7). The results show that indeed a fragment of about 250-bp was amplified with all type 7 strains used (FIG. 12, part B), whereas no PCR products were obtained with serotype 1, 2 and 9 strains (FIG. 12, part A). This suggests that the PCR test, as described here, is a rapid diagnostic tool for the identification of *S. suis* serotype 7 strains. Until now, such a diagnostic test was not available for serotype 7 strains. Together with the recently developed PCR assays for serotypes 1, 2, 1/2, 14 and 9, this assay may be an important diagnostic tool to detect pigs carrying serotype 2, 1/2, 1, 14, 9 and 7 strains and may facilitate control and eradication programs.



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 aacaattttt tttcttgtgt cttgctttta taccgatcta ttttaagtga tcgagaattg 3660  
 gtagtttatc gctagcaata ttaattatat gcttggtatg gagatatata ggtggaaaat 3720  
 ttgcttggtat aaaaaagcta atagtaatat ttgtaatact acttattatt ttaaatactg 3780  
 aattgcttta ccatgaaatt ttggctgttt ataattctag agaatcaagt aacgaagcta 3840  
 gatttattat ttatcaagga agtattgata aagtattaga aaacaatatt ttatttggat 3900  
 atggaatatc cgaatattca gttacgggaa cttggctcgg aagtcattca ggctatatat 3960  
 cattttttta taaatcagga atagttgggt tgattttact gatgttttct tttttttatg 4020

ttataaaaaa aagttatgga gttaatgggg aaacagcact attttatttt acatcattag 4080  
 ccatattttt catatatgaa acaatagatc cgattattat tatattagta ctattctttt 4140  
 cttcaatagg tatttggaat aatataaatt ttaaaaagga tatggagaca aaaaatgaat 4200  
 gatttaattt cagttattgt accaatttat aatgtccaag attatcttga taaatgtatt 4260  
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 actgatgatt ctgagaaaat ttgcttaaac tatatgaaga acgatggaag aattaaatat 4380  
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 atgcatgata atataactga gtataatgcc gatatagcag agatagattt ttgttttagta 4560  
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 gagactgtaa aagaattttt gtcaggatct aatatagaaa ataatgtttg gtgcaagctt 4680  
 tattcacgag atattataaa agatataaaa ttccaaatta ataatagaag tattggtgag 4740  
 gatttgcttt ttaatttgga ggtcttgaac aatgtaacac gtgtagtagt tgatactaga 4800  
 gaatattatt ataattatgt cattcgtaac agttcgctta ttaatcagaa attctctata 4860  
 aataatattg atttagtcac aagattggag aattaccctt ttaagttaaa aagagagttt 4920  
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 aaagagtaaa aatttttttc actaatcata gtggagtatc aaatgctaga aatcatggaa 5400  
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 gattagtaga aaaattatat tttaatatta taaaaagtag aagtgattta tctggttggt 5520  
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 aaaaagaact atttgaagat tttcgatttg aaaagggtaa gattcatgaa gatgaatact 6720  
 tcacttatcg cttgctctat gagttagaaa aagttgcaat agttaaggag tgcttgact 6780  
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 gcctactgga atttcaaaat gaacgaatgg acttctatga aagtagagga gataaagagc 6900  
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<210> 10

<211> 239

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> ORF2Z

<400> 10

Ser Leu Asp Ile Asp His Met Met Glu Val Met Glu Ala Ser Lys Ser  
1 5 10 15

Ala Ala Gly Ser Ala Cys Pro Ser Pro Gln Ala Tyr Gln Ala Ala Phe  
20 25 30

Glu Gly Ala Glu Asn Ile Ile Val Val Thr Ile Thr Gly Gly Leu Ser  
35 40 45

Gly Ser Phe Asn Ala Ala Arg Val Ala Arg Asp Met Tyr Ile Glu Glu  
50 55 60

His Pro Asn Val Asn Ile His Leu Ile Asp Ser Leu Ser Ala Ser Gly  
65 70 75 80

Glu Met Asp Leu Leu Val His Gln Ile Asn Arg Leu Ile Ser Ala Gly  
85 90 95

Leu Asp Phe Pro Gln Val Val Glu Ala Ile Thr His Tyr Arg Glu His  
100 105 110

Ser Lys Leu Leu Phe Val Leu Ala Lys Val Asp Asn Leu Val Lys Asn  
115 120 125

Gly Arg Leu Ser Lys Leu Val Gly Thr Val Val Gly Leu Leu Asn Ile  
130 135 140

Arg Met Val Gly Glu Ala Ser Ala Glu Gly Lys Leu Glu Leu Leu Gln  
145 150 155 160

Lys Ala Arg Gly His Lys Lys Ser Val Thr Ala Ala Phe Glu Glu Met  
165 170 175

Lys Lys Ala Gly Tyr Asp Gly Gly Arg Ile Val Met Ala His Arg Asn  
180 185 190

Asn Ala Lys Phe Phe Gln Gln Phe Ser Glu Leu Val Lys Ala Ser Phe  
195 200 205

Pro Thr Ala Val Ile Asp Glu Val Ala Thr Ser Gly Leu Cys Ser Phe  
210 215 220

Tyr Ala Glu Glu Gly Gly Leu Leu Met Gly Tyr Glu Val Lys Ala  
225 230 235

<210> 11

<211> 244

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> ORF2X

<400> 11

Met Lys Ile Ile Ile Pro Asn Ala Lys Glu Val Asn Thr Asn Leu Glu  
1 5 10 15

Asn Ala Ser Phe Tyr Leu Leu Ser Asp Arg Ser Lys Pro Val Leu Asp  
20 25 30

Ala Ile Ser Gln Phe Asp Val Lys Lys Met Ala Ala Phe Tyr Lys Leu  
35 40 45

Asn Glu Ala Lys Ala Glu Leu Glu Ala Asp Arg Trp Tyr Arg Ile Arg  
50 55 60

Thr Gly Gln Ala Lys Thr Tyr Pro Ala Trp Gln Leu Tyr Asp Gly Leu  
65 70 75 80

Met Tyr Arg Tyr Met Asp Arg Arg Gly Ile Asp Ser Lys Glu Glu Asn  
85 90 95

Tyr Leu Arg Asp His Val Arg Val Ala Thr Ala Leu Tyr Gly Leu Ile  
100 105 110

His Pro Phe Glu Phe Ile Ser Pro His Arg Leu Asp Phe Gln Gly Ser  
115 120 125

Leu Lys Ile Gly Asn Gln Ser Leu Lys Gln Tyr Trp Arg Pro Tyr Tyr  
130 135 140

Asp Gln Glu Val Gly Asp Asp Glu Leu Ile Leu Ser Leu Ala Ser Ser  
 145 150 155 160  
 Glu Phe Glu Gln Val Phe Ser Pro Gln Ile Gln Lys Arg Leu Val Lys  
 165 170 175  
 Ile Leu Phe Met Glu Glu Lys Ala Gly Gln Leu Lys Val His Ser Thr  
 180 185 190  
 Ile Ser Lys Lys Gly Arg Gly Arg Leu Leu Ser Trp Leu Ala Lys Asn  
 195 200 205  
 Asn Ile Gln Glu Leu Ser Asp Ile Gln Asp Phe Lys Val Asp Gly Phe  
 210 215 220  
 Glu Tyr Cys Thr Ser Glu Ser Thr Ala Asn Gln Leu Thr Phe Ile Arg  
 225 230 235 240  
 Ser Ile Lys Met

<210> 12

<211> 481

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS2A

<400> 12

Met Lys Lys Arg Ser Gly Arg Ser Lys Ser Ser Lys Phe Lys Leu Val  
 1 5 10 15  
 Asn Phe Ala Leu Leu Gly Leu Tyr Ser Ile Thr Leu Cys Leu Phe Leu  
 20 25 30  
 Val Thr Met Tyr Arg Tyr Asn Ile Leu Asp Phe Arg Tyr Leu Asn Tyr  
 35 40 45  
 Ile Val Thr Leu Leu Leu Val Gly Val Ala Val Leu Ala Gly Leu Leu  
 50 55 60

Met	Trp	Arg	Lys	Lys	Ala	Arg	Ile	Phe	Thr	Ala	Leu	Leu	Leu	Val	Phe	65	70	75	80
Ser	Leu	Val	Ile	Thr	Ser	Val	Gly	Ile	Tyr	Gly	Met	Gln	Glu	Val	Val	85	90	95	
Lys	Phe	Ser	Thr	Arg	Leu	Asn	Ser	Asn	Ser	Thr	Phe	Ser	Glu	Tyr	Glu	100	105	110	
Met	Ser	Ile	Leu	Val	Pro	Ala	Asn	Ser	Asp	Ile	Thr	Asp	Val	Arg	Gln	115	120	125	
Leu	Thr	Ser	Ile	Leu	Ala	Pro	Ala	Glu	Tyr	Asp	Gln	Asp	Asn	Ile	Thr	130	135	140	
Ala	Leu	Leu	Asp	Asp	Ile	Ser	Lys	Met	Glu	Ser	Thr	Gln	Leu	Ala	Thr	145	150	155	160
Ser	Pro	Gly	Thr	Ser	Tyr	Leu	Thr	Ala	Tyr	Gln	Ser	Met	Leu	Asn	Gly	165	170	175	
Glu	Ser	Gln	Ala	Met	Val	Phe	Asn	Gly	Val	Phe	Thr	Asn	Ile	Leu	Glu	180	185	190	
Asn	Glu	Asp	Pro	Gly	Phe	Ser	Ser	Lys	Val	Lys	Lys	Ile	Tyr	Ser	Phe	195	200	205	
Lys	Val	Thr	Gln	Thr	Val	Glu	Thr	Ala	Thr	Lys	Gln	Val	Ser	Gly	Asp	210	215	220	
Ser	Phe	Asn	Ile	Tyr	Ile	Ser	Gly	Ile	Asp	Ala	Tyr	Gly	Pro	Ile	Ser	225	230	235	240
Thr	Val	Ser	Arg	Ser	Asp	Val	Asn	Ile	Ile	Met	Thr	Val	Asn	Arg	Ala	245	250	255	
Thr	His	Lys	Ile	Leu	Leu	Thr	Thr	Thr	Pro	Arg	Asp	Ser	Tyr	Val	Ala	260	265	270	
Phe	Ala	Asp	Gly	Gly	Gln	Asn	Gln	Tyr	Asp	Lys	Leu	Thr	His	Ala	Gly	275	280	285	
Ile	Tyr	Gly	Val	Asn	Ala	Ser	Val	His	Thr	Leu	Glu	Asn	Phe	Tyr	Gly	290	295	300	
Ile	Asp	Ile	Ser	Asn	Tyr	Val	Arg	Leu	Asn	Phe	Ile	Ser	Phe	Leu	Gln	305	310	315	320
Leu	Ile	Asp	Leu	Val	Gly	Gly	Ile	Asp	Val	Tyr	Asn	Asp	Gln	Glu	Phe	325	330	335	

Thr Ser Leu His Gly Asn Tyr His Phe Pro Val Gly Gln Val His Leu  
 340 345 350  
 Asn Ser Asp Gln Ala Leu Gly Phe Val Arg Glu Arg Tyr Ser Leu Thr  
 355 360 365  
 Gly Gly Asp Asn Asp Arg Gly Lys Asn Gln Glu Lys Val Ile Ala Ala  
 370 375 380  
 Leu Ile Lys Lys Met Ser Thr Pro Glu Asn Leu Lys Asn Tyr Gln Ala  
 385 390 395 400  
 Ile Leu Ser Gly Leu Glu Gly Ser Ile Gln Thr Asp Leu Ser Leu Glu  
 405 410 415  
 Thr Ile Met Ser Leu Val Asn Thr Gln Leu Glu Ser Gly Thr Gln Phe  
 420 425 430  
 Thr Val Glu Ser Gln Ala Leu Thr Gly Thr Gly Arg Ser Asp Leu Ser  
 435 440 445  
 Ser Tyr Ala Met Pro Gly Ser Gln Leu Tyr Met Met Glu Ile Asn Gln  
 450 455 460  
 Asp Ser Leu Glu Gln Ser Lys Ala Ala Ile Gln Ser Val Leu Val Glu  
 465 470 475 480

Lys

<210> 13

<211> 229

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS2B

<400> 13

Met Asn Asn Gln Glu Val Asn Ala Ile Glu Ile Asp Val Leu Phe Leu  
 1 5 10 15  
 Leu Lys Thr Ile Trp Arg Lys Lys Phe Leu Ile Leu Leu Thr Ala Val  
 20 25 30



<223> CPS2C

<400> 14

Met	Ala	Met	Leu	Glu	Ile	Ala	Arg	Thr	Lys	Arg	Glu	Gly	Val	Asn	Lys	
1				5					10					15		
Thr	Glu	Glu	Tyr	Phe	Asn	Ala	Ile	Arg	Thr	Asn	Ile	Gln	Leu	Ser	Gly	
			20					25					30			
Ala	Asp	Ile	Lys	Val	Val	Gly	Ile	Thr	Ser	Val	Lys	Ser	Asn	Glu	Gly	
		35					40					45				
Lys	Ser	Thr	Thr	Ala	Ala	Ser	Leu	Ala	Ile	Ala	Tyr	Ala	Arg	Ser	Gly	
	50					55					60					
Tyr	Lys	Thr	Val	Leu	Val	Asp	Ala	Asp	Ile	Arg	Asn	Ser	Val	Met	Pro	
65					70					75					80	
Gly	Phe	Phe	Lys	Pro	Ile	Thr	Lys	Ile	Thr	Gly	Leu	Thr	Asp	Tyr	Leu	
				85					90					95		
Ala	Gly	Thr	Thr	Asp	Leu	Ser	Gln	Gly	Leu	Cys	Asp	Thr	Asp	Ile	Pro	
			100					105					110			
Asn	Leu	Thr	Val	Ile	Glu	Ser	Gly	Lys	Val	Ser	Pro	Asn	Pro	Thr	Ala	
		115					120					125				
Leu	Leu	Gln	Ser	Lys	Asn	Phe	Glu	Asn	Leu	Leu	Ala	Thr	Leu	Arg	Arg	
	130					135					140					
Tyr	Tyr	Asp	Tyr	Val	Ile	Val	Asp	Cys	Pro	Pro	Leu	Gly	Leu	Val	Ile	
145					150					155					160	
Asp	Ala	Ala	Ile	Ile	Ala	Gln	Lys	Cys	Asp	Ala	Met	Val	Ala	Val	Val	
				165					170					175		
Glu	Ala	Gly	Asn	Val	Lys	Cys	Ser	Ser	Leu	Lys	Lys	Val	Lys	Glu	Gln	
			180					185					190			
Leu	Glu	Gln	Thr	Gly	Thr	Pro	Phe	Leu	Gly	Val	Ile	Leu	Asn	Lys	Tyr	
		195					200					205				
Asp	Ile	Ala	Thr	Glu	Lys	Tyr	Ser	Glu	Tyr	Gly	Asn	Tyr	Gly	Lys	Lys	
	210					215					220					
Ala																
225																

<210> 15

<211> 243

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS2D

<400> 15

Met Ile Asp Ile His Ser His Ile Ile Phe Gly Val Asp Asp Gly Pro  
1 5 10 15

Lys Thr Ile Glu Glu Ser Leu Ser Leu Ile Ser Glu Ala Tyr Arg Gln  
20 25 30

Gly Val Arg Tyr Ile Val Ala Thr Ser His Arg Arg Lys Gly Met Phe  
35 40 45

Glu Thr Pro Glu Lys Ile Ile Met Ile Asn Phe Leu Gln Leu Lys Glu  
50 55 60

Ala Val Ala Glu Val Tyr Pro Glu Ile Arg Leu Cys Tyr Gly Ala Glu  
65 70 75 80

Leu Tyr Tyr Ser Lys Asp Ile Leu Ser Lys Leu Glu Lys Lys Lys Val  
85 90 95

Pro Thr Leu Asn Gly Ser Cys Tyr Ile Leu Leu Glu Phe Ser Thr Asp  
100 105 110

Thr Pro Trp Lys Glu Ile Gln Glu Ala Val Asn Glu Met Thr Leu Leu  
115 120 125

Gly Leu Thr Pro Val Leu Ala His Ile Glu Arg Tyr Asp Ala Leu Ala  
130 135 140

Phe Gln Ser Glu Arg Val Glu Lys Leu Ile Asp Lys Gly Cys Tyr Thr  
145 150 155 160

Gln Val Asn Ser Asn His Val Leu Lys Pro Ala Leu Ile Gly Glu Arg  
165 170 175

Ala Lys Glu Phe Lys Lys Arg Thr Arg Tyr Phe Leu Glu Gln Asp Leu  
180 185 190



Val	His	Cys	Val	Ala	Ser	Asp	Met	His	Asn	Leu	Tyr	Ser	Arg	Pro	Pro
		195					200					205			
Phe	Met	Arg	Glu	Ala	Tyr	Gln	Leu	Val	Lys	Lys	Glu	Tyr	Gly	Glu	Asp
	210					215					220				
Arg	Ala	Lys	Ala	Leu	Phe	Lys	Lys	Asn	Pro	Leu	Leu	Ile	Leu	Lys	Asn
225					230					235					240
Gln	Val	Gln													

<210> 16

<211> 459

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS2E

<400> 16

Met	Asn	Ile	Glu	Ile	Gly	Tyr	Arg	Gln	Thr	Lys	Leu	Ala	Leu	Phe	Asp
1				5					10					15	
Met	Ile	Ala	Val	Thr	Ile	Ser	Ala	Ile	Leu	Thr	Ser	His	Ile	Pro	Asn
			20					25					30		
Ala	Asp	Leu	Asn	Arg	Ser	Gly	Ile	Phe	Ile	Ile	Met	Met	Val	His	Tyr
		35					40					45			
Phe	Ala	Phe	Phe	Ile	Ser	Arg	Met	Pro	Val	Glu	Phe	Glu	Tyr	Arg	Gly
	50					55					60				
Asn	Leu	Ile	Glu	Phe	Glu	Lys	Thr	Phe	Asn	Tyr	Ser	Ile	Ile	Phe	Val
65					70					75					80
Ile	Phe	Leu	Met	Ala	Val	Ser	Phe	Met	Leu	Glu	Asn	Asn	Phe	Ala	Leu
				85					90					95	
Ser	Arg	Arg	Gly	Ala	Val	Tyr	Phe	Thr	Leu	Ile	Asn	Phe	Val	Leu	Val
			100					105					110		
Tyr	Leu	Phe	Asn	Val	Ile	Ile	Lys	Gln	Phe	Lys	Asp	Ser	Phe	Leu	Phe
		115					120					125			

Ser	Thr	Thr	Tyr	Gln	Lys	Lys	Thr	Ile	Leu	Ile	Thr	Thr	Ala	Glu	Leu			
	130					135					140							
Trp	Glu	Asn	Met	Gln	Val	Leu	Phe	Glu	Ser	Asp	Ile	Leu	Phe	Gln	Lys			
145					150					155					160			
Asn	Leu	Val	Ala	Leu	Val	Ile	Leu	Gly	Thr	Glu	Ile	Asp	Lys	Ile	Asn			
				165					170					175				
Leu	Pro	Leu	Pro	Leu	Tyr	Tyr	Ser	Val	Glu	Glu	Ala	Ile	Gly	Phe	Ser			
			180					185					190					
Thr	Arg	Glu	Val	Val	Asp	Tyr	Val	Phe	Ile	Asn	Leu	Pro	Ser	Glu	Tyr			
		195					200					205						
Phe	Asp	Leu	Lys	Gln	Leu	Val	Ser	Asp	Phe	Glu	Leu	Leu	Gly	Ile	Asp			
	210					215					220							
Val	Gly	Val	Asp	Ile	Asn	Ser	Phe	Gly	Phe	Thr	Val	Leu	Lys	Asn	Lys			
225					230					235					240			
Lys	Ile	Gln	Met	Leu	Gly	Asp	His	Ser	Ile	Val	Thr	Phe	Ser	Thr	Asn			
				245					250					255				
Phe	Tyr	Lys	Pro	Ser	His	Ile	Trp	Met	Lys	Arg	Leu	Leu	Asp	Ile	Leu			
			260					265					270					
Gly	Ala	Val	Val	Gly	Leu	Ile	Ile	Ser	Gly	Ile	Val	Ser	Ile	Leu	Leu			
		275					280					285						
Ile	Pro	Ile	Ile	Arg	Arg	Asp	Gly	Gly	Pro	Ala	Ile	Phe	Ala	Gln	Lys			
	290					295					300							
Arg	Val	Gly	Gln	Asn	Gly	Arg	Ile	Phe	Thr	Phe	Tyr	Lys	Phe	Arg	Ser			
305					310					315					320			
Met	Phe	Val	Asp	Ala	Glu	Val	Arg	Lys	Lys	Glu	Leu	Met	Ala	Gln	Asn			
				325					330					335				
Gln	Met	Gln	Gly	Gly	Met	Phe	Lys	Met	Asp	Asn	Asp	Pro	Arg	Ile	Thr			
			340					345					350					
Pro	Ile	Gly	His	Phe	Ile	Arg	Lys	Thr	Ser	Leu	Asp	Glu	Leu	Pro	Gln			
		355					360					365						
Phe	Tyr	Asn	Val	Leu	Ile	Gly	Asp	Met	Ser	Leu	Val	Gly	Thr	Arg	Pro			
	370					375					380							
Pro	Thr	Val	Asp	Glu	Phe	Glu	Lys	Tyr	Thr	Pro	Ser	Gln	Lys	Arg	Arg			
385					390					395					400			

Leu Ser Phe Lys Pro Gly Ile Thr Gly Leu Trp Gln Val Ser Gly Arg  
                     405                    410                    415  
 Ser Asp Ile Thr Asp Phe Asn Glu Val Val Arg Leu Asp Leu Thr Tyr  
                     420                    425                    430  
 Ile Asp Asn Trp Thr Ile Trp Ser Asp Ile Lys Ile Leu Leu Lys Thr  
                     435                    440                    445  
 Val Lys Val Val Leu Leu Arg Glu Gly Gly Gln  
             450                    455

<210> 17

<211> 389

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS2F

<400> 17

Met Arg Thr Val Tyr Ile Ile Gly Ser Lys Gly Ile Pro Ala Lys Tyr  
 1                    5                    10                    15  
 Gly Gly Phe Glu Thr Phe Val Glu Lys Leu Thr Glu Tyr Gln Lys Asp  
                     20                    25                    30  
 Lys Ser Ile Asn Tyr Phe Val Ala Cys Thr Arg Glu Asn Ser Ala Lys  
                     35                    40                    45  
 Ser Asp Ile Thr Gly Glu Val Phe Glu His Asn Gly Ala Thr Cys Phe  
                     50                    55                    60  
 Asn Ile Asp Val Pro Asn Ile Gly Ser Ala Lys Ala Ile Leu Tyr Asp  
 65                    70                    75                    80  
 Ile Met Ala Leu Lys Lys Ser Ile Glu Ile Ala Lys Asp Arg Asn Asp  
                     85                    90                    95  
 Thr Ser Pro Ile Phe Tyr Ile Leu Ala Cys Arg Ile Gly Pro Phe Ile  
                     100                    105                    110  
 Tyr Leu Phe Lys Lys Gln Ile Glu Ser Ile Gly Gly Gln Leu Phe Val  
                     115                    120                    125

Asn	Pro	Asp	Gly	His	Glu	Trp	Leu	Arg	Glu	Lys	Trp	Ser	Tyr	Pro	Val	
	130					135					140					
Arg	Gln	Tyr	Trp	Lys	Phe	Ser	Glu	Ser	Leu	Met	Leu	Lys	Tyr	Ala	Asp	
145					150					155					160	
Leu	Leu	Ile	Cys	Asp	Ser	Lys	Asn	Ile	Glu	Lys	Tyr	Ile	His	Glu	Asp	
				165					170					175		
Tyr	Arg	Lys	Tyr	Ala	Pro	Glu	Thr	Ser	Tyr	Ile	Ala	Tyr	Gly	Thr	Asp	
			180					185					190			
Leu	Asp	Lys	Ser	Arg	Leu	Ser	Pro	Thr	Asp	Ser	Val	Val	Arg	Glu	Trp	
	195						200				205					
Tyr	Lys	Glu	Lys	Glu	Ile	Ser	Glu	Asn	Asp	Tyr	Tyr	Leu	Val	Val	Gly	
210						215					220					
Arg	Phe	Val	Pro	Glu	Asn	Asn	Tyr	Glu	Val	Met	Ile	Arg	Glu	Phe	Met	
225					230					235					240	
Lys	Ser	Tyr	Ser	Arg	Lys	Asp	Phe	Val	Leu	Ile	Thr	Asn	Val	Glu	His	
				245					250					255		
Asn	Ser	Phe	Tyr	Glu	Lys	Leu	Lys	Lys	Glu	Thr	Gly	Phe	Asp	Lys	Asp	
			260					265					270			
Lys	Arg	Ile	Lys	Phe	Val	Gly	Thr	Val	Tyr	Asn	Gln	Glu	Leu	Leu	Lys	
	275						280					285				
Tyr	Ile	Arg	Glu	Asn	Ala	Phe	Ala	Tyr	Phe	His	Gly	His	Glu	Val	Gly	
290					295						300					
Gly	Thr	Asn	Pro	Ser	Leu	Leu	Glu	Ala	Leu	Ser	Ser	Thr	Lys	Leu	Asn	
305					310					315					320	
Leu	Leu	Leu	Asp	Val	Gly	Phe	Asn	Arg	Glu	Val	Gly	Glu	Glu	Gly	Ala	
				325					330					335		
Lys	Tyr	Trp	Asn	Lys	Asp	Asn	Leu	His	Arg	Val	Ile	Asp	Ser	Cys	Glu	
			340				345						350			
Gln	Leu	Ser	Gln	Glu	Gln	Ile	Asn	Asp	Met	Asp	Ser	Leu	Ser	Thr	Lys	
	355						360					365				
Gln	Val	Lys	Glu	Arg	Phe	Ser	Trp	Asp	Phe	Ile	Val	Asp	Glu	Tyr	Glu	
370						375					380					
Lys	Leu	Phe	Lys	Gly												
385																

<210> 18

<211> 385

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS2G

<400> 18

Met Lys Lys Ile Leu Tyr Leu His Ala Gly Ala Glu Leu Tyr Gly Ala  
1 5 10 15

Asp Lys Val Leu Leu Glu Leu Ile Lys Gly Leu Asp Lys Asn Glu Phe  
20 25 30

Glu Ala His Val Ile Leu Pro Asn Asp Gly Val Leu Val Pro Ala Leu  
35 40 45

Arg Glu Val Gly Ala Gln Val Glu Val Ile Asn Tyr Pro Ile Leu Arg  
50 55 60

Arg Lys Tyr Phe Asn Pro Lys Gly Ile Phe Asp Tyr Phe Ile Ser Tyr  
65 70 75 80

His His Tyr Ser Lys Gln Ile Ala Gln Tyr Ala Ile Glu Asn Lys Val  
85 90 95

Asp Ile Ile His Asn Asn Thr Thr Ala Val Leu Glu Gly Ile Tyr Leu  
100 105 110

Lys Arg Lys Leu Lys Leu Pro Leu Leu Trp His Val His Glu Ile Ile  
115 120 125

Val Lys Pro Lys Phe Ile Ser Asp Ser Ile Asn Phe Leu Met Gly Arg  
130 135 140

Phe Ala Asp Lys Ile Val Thr Val Ser Gln Ala Val Ala Asn His Ile  
145 150 155 160

Lys Gln Ser Pro His Ile Lys Asp Asp Gln Ile Ser Val Ile Tyr Asn  
165 170 175

Gly Val Asp Asn Lys Val Phe Tyr Gln Ser Asp Ala Arg Ser Val Arg  
180 185 190



<223> cps2h

<400> 19

Met	Lys	Ile	Ile	Ser	Phe	Thr	Met	Val	Asn	Asn	Glu	Ser	Glu	Ile	Ile	
1				5					10					15		
Glu	Ser	Phe	Ile	Arg	Tyr	Asn	Tyr	Asn	Phe	Ile	Asp	Glu	Met	Val	Ile	
			20					25					30			
Ile	Asp	Asn	Gly	Cys	Thr	Asp	Asn	Thr	Met	Gln	Ile	Ile	Phe	Asn	Leu	
		35					40					45				
Ile	Lys	Glu	Gly	Tyr	Lys	Ile	Ser	Val	Tyr	Asp	Glu	Ser	Leu	Glu	Ala	
	50					55					60					
Tyr	Asn	Gln	Tyr	Arg	Leu	Asp	Asn	Lys	Tyr	Leu	Thr	Lys	Ile	Ile	Ala	
65					70					75					80	
Glu	Lys	Asn	Pro	Asp	Leu	Ile	Ile	Pro	Leu	Asp	Ala	Asp	Glu	Phe	Leu	
				85					90					95		
Thr	Ala	Asp	Ser	Asn	Pro	Arg	Lys	Leu	Leu	Glu	Gln	Leu	Asp	Leu	Glu	
			100					105					110			
Lys	Ile	His	Tyr	Val	Asn	Trp	Gln	Trp	Phe	Val	Met	Thr	Lys	Lys	Asp	
		115					120					125				
Asp	Ile	Asn	Asp	Ser	Phe	Ile	Pro	Arg	Arg	Met	Gln	Tyr	Cys	Phe	Glu	
	130					135					140					
Lys	Pro	Val	Trp	His	His	Ser	Asp	Gly	Lys	Pro	Val	Thr	Lys	Cys	Ile	
145					150					155					160	
Ile	Ser	Ala	Lys	Tyr	Tyr	Lys	Lys	Met	Asn	Leu	Lys	Leu	Ser	Met	Gly	
				165					170					175		
His	His	Thr	Val	Phe	Gly	Asn	Pro	Asn	Val	Arg	Ile	Glu	His	His	Asn	
			180					185					190			
Asp	Leu	Lys	Phe	Ala	His	Tyr	Arg	Ala	Ile	Ser	Gln	Glu	Gln	Leu	Ile	
		195					200					205				
Tyr	Lys	Thr	Ile	Cys	Tyr	Thr	Ile	Arg	Asp	Ile	Ala	Thr	Met	Glu	Asn	
	210					215					220					
Asn	Ile	Glu	Thr	Ala	Gln	Arg	Thr	Asn	Gln	Met	Ala	Leu	Ile	Glu	Ser	
225					230					235					240	
Gly	Val	Asp	Met	Trp	Glu	Thr	Ala	Arg	Glu	Ala	Ser	Tyr	Ser	Gly	Tyr	
				245					250					255		

Asp	Cys	Asn	Val	Ile	His	Ala	Pro	Ile	Asp	Leu	Ser	Phe	Cys	Lys	Glu	
			260					265					270			
Asn	Ile	Val	Ile	Lys	Tyr	Asn	Glu	Leu	Ser	Arg	Glu	Thr	Val	Ala	Glu	
		275					280					285				
Arg	Val	Met	Lys	Thr	Gly	Arg	Glu	Met	Ala	Val	Arg	Ala	Tyr	Asn	Val	
	290					295					300					
Glu	Arg	Lys	Gln	Lys	Glu	Lys	Lys	Phe	Leu	Lys	Pro	Ile	Ile	Phe	Val	
305					310					315					320	
Leu	Asp	Gly	Leu	Lys	Gly	Asp	Glu	Tyr	Ile	His	Pro	Asn	Pro	Ser	Asn	
			325						330					335		
His	Leu	Thr	Ile	Leu	Thr	Glu	Met	Tyr	Asn	Val	Arg	Gly	Leu	Leu	Thr	
			340					345					350			
Asp	Asn	His	Gln	Ile	Lys	Phe	Leu	Lys	Val	Asn	Tyr	Arg	Leu	Ile	Ile	
		355					360					365				
Thr	Pro	Asp	Phe	Ala	Lys	Phe	Leu	Pro	His	Glu	Phe	Ile	Val	Val	Pro	
	370					375					380					
Asp	Thr	Leu	Asp	Ile	Glu	Gln	Val	Lys	Ser	Gln	Tyr	Val	Gly	Thr	Gly	
385					390					395					400	
Val	Asp	Leu	Ser	Lys	Ile	Ile	Ser	Leu	Lys	Glu	Tyr	Arg	Lys	Glu	Ile	
				405					410					415		
Gly	Phe	Ile	Gly	Asn	Leu	Tyr	Ala	Leu	Leu	Gly	Phe	Val	Pro	Asn	Met	
			420					425					430			
Leu	Asn	Arg	Ile	Tyr	Leu	Tyr	Ile	Gln	Arg	Asn	Gly	Ile	Ala	Asn	Thr	
		435					440					445				
Ile	Ile	Lys	Ile	Lys	Ser	Arg	Leu									
	450					455										

<210> 20

<211> 410

<212> PRT

<213> Streptococcus suis.

<220>

<221> misc\_feature



<223> CPS2I

<400> 20

Met	Gln	Ala	Asp	Arg	Arg	Lys	Thr	Phe	Gly	Lys	Met	Arg	Ile	Arg	Ile	
1				5					10					15		
Asn	Asn	Leu	Phe	Phe	Val	Ala	Ile	Ala	Phe	Met	Gly	Ile	Ile	Ile	Ser	
			20					25					30			
Asn	Ser	Gln	Val	Val	Leu	Ala	Ile	Gly	Lys	Ala	Ser	Val	Ile	Gln	Tyr	
		35					40					45				
Leu	Ser	Tyr	Leu	Val	Leu	Ile	Leu	Cys	Ile	Val	Asn	Asp	Leu	Leu	Lys	
	50					55					60					
Asn	Asn	Lys	His	Ile	Val	Val	Tyr	Lys	Leu	Gly	Tyr	Leu	Phe	Leu	Ile	
65					70					75					80	
Ile	Phe	Leu	Phe	Thr	Ile	Gly	Ile	Cys	Gln	Gln	Ile	Leu	Pro	Ile	Thr	
				85					90					95		
Thr	Lys	Ile	Tyr	Leu	Ser	Ile	Ser	Met	Met	Ile	Ile	Ser	Val	Leu	Ala	
			100					105					110			
Thr	Leu	Pro	Ile	Ser	Leu	Ile	Lys	Asp	Ile	Asp	Asp	Phe	Arg	Arg	Ile	
		115					120					125				
Ser	Asn	His	Leu	Leu	Phe	Ala	Leu	Phe	Ile	Thr	Ser	Ile	Leu	Gly	Ile	
	130					135					140					
Lys	Met	Gly	Ala	Thr	Met	Phe	Thr	Gly	Ala	Val	Glu	Gly	Ile	Gly	Phe	
145					150					155					160	
Ser	Gln	Gly	Phe	Asn	Gly	Gly	Leu	Thr	His	Lys	Asn	Phe	Phe	Gly	Ile	
				165					170					175		
Thr	Ile	Leu	Met	Gly	Phe	Val	Leu	Thr	Tyr	Leu	Ala	Tyr	Lys	Tyr	Gly	
		180						185					190			
Ser	Tyr	Lys	Arg	Thr	Asp	Arg	Phe	Ile	Leu	Gly	Leu	Glu	Leu	Phe	Leu	
		195					200					205				
Ile	Leu	Ile	Ser	Asn	Thr	Arg	Ser	Val	Tyr	Leu	Ile	Leu	Leu	Leu	Phe	
	210					215					220					
Leu	Phe	Leu	Val	Asn	Leu	Asp	Lys	Ile	Lys	Ile	Glu	Gln	Arg	Gln	Trp	
225				230						235					240	
Ser	Thr	Leu	Lys	Tyr	Ile	Ser	Met	Leu	Phe	Cys	Ala	Ile	Phe	Leu	Tyr	
				245					250					255		

Tyr Phe Phe Gly Phe Leu Ile Thr His Ser Asp Ser Tyr Ala His Arg  
                   260                                  265                                  270  
 Val Asn Gly Leu Ile Asn Phe Phe Glu Tyr Tyr Arg Asn Asp Trp Phe  
                   275                                  280                                  285  
 His Leu Met Phe Gly Ala Ala Asp Leu Ala Tyr Gly Asp Leu Thr Leu  
           290                                  295                                  300  
 Asp Tyr Ala Ile Arg Val Arg Arg Val Leu Gly Trp Asn Gly Thr Leu  
 305                                  310                                  315                                  320  
 Glu Met Pro Leu Leu Ser Ile Met Leu Lys Asn Gly Phe Ile Gly Leu  
                   325                                  330                                  335  
 Val Gly Tyr Gly Ile Val Leu Tyr Lys Leu Tyr Arg Asn Val Arg Ile  
                   340                                  345                                  350  
 Leu Lys Thr Asp Asn Ile Lys Thr Ile Gly Lys Ser Val Phe Ile Ile  
           355                                  360                                  365  
 Val Val Leu Ser Ala Thr Val Glu Asn Tyr Ile Val Asn Leu Ser Phe  
           370                                  375                                  380  
 Val Phe Met Pro Ile Cys Phe Cys Leu Leu Asn Ser Ile Ser Thr Met  
 385                                  390                                  395                                  400  
 Glu Ser Thr Ile Asn Lys Gln Leu Gln Thr  
                   405                                  410

<210> 21

<211> 332

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS2J

<400> 21

Met Glu Lys Val Ser Ile Ile Val Pro Ile Phe Asn Thr Glu Lys Tyr  
 1                  5                                  10                                  15  
 Leu Arg Glu Cys Leu Asp Ser Ile Ile Ser Gln Ser Tyr Thr Asn Leu  
           20                                  25                                  30

Glu	Ile	Leu	Leu	Ile	Asp	Asp	Gly	Ser	Ser	Asp	Ser	Ser	Thr	Asp	Ile	35	40	45
Cys	Leu	Glu	Tyr	Ala	Glu	Gln	Asp	Gly	Arg	Ile	Lys	Leu	Phe	Arg	Leu	50	55	60
Pro	Asn	Gly	Gly	Val	Ser	Asn	Ala	Arg	Asn	Tyr	Gly	Ile	Lys	Asn	Ser	65	70	75
Thr	Ala	Asn	Tyr	Ile	Met	Phe	Val	Asp	Ser	Asp	Asp	Ile	Val	Asp	Gly	85	90	95
Asn	Ile	Val	Glu	Ser	Leu	Tyr	Thr	Cys	Leu	Lys	Glu	Asn	Asp	Ser	Asp	100	105	110
Leu	Ser	Gly	Gly	Leu	Leu	Ala	Thr	Phe	Asp	Gly	Asn	Tyr	Gln	Glu	Ser	115	120	125
Glu	Leu	Gln	Lys	Cys	Gln	Ile	Asp	Leu	Glu	Glu	Ile	Lys	Glu	Val	Arg	130	135	140
Asp	Leu	Gly	Asn	Glu	Asn	Phe	Pro	Asn	His	Tyr	Met	Ser	Gly	Ile	Phe	145	150	155
Asn	Ser	Pro	Cys	Cys	Lys	Leu	Tyr	Lys	Asn	Ile	Tyr	Ile	Asn	Gln	Gly	165	170	175
Phe	Asp	Thr	Glu	Gln	Trp	Leu	Gly	Glu	Asp	Leu	Leu	Phe	Asn	Leu	Asn	180	185	190
Tyr	Leu	Lys	Asn	Ile	Lys	Lys	Val	Arg	Tyr	Val	Asn	Arg	Asn	Leu	Tyr	195	200	205
Phe	Ala	Arg	Arg	Ser	Leu	Gln	Ser	Thr	Thr	Asn	Thr	Phe	Lys	Tyr	Asp	210	215	220
Val	Phe	Ile	Gln	Leu	Glu	Asn	Leu	Glu	Glu	Lys	Thr	Phe	Asp	Leu	Phe	225	230	235
Val	Lys	Ile	Phe	Gly	Gly	Gln	Tyr	Glu	Phe	Ser	Val	Phe	Lys	Glu	Thr	245	250	255
Leu	Gln	Trp	His	Ile	Ile	Tyr	Tyr	Ser	Leu	Leu	Met	Phe	Lys	Asn	Gly	260	265	270
Asp	Glu	Ser	Leu	Pro	Lys	Lys	Leu	His	Ile	Phe	Lys	Tyr	Leu	Tyr	Asn	275	280	285
Arg	His	Ser	Leu	Asp	Thr	Leu	Ser	Ile	Lys	Arg	Thr	Ser	Ser	Val	Phe	290	295	300

Lys Arg Ile Cys Lys Leu Ile Val Ala Asn Asn Leu Phe Lys Ile Phe  
 305 310 315 320

Leu Asn Thr Leu Ile Arg Glu Glu Lys Asn Asn Asp  
 325 330

<210> 22

<211> 332

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS2K

<400> 22

Met Ile Asn Ile Ser Ile Ile Val Pro Ile Tyr Asn Val Glu Gln Tyr  
 1 5 10 15

Leu Ser Lys Cys Ile Asn Ser Ile Val Asn Gln Thr Tyr Lys His Ile  
 20 25 30

Glu Ile Leu Leu Val Asn Asp Gly Ser Thr Asp Asn Ser Glu Glu Ile  
 35 40 45

Cys Leu Ala Tyr Ala Lys Lys Asp Ser Arg Ile Arg Tyr Phe Lys Lys  
 50 55 60

Glu Asn Gly Gly Leu Ser Asp Ala Arg Asn Tyr Gly Ile Ser Arg Ala  
 65 70 75 80

Lys Gly Asp Tyr Leu Ala Phe Ile Asp Ser Asp Asp Phe Ile His Ser  
 85 90 95

Glu Phe Ile Gln Arg Leu His Glu Ala Ile Glu Arg Glu Asn Ala Leu  
 100 105 110

Val Ala Val Ala Gly Tyr Asp Arg Val Asp Ala Ser Gly His Phe Leu  
 115 120 125

Thr Ala Glu Pro Leu Pro Thr Asn Gln Ala Val Leu Ser Gly Arg Asn  
 130 135 140

Val Cys Lys Lys Leu Leu Glu Ala Asp Gly His Arg Phe Val Val Ala  
 145 150 155 160



<222> (1)..(467)

<223> Xaa may be any amino acid

<400> 23

Met	Ser	Lys	Lys	Ser	Ile	Val	Val	Ser	Gly	Leu	Val	Tyr	Thr	Ile	Gly	
1				5					10					15		
Thr	Ile	Leu	Val	Gln	Gly	Leu	Ala	Phe	Ile	Thr	Leu	Pro	Ile	Tyr	Thr	
			20					25					30			
Arg	Val	Ile	Ser	Gln	Glu	Val	Tyr	Gly	Gln	Phe	Ser	Leu	Tyr	Asn	Ser	
		35					40					45				
Trp	Val	Gly	Leu	Val	Gly	Leu	Phe	Ile	Gly	Leu	Gln	Leu	Gly	Gly	Ala	
	50					55					60					
Phe	Gly	Pro	Gly	Trp	Val	His	Phe	Arg	Glu	Lys	Phe	Asp	Asp	Phe	Val	
65					70					75					80	
Ser	Thr	Leu	Met	Val	Ser	Ser	Ile	Ala	Phe	Phe	Leu	Pro	Ile	Phe	Gly	
				85					90					95		
Leu	Ser	Phe	Leu	Leu	Ser	Gln	Pro	Leu	Ser	Leu	Leu	Phe	Gly	Leu	Pro	
			100					105					110			
Asp	Trp	Val	Val	Pro	Leu	Ile	Phe	Leu	Gln	Ser	Leu	Met	Ile	Val	Val	
		115					120					125				
Gln	Gly	Phe	Phe	Thr	Thr	Tyr	Leu	Val	Gln	Arg	Gln	Gln	Ser	Met	Trp	
	130					135					140					
Thr	Leu	Pro	Leu	Ser	Val	Leu	Ser	Ala	Val	Ile	Asn	Thr	Ala	Leu	Ser	
145					150					155					160	
Leu	Phe	Leu	Thr	Phe	Pro	Met	Glu	Asn	Asp	Phe	Ile	Ala	Arg	Val	Met	
				165					170					175		
Ala	Asn	Pro	Ala	Thr	Thr	Gly	Val	Leu	Ala	Cys	Val	Ser	Xaa	Trp	Phe	
			180					185					190			
Ser	Gln	Lys	Lys	Asn	Gly	Leu	His	Phe	Arg	Lys	Asp	Tyr	Leu	Arg	Tyr	
		195					200					205				
Gly	Leu	Ser	Ile	Ser	Ile	Pro	Leu	Ile	Phe	His	Gly	Leu	Gly	His	Asn	
	210					215					220					
Val	Leu	Asn	Gln	Phe	Asp	Arg	Ile	Met	Leu	Gly	Lys	Met	Leu	Thr	Leu	
225					230					235					240	

Ser Asp Val Ala Leu Tyr Ser Phe Gly Tyr Thr Leu Ala Ser Ile Leu  
 245 250 255  
 Gln Ile Val Phe Ser Ser Leu Asn Thr Val Trp Cys Pro Trp Tyr Phe  
 260 265 270  
 Glu Lys Lys Arg Gly Ala Asp Lys Asp Leu Leu Ser Tyr Val Arg Tyr  
 275 280 285  
 Tyr Leu Ala Ile Gly Leu Phe Val Thr Phe Gly Phe Leu Thr Ile Tyr  
 290 295 300  
 Pro Arg Leu Ala Met Leu Leu Gly Gly Ser Glu Tyr Arg Phe Ser Met  
 305 310 315 320  
 Gly Phe Ile Pro Met Ile Ile Val Gly Val Phe Phe Val Phe Leu Tyr  
 325 330 335  
 Ser Phe Pro Ala Asn Ile Gln Phe Tyr Ser Gly Asn Thr Lys Phe Leu  
 340 345 350  
 Pro Ile Gly Thr Phe Ile Ala Gly Val Leu Asn Ile Ser Val His Phe  
 355 360 365  
 Val Leu Ile Pro Thr Lys Asn Leu Trp Cys Cys Phe Ala Thr Thr Ala  
 370 375 380  
 Ser Tyr Leu Leu Leu Leu Val Leu His Tyr Phe Val Ala Lys Lys Lys  
 385 390 395 400  
 Tyr Ala Tyr Asp Glu Val Ala Ile Ser Thr Phe Val Lys Val Ile Ala  
 405 410 415  
 Leu Val Val Val Tyr Thr Gly Leu Met Thr Val Phe Val Gly Ser Ile  
 420 425 430  
 Trp Ile Arg Trp Ser Leu Gly Ile Ala Val Leu Val Val Tyr Ala Ile  
 435 440 445  
 Tyr Phe Arg Lys Glu Leu Thr Val Ala Leu Asn Thr Phe Arg Glu Lys  
 450 455 460  
 Arg Ser Lys  
 465

<210> 24

<211> 338

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS2P

<400> 24

Met	Val	Tyr	Ile	Ile	Ala	Glu	Ile	Gly	Cys	Asn	His	Asn	Gly	Asp	Val	
1				5					10					15		
His	Leu	Ala	Arg	Lys	Met	Val	Glu	Val	Ala	Val	Asp	Cys	Gly	Val	Asp	
			20					25					30			
Ala	Val	Lys	Phe	Gln	Thr	Glu	Lys	Ala	Asp	Leu	Leu	Ile	Ser	Lys	Tyr	
		35					40					45				
Ala	Pro	Lys	Ala	Glu	Tyr	Gln	Lys	Ile	Thr	Thr	Gly	Glu	Ser	Asp	Ser	
	50					55					60					
Gln	Leu	Glu	Met	Thr	Arg	Arg	Leu	Glu	Leu	Ser	Phe	Glu	Glu	Tyr	Leu	
65					70					75					80	
Asp	Leu	Arg	Asp	Tyr	Cys	Leu	Glu	Lys	Gly	Val	Asp	Val	Phe	Ser	Thr	
				85					90					95		
Pro	Glu	Asp	Glu	Glu	Ser	Leu	Asp	Phe	Leu	Ile	Ser	Thr	Asp	Met	Pro	
			100					105					110			
Val	Tyr	Lys	Ile	Pro	Ser	Gly	Glu	Ile	Thr	Asn	Leu	Pro	Tyr	Leu	Glu	
		115					120					125				
Lys	Ile	Gly	Arg	Gln	Ala	Lys	Lys	Val	Ile	Leu	Ser	Thr	Gly	Met	Ala	
	130					135					140					
Val	Met	Asp	Glu	Ile	His	Gln	Ala	Val	Lys	Ile	Leu	Gln	Glu	Asn	Gly	
145					150					155					160	
Thr	Thr	Asp	Ile	Ser	Ile	Leu	His	Cys	Thr	Thr	Glu	Tyr	Pro	Thr	Pro	
				165					170					175		
Tyr	Pro	Ala	Leu	Asn	Leu	Asn	Val	Leu	His	Thr	Leu	Lys	Lys	Glu	Phe	
			180					185					190			
Pro	Asn	Leu	Thr	Ile	Gly	Tyr	Ser	Asp	His	Ser	Val	Gly	Ser	Glu	Val	
	195						200					205				
Pro	Ile	Ala	Ala	Ala	Ala	Met	Gly	Ala	Glu	Leu	Ile	Glu	Lys	His	Phe	
	210					215					220					



Thr Leu Asp Asn Glu Met Glu Gly Pro Asp His Lys Ala Ser Ala Thr  
 225 230 235 240  
 Pro Asp Ile Leu Ala Ala Leu Val Lys Gly Val Arg Ile Val Glu Gln  
 245 250 255  
 Ser Leu Gly Lys Phe Glu Lys Glu Pro Glu Glu Val Glu Val Arg Asn  
 260 265 270  
 Lys Ile Val Ala Glu Lys Ser Ile Val Ala Lys Lys Ala Ile Ala Lys  
 275 280 285  
 Gly Glu Val Phe Thr Glu Glu Asn Ile Thr Val Lys Arg Pro Gly Asn  
 290 295 300  
 Gly Ile Ser Pro Met Glu Trp Tyr Lys Val Leu Gly Gln Val Ser Glu  
 305 310 315 320  
 Gln Asp Phe Glu Glu Asp Gln Asn Ile Cys His Ser Ala Phe Glu Asn  
 325 330 335

Gln Met

<210> 25

<211> 170

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS2Q

<400> 25

Met Lys Lys Ile Cys Phe Val Thr Gly Ser Arg Ala Glu Tyr Gly Ile  
 1 5 10 15  
 Met Arg Arg Leu Leu Ser Tyr Leu Gln Asp Asp Pro Glu Met Glu Leu  
 20 25 30  
 Asp Leu Val Val Ala Thr Met His Leu Glu Glu Lys Tyr Gly Met Thr  
 35 40 45  
 Val Lys Asp Ile Glu Ala Asp Lys Arg Arg Ile Val Lys Arg Ile Pro  
 50 55 60

Leu His Leu Thr Asp Thr Ser Lys Gln Thr Ile Val Lys Ser Leu Ala  
 65 70 75 80  
 Thr Leu Thr Glu Gln Leu Thr Val Leu Phe Glu Glu Val Gln Tyr Asp  
 85 90 95  
 Leu Val Leu Ile Leu Gly Asp Arg Tyr Glu Met Leu Pro Val Ala Asn  
 100 105 110  
 Ala Ala Leu Leu Tyr Asn Ile Pro Ile Cys His Ile His Gly Gly Glu  
 115 120 125  
 Lys Thr Met Gly Asn Phe Asp Glu Ser Ile Arg His Ala Ile Thr Lys  
 130 135 140  
 Met Ser His Leu His Leu Thr Ser Thr Asp Glu Phe Arg Asn Arg Val  
 145 150 155 160  
 Ile Gln Leu Gly Glu Asn Pro Thr Met Tyr  
 165 170

<210> 26

<211> 184

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS2R

<400> 26

Met Glu Leu Gly Ile Asp Phe Ala Glu Asp Tyr Tyr Val Val Leu Phe  
 1 5 10 15  
 His Pro Val Thr Leu Glu Asp Asn Thr Ala Glu Glu Gln Thr Gln Ala  
 20 25 30  
 Leu Leu Asp Ala Leu Lys Glu Asp Gly Ser Gln Cys Leu Ile Ile Gly  
 35 40 45  
 Ser Asn Ser Asp Thr His Ala Asp Lys Ile Met Glu Leu Met His Glu  
 50 55 60

Phe	Val	Lys	Gln	Asp	Ser	Asp	Ser	Tyr	Ile	Phe	Thr	Ser	Leu	Pro	Thr
65					70					75					80
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				85					90					95	
Ser	Ser	Ser	Gly	Leu	Ile	Glu	Val	Pro	Ser	Leu	Gln	Val	Pro	Thr	Leu
			100					105					110		
Asn	Ile	Gly	Asn	Arg	Gln	Phe	Gly	Arg	Leu	Ser	Gly	Pro	Ser	Val	Val
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His	Val	Gly	Thr	Ser	Lys	Glu	Ala	Ile	Val	Gly	Gly	Leu	Gly	Gln	Leu
	130					135					140				
Arg	Asp	Val	Ile	Asp	Phe	Thr	Asn	Pro	Phe	Glu	Gln	Pro	Asp	Ser	Ala
145					150					155					160
Leu	Gln	Gly	Tyr	Arg	Ala	Ile	Lys	Glu	Phe	Leu	Ser	Val	Gln	Ala	Ser
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<210> 27

<211> 208

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS2S

<400> 27

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			20					25					30		
Asp	Lys	Pro	Ile	Ser	Asp	Tyr	Arg	Gly	Tyr	Pro	Val	Phe	Gly	Pro	Leu
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Gln	Asp	Val	Leu	Thr	Tyr	Leu	Asp	Asp	Gly	Lys	Val	Asp	Ala	Val	Phe
	50					55					60				

Val	Thr	Ile	Gly	Asp	Asn	Val	Lys	Arg	Lys	Glu	Ile	Phe	Asp	Leu	Leu
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Ala	Lys	Asp	His	Tyr	Asp	Ala	Leu	Phe	Asn	Ile	Ile	Ser	Glu	Gln	Ala
			85						90					95	
Asn	Ile	Phe	Ser	Pro	Asp	Ser	Ile	Lys	Gly	Arg	Gly	Val	Phe	Ile	Gly
			100					105					110		
Phe	Ser	Ser	Phe	Val	Gly	Ala	Asp	Ser	Tyr	Val	Tyr	Asp	Asn	Cys	Ile
		115					120					125			
Ile	Asn	Thr	Gly	Ala	Ile	Val	Glu	His	His	Thr	Thr	Val	Glu	Ala	His
	130					135					140				
Cys	Asn	Ile	Thr	Pro	Gly	Val	Thr	Ile	Asn	Gly	Leu	Cys	Arg	Ile	Gly
145					150					155					160
Glu	Ser	Thr	Tyr	Ile	Gly	Ser	Gly	Ser	Thr	Val	Ile	Gln	Cys	Ile	Glu
				165					170					175	
Ile	Ala	Pro	Tyr	Thr	Thr	Leu	Gly	Ala	Gly	Thr	Val	Val	Leu	Lys	Ser
			180					185					190		
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<210> 28

<211> 410

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS2T

<400> 28

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			20					25					30		

Thr	Ile	Arg	Ala	Ala	Ile	Glu	Ser	Gly	Cys	Phe	Lys	Lys	Glu	Asn	Ile		
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Tyr	Val	Ser	Thr	Asp	Ser	Glu	Val	Tyr	Lys	Glu	Ile	Cys	Glu	Thr	Thr		
	50					55					60						
Gly	Val	Gln	Val	Leu	Met	Arg	Pro	Ala	Asp	Leu	Ala	Thr	Asp	Phe	Thr		
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Thr	Ser	Phe	Gln	Leu	Asn	Glu	His	Phe	Leu	Gln	Asp	Phe	Ser	Asp	Asp		
				85					90					95			
Gln	Val	Phe	Val	Leu	Leu	Gln	Val	Thr	Ser	Pro	Leu	Arg	Ser	Gly	Lys		
			100					105					110				
His	Val	Lys	Glu	Ala	Met	Glu	Leu	Tyr	Gly	Lys	Gly	Gln	Ala	Asp	His		
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Val	Val	Ser	Phe	Thr	Lys	Val	Asp	Lys	Ser	Pro	Thr	Leu	Phe	Ser	Thr		
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Tyr	Ile	Ser	Ser	Lys	Gln	Ala	Tyr	Leu	Ala	Asp	Lys	Thr	Tyr	Phe	Ser		
			180					185					190				
Glu	Lys	Thr	Ala	Ala	Tyr	Val	Met	Thr	Lys	Glu	Asp	Ser	Ile	Asp	Val		
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Asp	Asp	His	Phe	Asp	Phe	Thr	Gly	Val	Ile	Gly	Arg	Ile	Tyr	Phe	Asp		
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Tyr	Gln	Arg	Arg	Glu	Gln	Gln	Asn	Lys	Pro	Phe	Tyr	Lys	Arg	Glu	Leu		
225					230					235					240		
Lys	Arg	Leu	Cys	Glu	Gln	Arg	Val	His	Asp	Ser	Leu	Val	Ile	Gly	Asp		
				245					250					255			
Ser	Arg	Leu	Leu	Ala	Leu	Leu	Leu	Asp	Gly	Phe	Asp	Asn	Ile	Ser	Ile		
			260					265					270				
Gly	Gly	Met	Thr	Ala	Ser	Thr	Ser	Leu	Glu	Asn	Gln	Gly	Leu	Phe	Leu		
		275					280					285					
Ala	Thr	Pro	Ile	Lys	Lys	Val	Leu	Leu	Ser	Leu	Gly	Val	Asn	Asp	Leu		
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Ile	Thr	Asp	Tyr	Pro	Leu	His	Met	Ile	Glu	Asp	Thr	Ile	Arg	Gln	Leu
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Ile	Asp	Leu	Asn	Glu	Val	Val	Glu	Lys	Glu	Ala	Met	Leu	Asp	Tyr	Gln
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Tyr	Thr	Asn	Asp	Gly	Leu	His	Phe	Asn	Gln	Ile	Gly	Gln	Glu	Arg	Val
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<211> 6992

<212> DNA

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS1

<400> 29

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<210> 30

<211> 454

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS1E

<400> 30

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Met	Pro	Val	Glu	Phe	Glu	Tyr	Arg	Gly	Asn	Leu	Ile	Glu	Phe	Glu	Lys	50	55	60	
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Phe	Leu	Leu	Glu	Asn	Asn	Phe	Ala	Leu	Ser	Arg	Arg	Gly	Ala	Val	Tyr	85	90	95	
Phe	Thr	Leu	Ile	Asn	Phe	Val	Leu	Val	Tyr	Leu	Phe	Asn	Val	Ile	Ile	100	105	110	
Lys	Gln	Phe	Lys	Asp	Ser	Phe	Leu	Phe	Ser	Thr	Ile	Tyr	Gln	Lys	Lys	115	120	125	

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Phe	Glu	Lys	Tyr	Thr	Pro	Gly	Gln	Lys	Arg	Arg	Leu	Ser	Phe	Lys	Pro			
385					390					395					400			

Gly Ile Thr Gly Leu Trp Gln Val Ser Gly Arg Ser Asn Ile Thr Asp  
405 410 415  
Phe Asp Asp Val Val Arg Leu Asp Leu Ala Tyr Ile Asp Asn Trp Thr  
420 425 430  
Ile Trp Ser Asp Ile Lys Ile Leu Leu Lys Thr Val Lys Val Val Leu  
435 440 445  
Leu Arg Glu Gly Ser Lys  
450

<210> 31

<211> 149

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS1F

<400> 31

Met Lys Val Cys Leu Val Gly Ser Ser Gly Gly His Leu Thr His Leu  
1 5 10 15  
Tyr Leu Leu Lys Pro Phe Trp Lys Glu Glu Glu Arg Phe Trp Val Thr  
20 25 30  
Phe Asp Lys Glu Asp Ala Arg Ser Leu Leu Lys Asn Glu Lys Met Tyr  
35 40 45  
Pro Cys Tyr Phe Pro Thr Asn Arg Asn Leu Ile Asn Leu Val Lys Asn  
50 55 60  
Thr Phe Leu Ala Phe Lys Ile Leu Arg Asp Glu Lys Pro Asp Val Ile  
65 70 75 80  
Ile Ser Ser Gly Ala Ala Val Ala Val Pro Phe Phe Tyr Ile Gly Lys  
85 90 95  
Leu Phe Gly Ala Lys Thr Ile Tyr Ile Glu Val Phe Asp Arg Val Asn  
100 105 110  
Lys Ser Thr Leu Thr Gly Lys Leu Val Tyr Pro Val Thr Asp Ile Phe  
115 120 125

Ile Val Gln Trp Glu Glu Met Lys Lys Val Tyr Pro Lys Ser Ile Asn  
130 135 140

Leu Gly Ser Ile Phe  
145

<210> 32

<211> 164

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS1G

<400> 32

Met Ile Phe Val Thr Val Gly Thr His Glu Gln Gln Phe Asn Arg Leu  
1 5 10 15

Ile Lys Glu Ile Asp Leu Leu Lys Lys Asn Gly Ser Ile Thr Asp Glu  
20 25 30

Ile Phe Ile Gln Thr Gly Tyr Ser Asp Tyr Ile Pro Glu Tyr Cys Lys  
35 40 45

Tyr Lys Lys Phe Leu Ser Tyr Lys Glu Met Glu Gln Tyr Ile Asn Lys  
50 55 60

Ser Glu Val Val Ile Cys His Gly Gly Pro Ala Thr Phe Met Asn Ser  
65 70 75 80

Leu Ser Lys Gly Lys Lys Gln Leu Leu Phe Pro Arg Gln Lys Lys Tyr  
85 90 95

Gly Glu His Val Asn Asp His Gln Val Glu Phe Val Arg Arg Ile Leu  
100 105 110

Gln Asp Asn Asn Ile Leu Phe Ile Glu Asn Ile Asp Asp Leu Phe Glu  
115 120 125

Lys Ile Ile Glu Val Ser Lys Gln Thr Asn Phe Thr Ser Asn Asn Asn  
130 135 140

Phe	Phe	Cys	Glu	Arg	Leu	Lys	Gln	Ile	Val	Glu	Lys	Phe	Asn	Glu	Asp
145					150					155					160

Gln Glu Asn Glu

<210> 33

<211> 388

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS1H

<400> 33

Met	Phe	Lys	Leu	Phe	Lys	Tyr	Asp	Pro	Glu	Tyr	Phe	Ile	Phe	Lys	Tyr
1				5					10					15	

Phe	Trp	Leu	Ile	Ile	Phe	Ile	Pro	Glu	Gln	Lys	Tyr	Val	Phe	Leu	Leu
		20					25					30			

Ile	Phe	Met	Asn	Leu	Ile	Leu	Phe	His	Ile	Lys	Phe	Leu	Lys	Thr	Lys
		35					40					45			

Leu	Ile	Leu	Lys	Asn	Glu	Ile	Leu	Leu	Phe	Leu	Leu	Trp	Ser	Ile	Leu
	50					55					60				

Cys	Phe	Val	Ser	Val	Val	Thr	Ser	Met	Phe	Val	Glu	Ile	Asn	Phe	Glu
65					70					75					80

Arg	Leu	Phe	Ala	Asp	Phe	Thr	Ala	Pro	Ile	Ile	Trp	Ile	Ile	Ala	Ile
				85					90					95	

Met	Tyr	Tyr	Asn	Leu	Tyr	Ser	Phe	Ile	Asn	Ile	Asp	Tyr	Lys	Lys	Leu
			100					105					110		

Lys	Asn	Ser	Ile	Phe	Phe	Ser	Phe	Leu	Val	Leu	Leu	Gly	Ile	Ser	Ala
		115					120					125			

Leu	Tyr	Ile	Ile	Gln	Asn	Gly	Lys	Asp	Ile	Val	Phe	Leu	Asp	Arg	His
	130					135					140				

Leu	Ile	Gly	Leu	Asp	Tyr	Leu	Ile	Thr	Gly	Val	Lys	Thr	Arg	Leu	Val
145					150					155					160

Gly	Phe	Met	Asn	Tyr	Pro	Thr	Leu	Asn	Thr	Thr	Thr	Ile	Ile	Val	Ser	
			165						170					175		
Ile	Pro	Leu	Ile	Phe	Ala	Leu	Ile	Lys	Asn	Lys	Met	Gln	Gln	Phe	Phe	
			180					185					190			
Phe	Leu	Cys	Leu	Ala	Phe	Ile	Pro	Ile	Tyr	Leu	Ser	Gly	Ser	Arg	Ile	
		195					200					205				
Gly	Ser	Leu	Ser	Leu	Ala	Ile	Leu	Ile	Ile	Cys	Leu	Leu	Trp	Arg	Tyr	
	210					215					220					
Ile	Gly	Gly	Lys	Phe	Ala	Trp	Ile	Lys	Lys	Leu	Ile	Val	Ile	Phe	Val	
225					230					235					240	
Ile	Leu	Leu	Ile	Ile	Leu	Asn	Thr	Glu	Leu	Leu	Tyr	His	Glu	Ile	Leu	
				245					250					255		
Ala	Val	Tyr	Asn	Ser	Arg	Glu	Ser	Ser	Asn	Glu	Ala	Arg	Phe	Ile	Ile	
			260					265					270			
Tyr	Gln	Gly	Ser	Ile	Asp	Lys	Val	Leu	Glu	Asn	Asn	Ile	Leu	Phe	Gly	
		275					280					285				
Tyr	Gly	Ile	Ser	Glu	Tyr	Ser	Val	Thr	Gly	Thr	Trp	Leu	Gly	Ser	His	
	290					295					300					
Ser	Gly	Tyr	Ile	Ser	Phe	Phe	Tyr	Lys	Ser	Gly	Ile	Val	Gly	Leu	Ile	
305					310					315					320	
Leu	Leu	Met	Phe	Ser	Phe	Phe	Tyr	Val	Ile	Lys	Lys	Ser	Tyr	Gly	Val	
				325					330					335		
Asn	Gly	Glu	Thr	Ala	Leu	Phe	Tyr	Phe	Thr	Ser	Leu	Ala	Ile	Phe	Phe	
			340					345					350			
Ile	Tyr	Glu	Thr	Ile	Asp	Pro	Ile	Ile	Ile	Ile	Leu	Val	Leu	Phe	Phe	
		355					360					365				
Ser	Ser	Ile	Gly	Ile	Trp	Asn	Asn	Ile	Asn	Phe	Lys	Lys	Asp	Met	Glu	
	370					375					380					
Thr	Lys	Asn	Glu													
385																

<210> 34

<211> 322

<212> PRT



<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS1I

<400> 34

Met	Asn	Asp	Leu	Ile	Ser	Val	Ile	Val	Pro	Ile	Tyr	Asn	Val	Gln	Asp	
1				5					10					15		
Tyr	Leu	Asp	Lys	Cys	Ile	Asn	Ser	Ile	Ile	Asn	Gln	Thr	Tyr	Thr	Asn	
			20					25					30			
Leu	Glu	Val	Ile	Leu	Val	Asn	Asp	Gly	Ser	Thr	Asp	Asp	Ser	Glu	Lys	
		35					40					45				
Ile	Cys	Leu	Asn	Tyr	Met	Lys	Asn	Asp	Gly	Arg	Ile	Lys	Tyr	Tyr	Lys	
	50					55					60					
Lys	Ile	Asn	Gly	Gly	Leu	Ala	Asp	Ala	Arg	Asn	Phe	Gly	Leu	Glu	His	
65					70					75					80	
Ala	Thr	Gly	Lys	Tyr	Ile	Ala	Phe	Val	Asp	Ser	Asp	Asp	Tyr	Ile	Glu	
			85						90					95		
Val	Ala	Met	Phe	Glu	Arg	Met	His	Asp	Asn	Ile	Thr	Glu	Tyr	Asn	Ala	
		100						105					110			
Asp	Ile	Ala	Glu	Ile	Asp	Phe	Cys	Leu	Val	Asp	Glu	Asn	Gly	Tyr	Thr	
		115					120					125				
Lys	Lys	Lys	Arg	Asn	Ser	Asn	Phe	His	Val	Leu	Thr	Arg	Glu	Glu	Thr	
	130					135					140					
Val	Lys	Glu	Phe	Leu	Ser	Gly	Ser	Asn	Ile	Glu	Asn	Asn	Val	Trp	Cys	
145					150					155					160	
Lys	Leu	Tyr	Ser	Arg	Asp	Ile	Ile	Lys	Asp	Ile	Lys	Phe	Gln	Ile	Asn	
				165					170					175		
Asn	Arg	Ser	Ile	Gly	Glu	Asp	Leu	Leu	Phe	Asn	Leu	Glu	Val	Leu	Asn	
			180					185					190			
Asn	Val	Thr	Arg	Val	Val	Val	Asp	Thr	Arg	Glu	Tyr	Tyr	Tyr	Asn	Tyr	
		195					200					205				
Val	Ile	Arg	Asn	Ser	Ser	Leu	Ile	Asn	Gln	Lys	Phe	Ser	Ile	Asn	Asn	
	210					215					220					

Ile Asp Leu Val Thr Arg Leu Glu Asn Tyr Pro Phe Lys Leu Lys Arg  
 225 230 235 240  
 Glu Phe Ser His Tyr Phe Asp Ala Lys Val Ile Lys Glu Lys Val Lys  
 245 250 255  
 Cys Leu Asn Lys Met Tyr Ser Thr Asp Cys Leu Asp Asn Glu Phe Leu  
 260 265 270  
 Pro Ile Leu Glu Ser Tyr Arg Lys Glu Ile Arg Arg Tyr Pro Phe Ile  
 275 280 285  
 Lys Ala Lys Arg Tyr Leu Ser Arg Lys His Leu Val Thr Leu Tyr Leu  
 290 295 300  
 Met Lys Phe Ser Pro Lys Leu Tyr Val Met Leu Tyr Lys Lys Phe Gln  
 305 310 315 320  
 Lys Gln

<210> 35

<211> 322

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS1J

<400> 35

Met Asp Lys Ile Ser Val Ile Val Pro Val Tyr Asn Val Asp Lys Tyr  
 1 5 10 15  
 Leu Ser Ser Cys Ile Glu Ser Ile Ile Asn Gln Asn Tyr Lys Asn Ile  
 20 25 30  
 Glu Ile Leu Leu Ile Asp Asp Gly Ser Val Asp Asp Ser Ala Lys Ile  
 35 40 45  
 Cys Lys Glu Tyr Glu Lys Asp Lys Arg Val Lys Ile Phe Phe Thr Asn  
 50 55 60

His	Ser	Gly	Val	Ser	Asn	Ala	Arg	Asn	His	Gly	Ile	Lys	Arg	Ser	Thr	65	70	75	80
Ala	Glu	Tyr	Ile	Met	Phe	Val	Asp	Ser	Asp	Asp	Val	Val	Asp	Ser	Arg	85	90	95	
Leu	Val	Glu	Lys	Leu	Tyr	Phe	Asn	Ile	Ile	Lys	Ser	Arg	Ser	Asp	Leu	100	105	110	
Ser	Gly	Cys	Leu	Tyr	Ala	Thr	Phe	Ser	Glu	Asn	Ile	Asn	Asn	Phe	Glu	115	120	125	
Val	Asn	Asn	Pro	Asn	Ile	Asp	Phe	Glu	Ala	Ile	Asn	Thr	Val	Gln	Asp	130	135	140	
Met	Gly	Glu	Lys	Asn	Phe	Met	Asn	Leu	Tyr	Ile	Asn	Asn	Ile	Phe	Ser	145	150	155	160
Thr	Pro	Val	Cys	Lys	Leu	Tyr	Lys	Lys	Arg	Tyr	Ile	Thr	Asp	Leu	Phe	165	170	175	
Gln	Glu	Asn	Gln	Trp	Leu	Gly	Glu	Asp	Leu	Leu	Phe	Asn	Leu	His	Tyr	180	185	190	
Leu	Lys	Asn	Ile	Asp	Arg	Val	Ser	Tyr	Leu	Thr	Glu	His	Leu	Tyr	Phe	195	200	205	
Tyr	Arg	Arg	Gly	Ile	Leu	Ser	Thr	Val	Asn	Ser	Phe	Lys	Glu	Gly	Val	210	215	220	
Phe	Leu	Gln	Leu	Glu	Asn	Leu	Gln	Lys	Gln	Val	Ile	Val	Leu	Phe	Lys	225	230	235	240
Gln	Ile	Tyr	Gly	Glu	Asp	Phe	Asp	Val	Ser	Ile	Val	Lys	Asp	Thr	Ile	245	250	255	
Arg	Trp	Gln	Val	Phe	Tyr	Tyr	Ser	Leu	Leu	Met	Phe	Lys	Tyr	Gly	Lys	260	265	270	
Gln	Ser	Ile	Phe	Asp	Lys	Phe	Leu	Ile	Phe	Arg	Asn	Leu	Tyr	Lys	Lys	275	280	285	
Tyr	Tyr	Phe	Asn	Leu	Leu	Lys	Val	Ser	Asn	Lys	Asn	Ser	Leu	Ser	Lys	290	295	300	
Asn	Phe	Cys	Ile	Arg	Ile	Val	Ser	Asn	Lys	Val	Phe	Lys	Lys	Ile	Leu	305	310	315	320
Trp	Leu																		

<210> 36

<211> 278

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS1K

<400> 36

Met Asp Thr Ile Ser Lys Ile Ser Ile Ile Val Pro Ile Tyr Asn Val  
1 5 10 15

Glu Lys Tyr Leu Ser Lys Cys Ile Asp Ser Ile Val Asn Gln Thr Tyr  
20 25 30

Lys His Ile Glu Ile Leu Leu Val Asn Asp Gly Ser Thr Asp Asn Ser  
35 40 45

Glu Glu Ile Cys Leu Ala Tyr Ala Lys Lys Asp Ser Arg Ile Arg Tyr  
50 55 60

Phe Lys Lys Glu Asn Gly Gly Leu Ser Asp Ala Arg Asn Tyr Gly Ile  
65 70 75 80

Ser Arg Ala Lys Gly Asp Tyr Leu Ala Phe Ile Asp Ser Asp Asp Phe  
85 90 95

Ile His Ser Glu Phe Ile Gln Arg Leu His Glu Ala Ile Glu Arg Glu  
100 105 110

Asn Ala Leu Val Ala Val Ala Gly Tyr Asp Arg Val Asp Ala Ser Gly  
115 120 125

His Phe Leu Thr Ala Glu Pro Leu Pro Thr Asn Gln Ala Val Leu Ser  
130 135 140

Gly Arg Asn Val Cys Lys Lys Leu Leu Glu Ala Asp Gly His Arg Phe  
145 150 155 160

Val Val Ala Cys Asn Lys Leu Tyr Lys Lys Glu Leu Phe Glu Asp Phe  
165 170 175

Arg Phe Glu Lys Gly Lys Ile His Glu Asp Glu Tyr Phe Thr Tyr Arg  
180 185 190

Leu Leu Tyr Glu Leu Glu Lys Val Ala Ile Val Lys Glu Cys Leu Tyr  
 195 200 205  
 Tyr Tyr Val Asp Arg Glu Asn Ser Ile Thr Thr Ser Ser Met Thr Asp  
 210 215 220  
 His Arg Phe His Cys Leu Leu Glu Phe Gln Asn Glu Arg Met Asp Phe  
 225 230 235 240  
 Tyr Glu Ser Arg Gly Asp Lys Glu Leu Leu Leu Glu Cys Tyr Arg Ser  
 245 250 255  
 Phe Leu Ala Phe Ala Val Leu Phe Leu Gly Lys Tyr Asn His Trp Leu  
 260 265 270  
 Ser Lys Gln Gln Lys Lys  
 275

<210> 37

<211> 4519

<212> DNA

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS9

<400> 37

aagcttatcg tcaaggtggt cgctatatcg tggcgacatc tcatagacga aaagggatgt 60  
 ttgaaacacc agaaaaagtt atcatgacta actttcttca atttaaagac gcagtagcag 120  
 aagtttatcc tgaaatacga ttgtgctatg gtgctgaatt gtattatagt aaagatatat 180  
 taagcaaact tgaaaaaaag aaagtaccca cacttaatgg ctgcgctat attcttttgg 240  
 agttcagtag tgatactcct tggaaagaga ttcaagaagc agtgaacgaa gtgacgctac 300  
 ttgggctaac tcccgtactt gcccatatag aacgatatga cgccctagcg tttcatgcag 360  
 agagagtaga agagttaatt gacaagggat gctatactca ggtaaatagt aatcatgtgc 420  
 tgaagcccac ttttaattggt gatcgagcaa aagaatttaa aaaacgtact cggatattttt 480  
 tagagcagga tttagtagat tgtgttgcta gcgatatgca taatttatct agtagacctc 540

cgtttatgag ggaggcttat aagttgctaa cagaggaatt tggcaaagat aaagcgaaag 600  
 cgttgctaaa aaagaatcct cttatgctat taaaaaacca ggcgatttaa actggttact 660  
 ctagattgtg gagagaaaaa tggatttagg aactgttact gataaactgt tagaacgcaa 720  
 cagtaaacga ttgatactcg tgtgcatgga tacgtgtctt cttatagttt ccatgatttt 780  
 gagcagactg tttttggatg ttattattga cataccagat gaacgcttca ttcttgacgt 840  
 tttattcgta tcaattttat atttgattct atcgtttaga ttaaaagtct tttcattaat 900  
 tacgcgttac acagggatc agagttatgt aaaaatagga cttagttaa tatctgcgca 960  
 ttcattgttt ttaattatct caatgggtgt gtggcaggct tttagtata gtttcacctt 1020  
 agtatcctta tttttgtcgt atgtaatgct cattactccg aggattgttt ggaaagtctt 1080  
 acatgagacg agaaaaaatg ctatccgtaa gaaggatagc ccactaagaa tcttagtagt 1140  
 aggtgctgga gatgggtgga atatttttat caatactgtc aaagatcgaa aattgaattt 1200  
 tgaaattgtc ggtatcgttg atcgtgatcc aaataaactt ggaacattta tccgtacggc 1260  
 taaagtttta ggaaaccgta atgatattcc acgactggta gaggaattag ctgttgacca 1320  
 agtgacgatt gccatccctt ctttaaattg taaggagcga gagaagattg ttgaaatctg 1380  
 taacactaca ggagtgaccg tcaataatat gccgagtatt gaagacatta tggcggggaa 1440  
 catgtctgtc agtgcccttc aggaaattga cgtagcagac cttcttggtc gaccagaggt 1500  
 tgttttggat caggatgaat tgaatcagtt tttccaaggg aaaacaatcc ttgtcacagg 1560  
 agcagggtggc tctatcgtt cagagctatg tcgtcaaatt gctaagttta cgcctaaacg 1620  
 cttgttggtg cttggacatg gagaaaattc aatctatctc attcatcgag agttactgga 1680  
 aaagtaccaa ggtaagattg agttggtccc tctcattgca gatattcaag atagagaatt 1740  
 gatttttagc ataattggctg aatatcaacc cgatgttggt tatcatgctg cagcacataa 1800  
 gcatgttcct ttgatggaat ataatccaca tgaagcagtg aagaataata tttttggaac 1860  
 gaagaatgtg gctgaggcgg ctaaaactgc aaagggtgcc aaatttgta tggtttcaac 1920  
 agataaagct gttaatccac caaatgtcat gggagcgact aaacgtgttg cagaaatgat 1980  
 tgttacaggt ttaaacgagc caggtcagac tcaatttgcg gcagtccggt ttgggaatgt 2040  
 tctaggtagt cgtggaagtg ttgttccgct attcaaagag caaattagaa aaggtggacc 2100

tgttacggtt accgacttta ggatgactcg ttatttcatg acgattcctg aggcaagtcg 2160  
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 cgagccagta caaatcctgg aattggcaag aaaagttatc ttgttaagtg gacacacaga 2280  
 ggaagaaatc gggattgtag aatctggaat cagaccaggc gagaaactct acgaggaatt 2340  
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 aatgaatta aaaaatatgt tgattgaatt tgcaaaacaa gaataagaaa gtaaaaaata 2520  
 tttttacttt cctagagttt aaacgatgtt taagttctag gaaggttaga atacctaatt 2580  
 aacaacaata ttactattta ttaagagtca gataatagca actaagtgtc acaactatc 2640  
 ttataataa gtatatttgg tcaaaaggga gatgtgaaat gtatccaatt tgtaaacgta 2700  
 ttttagcaat tattatctca gggattgcta ttgttgttct gagtccaatt ttattattga 2760  
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 gtaaaaacaa gtcatacttt atgatttata aattccgttc tatgtacgtt gacgcaccaa 2880  
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 tggcgattgt tgggtccacgc ccagccttat ggaatcaata tgacttaatt gaagagcgag 3060  
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 gtgatgaatt ggaaattgat gaaaagtcaa aattagatgg atattatgtt caaaatatga 3180  
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 tcaaacaatg attccaacgg aggttgtctt ggtagaggat gggccactca atcagagctt 3420  
 atatagtatt ttagaagaat ttaaaagtcg attttcattt tttaaaacga tagccttgga 3480  
 aaagaattcg ggtttaggaa ttgcactgaa tgaaggtttg aaacattgta attatgagt 3540  
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 taactttata aaacaaaacc cgactataga tattgagata gatgagttct taaattctac 3660

tagtgaaata gtttctcata aaaatgttcc aaccagcac gatgaaatat taaagatggc 3720  
 aaggcgggag aaatccatgt gccacatgac tgtaatgttt aaaaagaaaa gtgtcgagag 3780  
 agcagggggg tatcaaacac ttccgtacgt agaagattat ttcctttggg tgcgcatgat 3840  
 tgcttcagga tcgaaatttg caaacattga tgaaacacta gttcttgcac gtgttgga 3900  
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 atttatgtta gctcaaggaa ttgttacacc actagatgta tttattaatc aaatttacat 4020  
 tagggctctt gtttatatgc caacttggat aaagaaactc atttatggaa aaatcttaag 4080  
 gaaatagtat gattacagta ttgatggcta catataatgg aagcccattt ataataaac 4140  
 agttagattc aattcgaaat caaagtgtat cagcagacaa agttattatt tgggatgatt 4200  
 gctcgacaga tgatacaata aaaataataa aagattatat aaaaaaatat tctttggatt 4260  
 catgggttgt ctctcaaaat aaatctaactc aggggcatta tcaaacattt ataaatttga 4320  
 caaagttagt tcaggaagga atagtctttt tttcagatca agatgatatt tgggactgtc 4380  
 ataaatttga gacaatgctt ccaatctttg acagagaaaa tgtatcaatg gtgttttgca 4440  
 aatccagatt gattgatgaa aacggaaata ttatcagtag ccagatact tcggatagaa 4500  
 tcaatacgta ctctctaga 4519

<210> 38

<211> 215

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS9D

<400> 38

Ala	Tyr	Arg	Gln	Gly	Val	Arg	Tyr	Ile	Val	Ala	Thr	Ser	His	Arg	Arg
1				5					10					15	



Lys	Gly	Met	Phe	Glu	Thr	Pro	Glu	Lys	Val	Ile	Met	Thr	Asn	Phe	Leu			
			20					25					30					
Gln	Phe	Lys	Asp	Ala	Val	Ala	Glu	Val	Tyr	Pro	Glu	Ile	Arg	Leu	Cys			
		35					40					45						
Tyr	Gly	Ala	Glu	Leu	Tyr	Tyr	Ser	Lys	Asp	Ile	Leu	Ser	Lys	Leu	Glu			
	50					55					60							
Lys	Lys	Lys	Val	Pro	Thr	Leu	Asn	Gly	Ser	Arg	Tyr	Ile	Leu	Leu	Glu			
65					70					75					80			
Phe	Ser	Ser	Asp	Thr	Pro	Trp	Lys	Glu	Ile	Gln	Glu	Ala	Val	Asn	Glu			
				85					90					95				
Val	Thr	Leu	Leu	Gly	Leu	Thr	Pro	Val	Leu	Ala	His	Ile	Glu	Arg	Tyr			
			100					105					110					
Asp	Ala	Leu	Ala	Phe	His	Ala	Glu	Arg	Val	Glu	Glu	Leu	Ile	Asp	Lys			
		115					120					125						
Gly	Cys	Tyr	Thr	Gln	Val	Asn	Ser	Asn	His	Val	Leu	Lys	Pro	Thr	Leu			
	130					135					140							
Ile	Gly	Asp	Arg	Ala	Lys	Glu	Phe	Lys	Lys	Arg	Thr	Arg	Tyr	Phe	Leu			
145					150					155					160			
Glu	Gln	Asp	Leu	Val	His	Cys	Val	Ala	Ser	Asp	Met	His	Asn	Leu	Ser			
				165					170					175				
Ser	Arg	Pro	Pro	Phe	Met	Arg	Glu	Ala	Tyr	Lys	Leu	Leu	Thr	Glu	Glu			
			180					185						190				
Phe	Gly	Lys	Asp	Lys	Ala	Lys	Ala	Leu	Leu	Lys	Lys	Asn	Pro	Leu	Met			
		195					200					205						
Leu	Leu	Lys	Asn	Gln	Ala	Ile												
	210					215												

<210> 39

<211> 608

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS9E

<400> 39

Met	Asp	Leu	Gly	Thr	Val	Thr	Asp	Lys	Leu	Leu	Glu	Arg	Asn	Ser	Lys
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Arg	Leu	Ile	Leu	Val	Cys	Met	Asp	Thr	Cys	Leu	Leu	Ile	Val	Ser	Met
			20					25					30		
Ile	Leu	Ser	Arg	Leu	Phe	Leu	Asp	Val	Ile	Ile	Asp	Ile	Pro	Asp	Glu
		35					40					45			
Arg	Phe	Ile	Leu	Ala	Val	Leu	Phe	Val	Ser	Ile	Leu	Tyr	Leu	Ile	Leu
	50					55					60				
Ser	Phe	Arg	Leu	Lys	Val	Phe	Ser	Leu	Ile	Thr	Arg	Tyr	Thr	Gly	Tyr
65					70					75					80
Gln	Ser	Tyr	Val	Lys	Ile	Gly	Leu	Ser	Leu	Ile	Ser	Ala	His	Ser	Leu
				85					90					95	
Phe	Leu	Ile	Ile	Ser	Met	Val	Leu	Trp	Gln	Ala	Phe	Ser	Tyr	Arg	Phe
			100					105					110		
Ile	Leu	Val	Ser	Leu	Phe	Leu	Ser	Tyr	Val	Met	Leu	Ile	Thr	Pro	Arg
		115					120					125			
Ile	Val	Trp	Lys	Val	Leu	His	Glu	Thr	Arg	Lys	Asn	Ala	Ile	Arg	Lys
	130					135					140				
Lys	Asp	Ser	Pro	Leu	Arg	Ile	Leu	Val	Val	Gly	Ala	Gly	Asp	Gly	Gly
145					150					155					160
Asn	Ile	Phe	Ile	Asn	Thr	Val	Lys	Asp	Arg	Lys	Leu	Asn	Phe	Glu	Ile
				165					170					175	
Val	Gly	Ile	Val	Asp	Arg	Asp	Pro	Asn	Lys	Leu	Gly	Thr	Phe	Ile	Arg
			180					185					190		
Thr	Ala	Lys	Val	Leu	Gly	Asn	Arg	Asn	Asp	Ile	Pro	Arg	Leu	Val	Glu
		195					200					205			
Glu	Leu	Ala	Val	Asp	Gln	Val	Thr	Ile	Ala	Ile	Pro	Ser	Leu	Asn	Gly
	210					215					220				
Lys	Glu	Arg	Glu	Lys	Ile	Val	Glu	Ile	Cys	Asn	Thr	Thr	Gly	Val	Thr
225					230					235					240
Val	Asn	Asn	Met	Pro	Ser	Ile	Glu	Asp	Ile	Met	Ala	Gly	Asn	Met	Ser
				245					250					255	

Val	Ser	Ala	Phe	Gln	Glu	Ile	Asp	Val	Ala	Asp	Leu	Leu	Gly	Arg	Pro		
			260					265					270				
Glu	Val	Val	Leu	Asp	Gln	Asp	Glu	Leu	Asn	Gln	Phe	Phe	Gln	Gly	Lys		
		275					280					285					
Thr	Ile	Leu	Val	Thr	Gly	Ala	Gly	Gly	Ser	Ile	Gly	Ser	Glu	Leu	Cys		
	290					295					300						
Arg	Gln	Ile	Ala	Lys	Phe	Thr	Pro	Lys	Arg	Leu	Leu	Leu	Leu	Gly	His		
305					310					315					320		
Gly	Glu	Asn	Ser	Ile	Tyr	Leu	Ile	His	Arg	Glu	Leu	Leu	Glu	Lys	Tyr		
				325					330					335			
Gln	Gly	Lys	Ile	Glu	Leu	Val	Pro	Leu	Ile	Ala	Asp	Ile	Gln	Asp	Arg		
			340					345					350				
Glu	Leu	Ile	Phe	Ser	Ile	Met	Ala	Glu	Tyr	Gln	Pro	Asp	Val	Val	Tyr		
		355					360					365					
His	Ala	Ala	Ala	His	Lys	His	Val	Pro	Leu	Met	Glu	Tyr	Asn	Pro	His		
	370					375					380						
Glu	Ala	Val	Lys	Asn	Asn	Ile	Phe	Gly	Thr	Lys	Asn	Val	Ala	Glu	Ala		
385					390					395					400		
Ala	Lys	Thr	Ala	Lys	Val	Ala	Lys	Phe	Val	Met	Val	Ser	Thr	Asp	Lys		
				405					410					415			
Ala	Val	Asn	Pro	Pro	Asn	Val	Met	Gly	Ala	Thr	Lys	Arg	Val	Ala	Glu		
			420					425					430				
Met	Ile	Val	Thr	Gly	Leu	Asn	Glu	Pro	Gly	Gln	Thr	Gln	Phe	Ala	Ala		
	435						440					445					
Val	Arg	Phe	Gly	Asn	Val	Leu	Gly	Ser	Arg	Gly	Ser	Val	Val	Pro	Leu		
	450					455					460						
Phe	Lys	Glu	Gln	Ile	Arg	Lys	Gly	Gly	Pro	Val	Thr	Val	Thr	Asp	Phe		
465					470					475					480		
Arg	Met	Thr	Arg	Tyr	Phe	Met	Thr	Ile	Pro	Glu	Ala	Ser	Arg	Leu	Val		
				485					490					495			
Ile	Gln	Ala	Gly	His	Leu	Ala	Lys	Gly	Gly	Glu	Ile	Phe	Val	Leu	Asp		
			500					505					510				
Met	Gly	Glu	Pro	Val	Gln	Ile	Leu	Glu	Leu	Ala	Arg	Lys	Val	Ile	Leu		
		515					520					525					

Leu Ser Gly His Thr Glu Glu Glu Ile Gly Ile Val Glu Ser Gly Ile  
530 535 540

Arg Pro Gly Glu Lys Leu Tyr Glu Glu Leu Leu Ser Thr Glu Glu Arg  
545 550 555 560

Val Ser Glu Gln Ile His Glu Lys Ile Phe Val Gly Arg Val Thr Asn  
565 570 575

Lys Gln Ser Asp Ile Val Asn Ser Phe Ile Asn Gly Leu Leu Gln Lys  
580 585 590

Asp Arg Asn Glu Leu Lys Asn Met Leu Ile Glu Phe Ala Lys Gln Glu  
595 600 605

<210> 40

<211> 200

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS9F

<400> 40

Met Tyr Pro Ile Cys Lys Arg Ile Leu Ala Ile Ile Ile Ser Gly Ile  
1 5 10 15

Ala Ile Val Val Leu Ser Pro Ile Leu Leu Leu Ile Ala Leu Ala Ile  
20 25 30

Lys Leu Asp Ser Lys Gly Pro Val Leu Phe Lys Gln Lys Arg Val Gly  
35 40 45

Lys Asn Lys Ser Tyr Phe Met Ile Tyr Lys Phe Arg Ser Met Tyr Val  
50 55 60

Asp Ala Pro Ser Asp Met Pro Thr His Leu Leu Lys Asp Pro Lys Ala  
65 70 75 80

Met Ile Thr Lys Val Gly Ala Phe Leu Arg Lys Thr Ser Leu Asp Glu  
85 90 95

Leu Pro Gln Leu Phe Asn Ile Phe Lys Gly Glu Met Ala Ile Val Gly  
100 105 110

Pro	Arg	Pro	Ala	Leu	Trp	Asn	Gln	Tyr	Asp	Leu	Ile	Glu	Glu	Arg	Asp
		115					120					125			
Lys	Tyr	Gly	Ala	Asn	Asp	Ile	Arg	Pro	Gly	Leu	Thr	Gly	Trp	Ala	Gln
	130					135					140				
Ile	Asn	Gly	Arg	Asp	Glu	Leu	Glu	Ile	Asp	Glu	Lys	Ser	Lys	Leu	Asp
145					150					155					160
Gly	Tyr	Tyr	Val	Gln	Asn	Met	Ser	Leu	Gly	Leu	Asp	Ile	Lys	Cys	Phe
				165					170					175	
Leu	Gly	Thr	Phe	Leu	Ser	Val	Ala	Arg	Ser	Glu	Gly	Val	Val	Glu	Gly
			180					185					190		
Gly	Thr	Gly	Gln	Lys	Gly	Lys	Gly								
		195					200								

<210> 41

<211> 269

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS2G

<400> 41

Met	Lys	Phe	Ser	Val	Leu	Met	Ser	Val	Tyr	Glu	Lys	Glu	Lys	Pro	Glu
1				5					10					15	
Phe	Leu	Arg	Glu	Ser	Leu	Glu	Ser	Ile	Leu	Val	Asn	Gln	Thr	Met	Ile
			20					25					30		
Pro	Thr	Glu	Val	Val	Leu	Val	Glu	Asp	Gly	Pro	Leu	Asn	Gln	Ser	Leu
		35					40					45			
Tyr	Ser	Ile	Leu	Glu	Glu	Phe	Lys	Ser	Arg	Phe	Ser	Phe	Phe	Lys	Thr
	50					55					60				
Ile	Ala	Leu	Glu	Lys	Asn	Ser	Gly	Leu	Gly	Ile	Ala	Leu	Asn	Glu	Gly
65					70					75					80

Leu	Lys	His	Cys	Asn	Tyr	Glu	Trp	Val	Cys	Thr	Lys	Trp	Ile	Leu	Met	
				85					90					95		
Met	Leu	His	Ile	His	Thr	Arg	Phe	Glu	Lys	Gln	Val	Asn	Phe	Ile	Lys	
			100					105					110			
Gln	Asn	Pro	Thr	Ile	Asp	Ile	Glu	Ile	Asp	Glu	Phe	Leu	Asn	Ser	Thr	
		115					120					125				
Ser	Glu	Ile	Val	Ser	His	Lys	Asn	Val	Pro	Thr	Gln	His	Asp	Glu	Ile	
	130					135					140					
Leu	Lys	Met	Ala	Arg	Arg	Glu	Lys	Ser	Met	Cys	His	Met	Thr	Val	Met	
145					150					155					160	
Phe	Lys	Lys	Lys	Ser	Val	Glu	Arg	Ala	Gly	Gly	Tyr	Gln	Thr	Leu	Pro	
				165					170					175		
Tyr	Val	Glu	Asp	Tyr	Phe	Leu	Trp	Val	Arg	Met	Ile	Ala	Ser	Gly	Ser	
			180					185					190			
Lys	Phe	Ala	Asn	Ile	Asp	Glu	Thr	Leu	Val	Leu	Ala	Arg	Val	Gly	Asn	
		195					200					205				
Gly	Met	Phe	Asn	Arg	Arg	Gly	Asn	Arg	Glu	Gln	Ile	Asn	Ser	Trp	Thr	
	210					215					220					
Leu	Leu	Ile	Glu	Phe	Met	Leu	Ala	Gln	Gly	Ile	Val	Thr	Pro	Leu	Asp	
225					230					235					240	
Val	Phe	Ile	Asn	Gln	Ile	Tyr	Ile	Arg	Val	Phe	Val	Tyr	Met	Pro	Thr	
				245					250					255		
Trp	Ile	Lys	Lys	Leu	Ile	Tyr	Gly	Lys	Ile	Leu	Arg	Lys				
			260					265								

<210> 42

<211> 143

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS9H

<400> 42

Met	Ile	Thr	Val	Leu	Met	Ala	Thr	Tyr	Asn	Gly	Ser	Pro	Phe	Ile	Ile	
1				5					10					15		
Lys	Gln	Leu	Asp	Ser	Ile	Arg	Asn	Gln	Ser	Val	Ser	Ala	Asp	Lys	Val	
			20					25					30			
Ile	Ile	Trp	Asp	Asp	Cys	Ser	Thr	Asp	Asp	Thr	Ile	Lys	Ile	Ile	Lys	
		35					40					45				
Asp	Tyr	Ile	Lys	Lys	Tyr	Ser	Leu	Asp	Ser	Trp	Val	Val	Ser	Gln	Asn	
	50					55					60					
Lys	Ser	Asn	Gln	Gly	His	Tyr	Gln	Thr	Phe	Ile	Asn	Leu	Thr	Lys	Leu	
65					70					75					80	
Val	Gln	Glu	Gly	Ile	Val	Phe	Phe	Ser	Asp	Gln	Asp	Asp	Ile	Trp	Asp	
				85					90					95		
Cys	His	Lys	Ile	Glu	Thr	Met	Leu	Pro	Ile	Phe	Asp	Arg	Glu	Asn	Val	
			100					105					110			
Ser	Met	Val	Phe	Cys	Lys	Ser	Arg	Leu	Ile	Asp	Glu	Asn	Gly	Asn	Ile	
		115					120					125				
Ile	Ser	Ser	Pro	Asp	Thr	Ser	Asp	Arg	Ile	Asn	Thr	Tyr	Ser	Leu		
	130					135					140					

<210> 43

<211> 3738

<212> DNA

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS7

<400> 43

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ttatgggtttc	aacagataaa	gctgttaatc	cgccaaatgt	catgggagcg	actaaacgtg	180

ttgcagaaat gattgtaaca ggtttaaacg agccaggtca gactcaattt gcggcagttc 240  
 gttttgggaa tgttctaggt agtcgtggaa gtgttggtcc gctattcaaa gagcaaatta 300  
 gaaaaggtgg acctgttacg gttaccgact ttaggatgac tcgttatttc atgacgattc 360  
 ctgaggcaag tcgtttgggt atccaagctg gacatttggc aaaaggtgga gaaatctttg 420  
 tcttgatat gggtgagcca gtacaaatcc tggaattggc aagaaaagtt atcttgtaa 480  
 gcggacatac agaggaagaa atcgggattg tagaatctgg aatcagacca ggcgagaaac 540  
 tctacgagga attgttatca acagaagaac gtgtcagcga acagattcat gaaaaaatat 600  
 ttgtgggtcg cgttacaaat aagcagtcgg acattgtcaa ttcatttatt aatggattac 660  
 tccaaaaaga tagaaatgaa ttaaaagata tgttgattga atttgcaaaa caagaataag 720  
 aaagtaaaaa atattttttac tttcctagag tttaaacgat gtttaagtcc taggaagggt 780  
 ggaattgctt tcgtggaggt gatagataga aacctatata tttgtagaag aaaggatatt 840  
 aaactaaagg tgaatcggaa cataaagttt agatagagtt ggtatttaat gccaaacagg 900  
 tgaatgcaac ctctcgctcg ttactaagca ggagatagta aagttgcttg aaagagagtt 960  
 tgttaatcag tataagtagg cttaaagtga aatatatatc tattattatc ggtaatgata 1020  
 ctattattga gaattattgt agtggggata aaaataattt ttggtgattt tatcgtccga 1080  
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 agaagaatga agcaaccagt aaatatcttc agaagataga atcaagaaga ggtgaattat 1260  
 ttattaaatt ctttatggat aagttacttg cgcttatcct attattgcta ttatccccag 1320  
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 aagaacgtgt tacgagatat ggtcgaattt ttagaatatt taagtttaga acaatgattt 1440  
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 ctgatgaaat gtttgcgacg ttacttttac ctgcaggaat tacttcacca gcgagtattg 1680  
 catataagga tgaagatatt gttttagaag aatattgttc tcaaggctat agtcctgatg 1740



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acttttggaaat tattttctgat tttaaagtaa tgattgatac agtaattaaa gtaataaaat 1860  
aggagattaa aatgacaaaa agacaaaata ttccattttc accaccagat attacccaag 1920  
ctgaaattga tgaagttatt gacacactaa aatctgggtg gattacaaca ggaccaaaga 1980  
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caactcctgt gatggttgat attcaaaaaa acagctttga gatggaatat gatgctttgg 2220  
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 gctagaaaaa cagcttgaat ttatgaaaaa taatggatat tcatttactt atcacaattt 3480  
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 gactagaaaa atgatgtaca attacggcta tccagggtgt ttgactttca tgtatgatgc 3600  
 agacaaaatg ggtttaattc agataaaaaga tataaagaaa aataacgatt atgcgatatt 3660  
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<210> 44

<211> 238

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS7E

<400> 44

Ala	Ala	His	Lys	His	Val	Pro	Leu	Met	Glu	Tyr	Asn	Pro	His	Glu	Ala
1				5					10					15	
Val	Lys	Asn	Asn	Ile	Phe	Gly	Thr	Lys	Asn	Val	Ala	Glu	Ala	Ala	Lys
			20					25					30		
Thr	Ala	Lys	Val	Ala	Lys	Phe	Val	Met	Val	Ser	Thr	Asp	Lys	Ala	Val
		35					40					45			
Asn	Pro	Pro	Asn	Val	Met	Gly	Ala	Thr	Lys	Arg	Val	Ala	Glu	Met	Ile
	50					55					60				
Val	Thr	Gly	Leu	Asn	Glu	Pro	Gly	Gln	Thr	Gln	Phe	Ala	Ala	Val	Arg
65				70					75					80	
Phe	Gly	Asn	Val	Leu	Gly	Ser	Arg	Gly	Ser	Val	Val	Pro	Leu	Phe	Lys
			85					90						95	

Glu Gln Ile Arg Lys Gly Gly Pro Val Thr Val Thr Asp Phe Arg Met  
 100 105 110  
 Thr Arg Tyr Phe Met Thr Ile Pro Glu Ala Ser Arg Leu Val Ile Gln  
 115 120 125  
 Ala Gly His Leu Ala Lys Gly Gly Glu Ile Phe Val Leu Asp Met Gly  
 130 135 140  
 Glu Pro Val Gln Ile Leu Glu Leu Ala Arg Lys Val Ile Leu Leu Ser  
 145 150 155 160  
 Gly His Thr Glu Glu Glu Ile Gly Ile Val Glu Ser Gly Ile Arg Pro  
 165 170 175  
 Gly Glu Lys Leu Tyr Glu Glu Leu Leu Ser Thr Glu Glu Arg Val Ser  
 180 185 190  
 Glu Gln Ile His Glu Lys Ile Phe Val Gly Arg Val Thr Asn Lys Gln  
 195 200 205  
 Ser Asp Ile Val Asn Ser Phe Ile Asn Gly Leu Leu Gln Lys Asp Arg  
 210 215 220  
 Asn Glu Leu Lys Asp Met Leu Ile Glu Phe Ala Lys Gln Glu  
 225 230 235

<210> 45

<211> 232

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS7F

<400> 45

Met Thr Arg Val Glu Leu Ile Thr Arg Glu Phe Phe Lys Lys Asn Glu  
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 Ala Thr Ser Lys Tyr Phe Gln Lys Ile Glu Ser Arg Arg Gly Glu Leu  
 20 25 30  
 Phe Ile Lys Phe Phe Met Asp Lys Leu Leu Ala Leu Ile Leu Leu Leu  
 35 40 45

Leu Leu Ser Pro Val Ile Ile Ile Leu Ala Ile Trp Ile Lys Leu Asp  
 50 55 60  
 Ser Lys Gly Pro Ile Phe Tyr Arg Gln Glu Arg Val Thr Arg Tyr Gly  
 65 70 75 80  
 Arg Ile Phe Arg Ile Phe Lys Phe Arg Thr Met Ile Ser Asp Ala Asp  
 85 90 95  
 Lys Val Gly Ser Leu Val Thr Val Gly Gln Asp Asn Arg Ile Thr Lys  
 100 105 110  
 Val Gly His Ile Ile Arg Lys Tyr Arg Leu Asp Glu Val Pro Gln Leu  
 115 120 125  
 Phe Asn Val Leu Met Gly Asp Met Ser Phe Val Gly Val Arg Pro Glu  
 130 135 140  
 Val Gln Lys Tyr Val Asn Gln Tyr Thr Asp Glu Met Phe Ala Thr Leu  
 145 150 155 160  
 Leu Leu Pro Ala Gly Ile Thr Ser Pro Ala Ser Ile Ala Tyr Lys Asp  
 165 170 175  
 Glu Asp Ile Val Leu Glu Glu Tyr Cys Ser Gln Gly Tyr Ser Pro Asp  
 180 185 190  
 Glu Ala Tyr Val Gln Lys Val Leu Pro Glu Lys Met Lys Tyr Asn Leu  
 195 200 205  
 Glu Tyr Ile Arg Asn Phe Gly Ile Ile Ser Asp Phe Lys Val Met Ile  
 210 215 220  
 Asp Thr Val Ile Lys Val Ile Lys  
 225 230

<210> 46

<211> 404

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS7G

<400> 46

Met	Thr	Lys	Arg	Gln	Asn	Ile	Pro	Phe	Ser	Pro	Pro	Asp	Ile	Thr	Gln	1	5	10	15
Ala	Glu	Ile	Asp	Glu	Val	Ile	Asp	Thr	Leu	Lys	Ser	Gly	Trp	Ile	Thr	20	25	30	
Thr	Gly	Pro	Lys	Thr	Lys	Glu	Leu	Glu	Arg	Arg	Leu	Ser	Val	Phe	Thr	35	40	45	
Gly	Thr	Asn	Lys	Thr	Val	Cys	Leu	Asn	Ser	Ala	Thr	Ala	Gly	Leu	Glu	50	55	60	
Leu	Val	Leu	Arg	Ile	Leu	Gly	Val	Gly	Pro	Gly	Asp	Glu	Val	Ile	Val	65	70	75	
Pro	Ala	Met	Thr	Tyr	Thr	Ala	Ser	Cys	Ser	Val	Ile	Thr	His	Val	Gly	85	90	95	
Ala	Thr	Pro	Val	Met	Val	Asp	Ile	Gln	Lys	Asn	Ser	Phe	Glu	Met	Glu	100	105	110	
Tyr	Asp	Ala	Leu	Glu	Lys	Ala	Ile	Thr	Pro	Lys	Thr	Lys	Val	Ile	Ile	115	120	125	
Pro	Val	Asp	Leu	Ala	Gly	Ile	Pro	Cys	Asp	Tyr	Asp	Lys	Ile	Tyr	Thr	130	135	140	
Ile	Val	Glu	Asn	Lys	Arg	Ser	Leu	Tyr	Val	Ala	Ser	Asp	Asn	Lys	Trp	145	150	155	
Gln	Lys	Leu	Phe	Gly	Arg	Val	Ile	Ile	Leu	Ser	Asp	Ser	Ala	His	Ser	165	170	175	
Leu	Gly	Ala	Ser	Tyr	Lys	Gly	Lys	Pro	Ala	Gly	Ser	Leu	Ala	Asp	Phe	180	185	190	
Thr	Ser	Phe	Ser	Phe	His	Ala	Val	Lys	Asn	Phe	Thr	Thr	Ala	Glu	Gly	195	200	205	
Gly	Ser	Val	Thr	Trp	Arg	Ser	His	Pro	Asp	Leu	Asp	Asp	Glu	Glu	Met	210	215	220	
Tyr	Lys	Glu	Phe	Gln	Ile	Tyr	Ser	Leu	His	Gly	Gln	Thr	Lys	Asp	Ala	225	230	235	
Leu	Ala	Lys	Thr	Gln	Leu	Gly	Ser	Trp	Glu	Tyr	Asp	Ile	Val	Ile	Pro	245	250	255	
Gly	Tyr	Lys	Cys	Asn	Met	Thr	Asp	Ile	Met	Ala	Gly	Ile	Gly	Leu	Val	260	265	270	

Gln Leu Glu Arg Tyr Pro Ser Leu Leu Asn Arg Arg Arg Glu Ile Ile  
           275                                  280                                  285  
 Glu Lys Tyr Asn Ala Gly Phe Glu Gly Thr Ser Ile Lys Pro Leu Val  
           290                                  295                                  300  
 His Leu Thr Glu Asp Lys Gln Ser Ser Met His Leu Tyr Ile Thr His  
 305                                  310                                  315                                  320  
 Leu Gln Gly Tyr Thr Leu Glu Gln Arg Asn Glu Val Ile Gln Lys Met  
                                   325                                  330                                  335  
 Ala Glu Ala Gly Ile Ala Cys Asn Val His Tyr Lys Pro Leu Pro Leu  
                                   340                                  345                                  350  
 Leu Thr Ala Tyr Lys Asn Leu Gly Phe Glu Met Lys Asp Phe Pro Asn  
                                   355                                  360                                  365  
 Ala Tyr Gln Tyr Phe Glu Asn Glu Val Thr Leu Pro Leu His Thr Asn  
                                   370                                  375                                  380  
 Leu Ser Asp Glu Asp Val Glu Tyr Val Ile Glu Met Phe Leu Lys Ile  
 385                                  390                                  395                                  400  
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<210> 47

<211> 210

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> CPS7H

<400> 47

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                                   20                                  25                                  30  
 Val Leu Asp Gln Thr His Gln Asn Trp Glu Leu Ile Ile Val Asp Asp  
                                   35                                  40                                  45

Cys	Ser	Asn	Asp	Glu	Thr	Glu	Lys	Val	Val	Ser	His	Phe	Lys	Asp	Ser
50						55					60				
Arg	Ile	Lys	Phe	Phe	Lys	Asn	Ser	Asn	Asn	Leu	Gly	Ala	Ala	Leu	Thr
65					70					75					80
Arg	Asn	Lys	Ala	Leu	Arg	Lys	Ala	Arg	Gly	Arg	Trp	Ile	Ala	Phe	Leu
				85					90					95	
Asp	Ser	Asp	Asp	Leu	Trp	His	Pro	Ser	Lys	Leu	Glu	Lys	Gln	Leu	Glu
			100					105					110		
Phe	Met	Lys	Asn	Asn	Gly	Tyr	Ser	Phe	Thr	Tyr	His	Asn	Phe	Glu	Lys
		115					120					125			
Ile	Asp	Glu	Ser	Ser	Gln	Ser	Leu	Arg	Val	Leu	Val	Ser	Gly	Pro	Ala
	130					135					140				
Ile	Val	Thr	Arg	Lys	Met	Met	Tyr	Asn	Tyr	Gly	Tyr	Pro	Gly	Cys	Leu
145					150					155					160
Thr	Phe	Met	Tyr	Asp	Ala	Asp	Lys	Met	Gly	Leu	Ile	Gln	Ile	Lys	Asp
				165					170					175	
Ile	Lys	Lys	Asn	Asn	Asp	Tyr	Ala	Ile	Leu	Leu	Gln	Leu	Cys	Lys	Lys
			180					185					190		
Tyr	Asp	Cys	Tyr	Leu	Leu	Asn	Glu	Ser	Leu	Ala	Ser	Tyr	Arg	Ile	Arg
		195					200					205			
Lys	Lys														
	210														

<210> 48

<211> 101

<212> DNA

<213> Streptococcus suis

<220>

<221> misc\_feature

<222> (1)..(101)

<223> N may be any nucleotide

<220>

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<221> misc_feature
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<400> 48
aagggcacct ctataaactc ccaaaattgc gaatttggag ttacgaaagc cttgttaa 60
caancatttt aaattttaga aaattagttt ttagagctcc c 101

<210> 49
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<212> DNA
<213> Streptococcus suis
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<222> (1)..(101)
<223> N may be any nucleotide
<220>
<221> misc_feature
<223> 100 base pair repeat within CPS2M
<400> 49
ggcgccacct ctataaattc ccaaaattgc gaatttcgag ttacgaaagc cttgttaa 60
caancatctt aaattttaga aaattagttt ttagaggctcc c 101

<210> 50
<211> 101
<212> DNA
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<220>
<221> misc_feature

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<223> 100 base pair repeat between CPS2O and CPS2P

<400> 50

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caaacatttt aaattttaga aaattagttt ttagaggtcc c 101
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<210> 51

<211> 120

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> N-terminal part of CPS2J

<400> 51

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Leu Arg Glu Cys Leu Asp Ser Ile Ile Ser Gln Ser Tyr Thr Asn Leu
          20          25          30
Glu Ile Leu Leu Ile Asp Asp Gly Ser Ser Asp Ser Ser Thr Asp Ile
          35          40          45
Cys Leu Glu Tyr Ala Glu Gln Asp Gly Arg Ile Lys Leu Phe Arg Leu
          50          55          60
Pro Asn Gly Gly Val Ser Asn Ala Arg Asn Tyr Gly Ile Lys Asn Ser
65          70          75          80
Thr Ala Asn Tyr Ile Met Phe Val Asp Ser Asp Asp Ile Val Asp Gly
          85          90          95
Asn Ile Val Glu Ser Leu Tyr Thr Cys Leu Lys Glu Asn Asp Ser Asp
          100          105          110
Leu Ser Gly Gly Leu Leu Ala Thr
          115          120
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<210> 52

<211> 120

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> N-terminal part of CPS2K

<220>

<221> misc\_feature

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<223> Xaa may be any amino acid

<400> 52

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1				5					10					15	

Leu	Ser	Lys	Cys	Ile	Asn	Ser	Ile	Val	Asn	Gln	Thr	Tyr	Lys	His	Ile
			20					25					30		

Glu	Leu	Leu	Val	Asn	Asp	Gly	Ser	Ser	Thr	Asp	Asn	Ser	Glu	Glu	Ile
			35				40					45			

Cys	Leu	Ala	Tyr	Ala	Lys	Lys	Asp	Ser	Arg	Ile	Arg	Tyr	Phe	Lys	Lys
	50					55					60				

Glu	Asn	Gly	Gly	Leu	Ser	Asp	Ala	Arg	Asn	Tyr	Gly	Ile	Ser	Arg	Ala
65					70					75					80

Lys	Gly	Asp	Tyr	Leu	Ala	Phe	Ile	Asp	Ser	Asp	Asp	Phe	Ile	His	Ser
				85					90					95	

Glu	Phe	Ile	Gln	Arg	Leu	Xaa	His	Glu	Ala	Ile	Glu	Arg	Glu	Asn	Ala
			100					105					110		

Leu	Xaa	Xaa	Val	Ala	Val	Ala	Gly
			115				120

<210> 53

<211> 419

<212> PRT

<213> Streptococcus suis

<220>

<221> misc\_feature

<223> ORF2Y

<400> 53

Met	Lys	Lys	Tyr	Gln	Val	Ile	Ile	Gln	Asp	Ile	Leu	Thr	Gly	Ile	Glu	
1				5					10					15		
Glu	His	Arg	Phe	Lys	Arg	Gly	Glu	Lys	Leu	Pro	Ser	Ile	Arg	Gln	Leu	
			20					25					30			
Arg	Glu	Gln	Tyr	His	Cys	Ser	Lys	Asp	Thr	Val	Gln	Lys	Ala	Met	Leu	
		35					40					45				
Glu	Leu	Lys	Tyr	Gln	Asn	Lys	Ile	Tyr	Ala	Val	Glu	Lys	Ser	Gly	Tyr	
	50					55					60					
Tyr	Ile	Leu	Glu	Asp	Arg	Asp	Phe	Gln	Asp	His	Thr	Cys	Arg	Ala	Gln	
65					70					75					80	
Ser	Tyr	Arg	Leu	Ser	Arg	Ile	Thr	Tyr	Glu	Asp	Phe	Arg	Ile	Cys	Leu	
				85					90					95		
Lys	Glu	Ser	Leu	Ile	Gly	Arg	Glu	Asn	Tyr	Leu	Phe	Asn	Tyr	Tyr	His	
			100					105					110			
Gln	Gln	Glu	Gly	Leu	Ala	Glu	Leu	Ile	Ser	Ser	Val	Gln	Ser	Leu	Leu	
		115					120					125				
Met	Asp	Tyr	His	Val	Tyr	Thr	Lys	Lys	Asp	Gln	Leu	Val	Ile	Thr	Ala	
	130					135					140					
Gly	Ser	Gln	Gln	Ala	Leu	Tyr	Ile	Leu	Thr	Gln	Met	Glu	Thr	Leu	Ala	
145					150					155					160	
Gly	Lys	Thr	Glu	Ile	Leu	Ile	Glu	Asn	Pro	Thr	Tyr	Ser	Arg	Met	Ile	
			165						170					175		
Glu	Leu	Ile	Arg	His	Gln	Gly	Ile	Pro	Tyr	Gln	Thr	Ile	Glu	Arg	Asn	
			180					185					190			
Leu	Asp	Gly	Ile	Asp	Leu	Glu	Glu	Leu	Glu	Ser	Ile	Phe	Gln	Thr	Gly	
		195					200						205			

Lys	Ile	Lys	Phe	Phe	Tyr	Thr	Ile	Pro	Arg	Leu	His	Asn	Pro	Leu	Gly
210						215					220				
Ser	Thr	Tyr	Asp	Ile	Ala	Thr	Lys	Thr	Ala	Ile	Val	Lys	Leu	Ala	Lys
225					230					235					240
Gln	Tyr	Asp	Val	Tyr	Ile	Ile	Glu	Asp	Asp	Tyr	Leu	Ala	Asp	Phe	Asp
				245					250					255	
Ser	Ser	His	Ser	Leu	Pro	Leu	His	Tyr	Leu	Asp	Thr	Asp	Asn	Arg	Val
			260					265					270		
Ile	Tyr	Ile	Lys	Ser	Phe	Thr	Pro	Thr	Leu	Phe	Pro	Ala	Leu	Arg	Ile
		275					280					285			
Gly	Ala	Ile	Ser	Leu	Pro	Asn	Gln	Leu	Arg	Asp	Ile	Phe	Ile	Lys	His
	290					295					300				
Lys	Ser	Leu	Ile	Asp	Tyr	Asp	Thr	Asn	Leu	Ile	Met	Gln	Lys	Ala	Leu
305					310					315					320
Ser	Leu	Tyr	Ile	Asp	Asn	Gly	Met	Phe	Ala	Arg	Asn	Thr	Gln	His	Leu
				325					330					335	
His	His	Ile	Tyr	His	Ala	Gln	Trp	Asn	Lys	Ile	Lys	Asp	Cys	Leu	Glu
			340					345					350		
Lys	Tyr	Ala	Leu	Asn	Ile	Pro	Tyr	Arg	Ile	Pro	Lys	Gly	Ser	Val	Thr
		355					360					365			
Phe	Gln	Leu	Ser	Lys	Gly	Ile	Leu	Ser	Pro	Ser	Ile	Gln	His	Met	Phe
	370					375					380				
Gly	Lys	Cys	Tyr	Tyr	Phe	Ser	Gly	Gln	Lys	Ala	Asp	Phe	Leu	Gln	Ile
385					390					395					400
Phe	Phe	Glu	Gln	Asp	Phe	Ala	Asp	Lys	Leu	Glu	Gln	Phe	Val	Arg	Tyr
				405					410					415	
Leu	Asn	Glu													

**TABLE 1.                    Bacterial strains and plasmids**

strain/plasmid	relevant characteristics	source/reference
<b>Strain</b>		
<i>E. coli</i>		
CC118	PhoA <sup>-</sup>	(28)
XL2 blue	Stratagene	
<i>E. coli</i>		
XL2 blue	Stratagene	
<i>S. suis</i>		
10	virulent serotype 2 strain	(49)
3	serotype 2	(63)
17	serotype 2	(63)
735	reference strain serotype 2	(63)
T15	serotype 2	(63)
6555	reference strain serotype 1	(63)
6388	serotype 1	(63)
6290	serotype 1	(63)
5637	serotype 1	(63)
5673	serotype 1/2	(63)
5679	serotype 1/2	(63)
5928	serotype 1/2	(63)
5934	serotype 1/2	(63)
5209	reference strains serotype 1/2	(63)
5218	reference strain serotype 9	(63)

strain/plasmid	relevant characteristics	source/reference
5973	serotype 9	(63)
6437	serotype 9	(63)
6207	serotype 9	(63)
reference strains	serotypes 1-34	(9, 56, 14)
<i>S. suis</i>		
10	virulent serotype 2 strain	(51)
10cpsB	isogenic cpsB mutant of strain 10	this work
10cpsEF	isogenic cpsEF mutant of strain 10	this work
<b>Plasmid</b>		
pKUN19	replication functions pUC, Amp <sup>R</sup>	(23)
pGEM7Zf (+)	replication functions pUC, Amp <sup>R</sup>	Promega Corp.
pIC19R	replication functions pUC, Amp <sup>R</sup>	(29)
pIC20R	replication functions pUC, Amp <sup>R</sup>	(29)
pIC-spc	pIC19R containing spc <sup>R</sup> gene of pDL282	labcollection
pDL282	replication functions of pBR 322 and pVT736-1, Amp <sup>R</sup> , Spc <sup>R</sup>	(43)
pPHOS2	pIC-spc containing the truncated <i>phoA</i> gene of pPHO7 as a <i>Pst</i> I- <i>Bam</i> HI fragment	this work
pPHO7	contains truncated <i>phoA</i> gene	(15)
pPHOS7	pPHOS2 containing chromosomal <i>S. suis</i> DNA	this work
pCPS6	pKUN19 containing 6 kb <i>Hind</i> III fragment of <i>cps</i> operon	this work (Fig. 1)
pCPS7	pKUN19 containing 3,5 kb <i>Eco</i> RI- <i>Hind</i> III fragment of <i>cps</i> operon	this work (Fig. 1)
pCPS11	pCPS7 in which 0.4 kb <i>Pst</i> I- <i>Bam</i> HI fragment of <i>cpsB</i> gene is replaced by Spc <sup>R</sup> gene of pIC-spec	this work (Fig. 1)

strain/plasmid	relevant characteristics	source/reference
pCPS17	pKUN19 containing 3.1 kb <i>Kpn</i> I fragment of <i>cps</i> operon	this work (Fig. 1)
pCPS18	pKUN19 containing 1.8 kb <i>Sna</i> BI fragment of <i>cps</i> operon	this work (Fig. 1)
pCPS20	pKUN19 containing 3.3 kb <i>Xba</i> I- <i>Hind</i> III fragment of <i>cps</i> operon	this work (Fig. 1)
pCPS23	pGEM7Zf (+) containing 1.5 kb <i>Mlu</i> I fragment of <i>cps</i> operon	this work (Fig. 1)
pCPS25	pIC20R containing 2.5 kb <i>Kpn</i> I- <i>Sal</i> I fragment of pCPS17	this work (Fig. 1)
pCPS26	pKUN19 containing 3.0 kb <i>Hind</i> III fragment of <i>cps</i> operon	this work (Fig. 1)
pCPS27	pCPS25 containing 2.3 kb <i>Xba</i> I (blunt) - <i>Cl</i> aI fragment of pCPS20	this work (Fig. 1)
pCPS28	pCPS27 containing the 1.2 kb <i>Pst</i> I- <i>Xho</i> I <i>Spc</i> <sup>R</sup> gene of pIC-spc	this work (Fig. 1)
pCPS29	pKUN19 containing 2.2 kb <i>Sac</i> I- <i>Pst</i> I fragment of <i>cps</i> operon	this work (Fig. 1)
pCPS1-1	pKUN19 containing 5 kb <i>Eco</i> RV fragment of <i>cps</i> operon of type 1	this work (Fig. 1)
pCPS1-2	pKUN19 containing 2.2 kb <i>Hind</i> III fragment of <i>cps</i> operon of type 1	this work (Fig. 1)
pCPS9-1	pKUN19 containing 1 kb <i>Hind</i> III- <i>Xba</i> I fragment of <i>cps</i> operon of serotype 9	this work (Fig. 1)
pCPS9-2	pKUN19 containing 4.0 kb <i>Xba</i> I- <i>Xba</i> I fragment of <i>cps</i> operon of serotype 9	this work (Fig. 1)

Amp<sup>R</sup>: ampicillin resistant  
 Spc<sup>R</sup>: spectinomycin resistant  
 cps: capsular polysaccharide

**Table 2. Properties of Orfs in the cps locus of *S. suis* serotype 2 and similarities to gene products of other bacteria**

<b>ORF</b>	<b>nucleotide position in sequence</b>	<b>number of amino acids</b>	<b>GC%</b>	<b>proposed function of gene product<sup>1</sup></b>	<b>similar gene product (% identity)</b>
Orf2Z	1- 719	240	44	Unknown	<i>B. subtilis</i> YitS (26%)
Orf2Y	2079- 822	419	38	Transcription regulation	<i>B. subtilis</i> YcxD (39%)
Orf2X	2202- 2934	244	39	Unknown	<i>H. influenzae</i> YAAA (24%)
Cps2A	3041- 4484	481	39	Regulation	<i>S. pneumoniae</i> Cps19fA (58%)
Cps2B	4504- 5191	229	40	Chain length determination	<i>S. pneumoniae</i> type 3 Orf1 (58%)
Cps2C	5203- 5878	225	40	Chain length determination/ Export	<i>S. pneumoniae</i> Cps23fD (63%)
Cps2D	5919- 6648	243	38	Unknown	<i>S. pneumoniae</i> CpsB (62%)
Cps2E	6675- 8052	459	33	Glycosyltransferase	<i>S. pneumoniae</i> Cps14E (56%)
Cps2F	8089- 9256	389	32	Glycosyltransferase	<i>S. pneumoniae</i> Cps23fT
Cps2G	9262-10417	385	36	Glycosyltransferase	<i>S. thermophilus</i> EpsF (25%)
Cps2H	10808-12176	457	31	Glycosyltransferase	<i>S. mutans</i> RGPEC, <sup>N</sup> (29%)
Cps2I	12213-13443	410	29	CP polymerase	<i>S. pneumoniae</i> Cps23fI (48%)
Cps2J	13583-14579	332	29	Glycosyltransferase	<i>S. pneumoniae</i> Cps14J (31%)
Cps2K	14574-15576	334	37	Glycosyltransferase	<i>S. pneumoniae</i> Cps14J (40%)
“Cps2L”	15618-16635	103	37	Unknown	-
“Cps2M”	16811-17322	-	38	-	<i>S. agalactiae</i> CpsF <sup>N</sup> (77%) <i>E. coli</i> NeuA, <sup>N</sup> (47%)



ORF	nucleotide position in sequence	number of amino acids	GC%	proposed function of gene product <sup>1</sup>	similar gene product (% identity)
“Cps2N”	17559-18342	-	39	-	<i>S. agalactiae</i> CpsJ (43%)
Cps2O	18401-19802	476	40	Repeat unit transporter	<i>S. agalactiae</i> CpsK (41%)
Cps2P	20327-21341	338	39	Sialic acid synthesis	<i>S. agalactiae</i> NeuB (80%) <i>E. coli</i> NeuB (59%)
Cps2Q	21355-21865	170	42	Sialic acid synthesis	<i>S. agalactiae</i> NeuC <sup>N</sup> (61%) <i>E. coli</i> NeuC <sup>N</sup> (54%)
Cps2R	21933-22483	184	40	Sialic acid synthesis	<i>S. agalactiae</i> NeuC <sup>C</sup> (55%) <i>E. coli</i> NeuC <sup>C</sup> (40%)
Cps2S	22501-23125	208	42	Sialic acid synthesis	<i>E. coli</i> NeuD (32%)
Cps2T	23136-24366	395	40	CMP-NeuNAc synthetase	<i>S. agalactiae</i> CpsF (49%) <i>E. coli</i> NeuA (34%)
“Orf2U”	24566-25488	168	42	Transposase	<i>S. thermophilus</i> IS1194 (51%)
“Orf2V”	25691-26281	116	37	Transposase	<i>S. pneumoniae</i> orf1 (85%)

<sup>1</sup> Predicted by sequence similarity

<sup>N</sup> Similarity refers to the amino-terminal part of the gene product

<sup>C</sup> Similarity refers to the carboxy-terminal part of the gene product

ORFS between “ ” are truncated or non-functional as the result of frame-shift or point mutations

**TABLE 3. Properties of ORFs in the cps gene of *S. suis* serotypes 1 and 9 and similarities to gene products of other bacteria**

ORF	nucleotide position in sequence	G + C%	number of amino acids	predicted mol. mass (kDa)	predicted pI	proposed function of gene product <sup>1</sup>	similar gene product (%) identity)	reference/ accession nr.
Cps1E <sup>2</sup>	1-1363	34%	454	52.2	8.0	Glucosyltransferase	<i>Streptococcus suis</i> Cps2E (86%)	(26)
Cps1F	1374-1821	33%	149	17.3	8.2	Unknown	<i>Streptococcus pneumoniae</i> Cps14E (48%)	(12)
Cps1G	1823-2315	25%	164	19.5	7.5	Glycosyltransferase	<i>Streptococcus pneumoniae</i> Cps14F (83%)	(14)
Cps1H	3035-4202	24%	389	45.5	8.4	CP polymerase	<i>Streptococcus pneumoniae</i> Cps14G (50%)	(14)
Cps1I	4917-					Glycosyltransferase	<i>Streptococcus pneumoniae</i> Cps14H (30%)	(14)
							<i>Streptococcus pneumoniae</i> Cps14J (38%)	(13)
							<i>Lactococcus lactis</i> EpsG (31%)	(29)
							<i>Streptococcus thermophilus</i> EpsI (33%)	(28)
Cps1J						Glycosyltransferase	<i>Streptococcus pneumoniae</i> Cps14J ( %)	(13)

ORF	nucleotide position in sequence	G + C%	number of amino acids	predicted mol. mass (kDa)	predicted pI	proposed function of gene product <sup>1</sup>	similar gene product (% identity)	reference/ accession nr.
Cps1K <sup>3</sup>		37%	278	32.5	7.8	Glycosyltransferase	<i>Streptococcus pneumoniae</i> Cps14J (44%)	(13)
Cps9D <sup>2</sup>	1-646	37%	215	24.9	8.1	Unknown	<i>Streptococcus suis</i> Cps2D (89%)	(26)
Cps9E	680-					Glycosyltransferase	<i>Staphylococcus aureus</i> Cap1D (27%)	(18)
Cps9F		36%	200	22.3	8.2	Glycosyltransferase	<i>Staphylococcus aureus</i> Cap5M (52%)	(17)
Cps9G		35%	269	31.5	8.0	Unknown	<i>Actinobacillus actinomycetemcomitans</i> (43%)	(AB002668_4)
Cps9H <sup>3</sup>		30%	143	16.5	7.2	Unknown	<i>Haemophilus influenzae</i> Lsg (43%)	(O05081)
							<i>Yersinia enterocolitica</i> RfbB (28%)	(33)

<sup>1</sup> Predicted by sequence similarity

<sup>2</sup> N-terminal part of protein is lacking

<sup>3</sup> C-terminal part of protein is lacking

Table 4. Hybridization of serotype 2 cps genes and neighboring sequences with chromosomal DNA of serotypes

serotypes	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	1/2
DNA probes																																			
<i>orf2Z</i>	+	+	+	+	+	+	+	+	+	+	+	+	±	+	+	+	+	+	+	+	-	+	-	+	+	+	+	+	+	+	+	+	-	-	+
<i>orf2Y</i>	+	+	+	+	+	+	+	+	+	+	+	±	±	+	+	+	+	+	+	±	+	±	+	+	+	+	+	+	+	+	+	+	-	-	+
<i>orf2X</i>	+	+	+	+	+	+	+	+	+	+	+	±	±	+	+	+	+	+	+	-	+	-	+	+	+	+	+	+	+	+	+	+	-	-	+
<i>cps2A</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	-	+	-	+	+	+	+	+	+	+	+	+	+	-	-	+
<i>cps2B</i>	+	+	+	+	+	+	+	+	+	+	-	-	±	+	-	±	±	±	±	±	-	-	+	+	+	+	+	-	-	+	±	+	-	±	+
<i>cps2C</i>	+	+	+	+	+	+	+	+	+	+	+	-	±	+	-	±	-	-	-	-	-	-	+	+	+	+	+	±	+	-	+	+	-	-	+
<i>cps2D</i>	+	+	+	+	+	+	+	+	+	+	+	±	±	+	-	±	+	+	+	±	±	-	+	+	+	+	+	+	+	±	+	+	-	-	+
<i>cps2E</i>	+	+	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	+
<i>cps2F</i>	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+
<i>cps2G</i>	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	±	+	
<i>cps2H</i>	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+
<i>cps2I</i>	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+
<i>cps2J</i>	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+
<i>cps2K</i>	+	+	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+
“ <i>cps2L</i> ”	+	+	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+
“ <i>cps2M</i> ”	+	+	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	+
<i>cps2N</i> ”	+	+	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+
<i>cps2O</i>	+	+	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	+
<i>cps2P</i>	+	+	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	+
<i>cps2Q</i>	+	+	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	+
<i>cps2R</i>	+	+	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	+
<i>cps2S</i>	+	+	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	+

serotypes	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	1/2
<i>cps2T</i>	+	+	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	+
“ <i>orf2U</i> ”	+	+	-	-	-	-	+	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	+	-	-	+	-	-	-	+	-	+	+	+
“ <i>orf2V</i> ”	+	+	±	±	±	-	±	-	-	-	-	-	-	+	+	-	+	±	-	-	-	±	+	-	-	+	-	-	-	+	+	±	±	+	+
<i>100-bp repeat</i>	+	+	-	-	-	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-	-	+	-	+	-	-	+	-	-	-	-	+	+	+	+
<i>16SrRNA</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+

Table 5. Hybridization of serotypes 1 and 9 *cps* genes with chromosomal DNA of other *S. suis* serotypes

Serotype	DNA probes									
	<i>cps1E</i>	<i>cps1F</i>	<i>cps1G</i>	<i>cps1H</i>	<i>cps1I</i>	<i>cps9E</i>	<i>cps9F</i>	<i>cps9G</i>	<i>cps9H</i>	16rRNA
1	+	+	+	+	+	-	-	-	-	-
2	+	-	-	-	-	-	-	-	-	+
3	-	-	-	+	-	+	-	-	-	+
4	-	-	-	+	-	+	-	-	-	+
5	-	-	-	+	-	+	-	-	-	+
6	-	-	-	-	-	-	-	-	-	+
7	-	-	-	+	-	+	-	-	-	+
8	-	-	-	-	-	-	-	-	-	+
9	-	-	-	+	-	+	+	+	+	+
10	-	-	-	+	-	+	+	-	-	+
11	-	-	-	+	-	+	±	-	-	+
12	-	-	-	±	-	+	±	-	-	+
13	-	-	-	+	-	+	-	-	-	+
14	+	+	+	+	+	-	-	-	-	+
15	-	-	-	-	-	-	-	-	-	+

Serotype	<i>cps1E</i>	<i>cps1F</i>	<i>cps1G</i>	<i>cps1H</i>	<i>cps1I</i>	<i>cps9E</i>	<i>cps9F</i>	<i>cps9G</i>	<i>cps9H</i>	16rRNA
16	-	-	-	-	-	-	-	-	-	+
17	-	-	-	+	-	+	-	-	-	+
18	-	-	-	+	-	+	-	-	-	+
19	-	-	-	+	-	+	-	-	-	+
20	-	-	-	-	-	-	-	-	-	+
21	-	-	-	+	-	+	±	-	-	+
22	-	-	-	-	-	-	-	-	-	+
23	-	-	-	+	-	+	-	-	-	+
24	-	-	-	+	-	+	+	-	-	+
25	-	-	-	-	-	-	-	-	-	+
26	-	-	-	-	-	-	±	-	-	+
27	+	-	-	-	-	-	-	-	-	+
28	-	-	-	+	-	+	±	-	-	+
29	-	-	-	+	-	+	-	-	-	+
30	-	-	-	+	-	+	±	-	-	+
31	-	-	-	+	-	+	-	-	-	+
32	-	-	-	-	-	-	-	-	-	+
33	-	-	-	-	-	-	±	-	-	+
34	-	-	-	-	-	-	-	-	-	+
½	+	-	-	-	-	-	-	-	-	+

TABLE 6. Virulence of wild-type and capsular mutant *S. suis* strains in germfree pigs

<i>S. suis</i> strains <sup>1</sup>	pigs/ group (n)	mortality <sup>2</sup> (%)	morbidity <sup>3</sup> (%)	clinical index of the group	spec symptoms <sup>5</sup>	non-spec. symptoms <sup>6</sup>	fever index <sup>7</sup>	leucocyte index <sup>8</sup>	isolation of <i>S. suis</i> in pigs (n) per group in	CNS	serosae	joints
10	4	100	100		11	88	43	44	2	3		4
10cpsB	4	0	0		0	10	1	3	1	3		2
10cpsEF	4	0	0		0	0	1	0	1	3		2

<sup>1</sup> strain10 in the wild-type strain, strains 10cpsB and 10cpsEF are isogenic capsular mutant strains

<sup>2</sup> piglets which died spontaneously or had to be killed for animal welfare reasons

<sup>3</sup> only considering pigs with specific symptoms

<sup>4</sup> clinical index: % of observations which matched the described criteria

<sup>5</sup> specific symptoms: ataxia, lameness on at lest one joint, stiffness

<sup>6</sup> non-specific symptoms: inappetance, depression

<sup>7</sup> % of observations in the experimental group with a body temperature > 40°C

<sup>8</sup> % of blood samples in the group in which number of granulocytes > 10<sup>10</sup>/l



**Table 7. Bacterial strains and plasmids**

strain/plasmid	relevant characteristics
<b>Strain</b>	
<i>E. coli</i>	
XL2 blue	
<i>S. suis</i>	
reference strains	
5667	serotypes 1-34
7037	serotype 7, tonsil (1993)
7044	serotype 7, organs (1994)
7068	serotype 7, brains (1994)
7646	serotype 7 (1994)
7744	serotype 7, lungs (1996)
7759	serotype 7, joints (1996)
8169	serotype 7 (1997)
15913	serotype 7, meninges (1998)
<b>Plasmid</b>	
pKUN19	replication functions pUC, Amp <sup>R</sup>
pGEM7Zf (+)	replication functions pUC, Amp <sup>R</sup>
pCPS9-1	pKUN19 containing 1 kb <i>HindIII</i> - <i>XbaI</i> fragment of <i>cps</i> operon of serotype 9
pCPS9-2	pKUN19 containing 4.09 kb <i>XbaI</i> - <i>XbaI</i> fragment of <i>cps</i> operon of serotype 9
pCPS7-1	pKUN19 containing 1.6-kb <i>PstI</i> fragment of <i>cps</i> operon of type 7
pCPS7-2	pGEM7 containing 2.7-kb <i>ScaI</i> - <i>Clal</i> fragment of <i>cps</i> operon of type 7

Amp<sup>R</sup>: ampicillin resistant  
cps: capsular polysaccharide

**Table 8. Properties of Orfs in the *cps* genes of *S. suis* serotype 7 and similarities to gene products of other bacteria**

<b>Orf</b>	<b>nucleotide position in sequence</b>	<b>proposed function of gene product</b>	<b>similar gene product (% identity)</b>
Cps7E	1-719	Glycosyltransferase	<i>Streptococcus suis</i> Cps9E (99%)
Cps7F	1164-1863	Glycosyltransferase	<i>Bordetella pertussis</i> BpIG <sup>1</sup> (43%) <i>Streptococcus suis</i> Cps2E <sup>1</sup> (33%)
Cps7G	1872-3086	Biosynthesis amino sugar	<i>Bordetella pertussis</i> BpIF (48%)
Cps7H	3104-3737	Glycosyltransferase	<i>Escherichia coli</i> WbdN (35%) <i>Streptococcus suis</i> Cps2K <sup>2</sup> (31%)

<sup>1</sup> similarity refers to the C-terminal part of the gene product  
<sup>2</sup> similarity refers to the N-terminal part of the gene product

Table 9. Hybridization of serotype 7 *cps* probes with chromosomal DNA of *S. suis* serotypes

serotypes	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	1/2			
DNA probes																																						
<i>cps7E</i>	-	-	+	+	+	-	+	-	+	+	+	+	+	+	-	+	+	+	+	-	+	-	+	+	-	-	-	-	+	+	-	-	-	-	-			
<i>cps7F</i>	-	-	+	+	-	+	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-			
<i>cps7G</i>	-	-	+	+	+	-	+	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-			
<i>cps7H</i>	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-			
<i>16SrRNA</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+			

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## ABSTRACT OF THE DISCLOSURE

The invention relates to *Streptococcus suis* infection in pigs, vaccines directed against those infections and tests for diagnosing *Streptococcus suis* infections. The invention provides an isolated or recombinant nucleic acid encoding a capsular gene cluster of *Streptococcus suis* or a gene or gene fragment derived thereof. The invention further provides a nucleic acid probe or primer allowing species or serotype-specific detection of *Streptococcus suis*. The invention also provides a *Streptococcus suis* antigen and vaccine derived thereof.

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